Overview

Develop a rapid procedure for the quantitation of the N-linked glycans attached to the Fc region of recombinant human IgG from direct analysis of cell culture media after trypsin digestion.

Introduction

The glycosylation on therapeutic IgGs is known to have profound effects on various therapeutic properties, such as efficacy, serum half-life, anti-inflammatory attributes, etc.

The current methodology for monitoring glycosylation consists of a complex process involving the purification of the IgG, release/derivatization of the glycans followed by their analysis. The numerous steps introduce a variety of potential sources for experimental error. For instance, it is known that large highly sialylated glycans are more difficult to release than smaller neutral glycans causing the glycan profile to change based on the extent of the de-glycosylation procedure.

To reduce time, cost, one would like to minimize the number of sample handling procedures, which would also improve reproducibility, and decrease experimental artifacts.

We have developed a HILIC-SRM procedure that provides the N-glycosylation profile of recombinant IgGs directly from tryptic-digested culture media, and thus obviates the need for purification and glycan release/derivatization, two of the more time-consuming steps. Essentially, the sample is spiked with an internal standard consisting of an isotopically labeled IgG. The material is then reduced, alkylated, digested with trypsin, and analyzed by HILIC-SRM.

The internal standard allows for both absolute and relative quantitation across the multiple samples and reduces the experimental accuracy to <10%. To demonstrate utility our HILIC-SRM approach, we have performed a time course experiment to evaluate how glycosylation changes over the course of an expression and compared glycosylation profiles obtained from IgGs expressed under different conditions.

Experimental

A recombinant human IgG was overexpressed in CHO cells using standard expression conditions. Aliquots of cell culture media were collected, spiked with an internal standard consisting of an IgG whose glycans are isotopically labeled. Proteins/glycoproteins in the sample are reduced with diethiothreitol, alkylated with iodoacetamide, and then digested with trypsin. The resulting peptide/glycopeptide mixture is analyzed by HILIC-MS using a Hilo-penta-HILIC column at 0.6mL/min and a 65-57% ACN in 0.1% formic acid and water gradient for 30 min and an ABSCIEX Q-Trap 4000 utilizing multiple reaction monitoring. The relative ratios of the glycans are obtained by SRM detection of the analyte glycoforms, and the isotopically labeled glycoforms were used as an internal standard for relative and absolute quantitation.

Use of an IgG with Isotopically Labeled Glycans as an Internal Standard

The use of internal standards is the accepted strategy to facilitate quantitation via MS. The closer the chemical properties of the internal standard to its analyte, the better it compensates for the various sources of error, and thus the most desirable internal standard is typically an isotopically labeled version of the analyte itself. Consequently, the optimal internal standard for glycoproteins is a labeled glycoprotein, which led GlycoScientific to develop a monoclonal antibody with 15N labeled glycans (iGlycoMAb). This enables the addition of the internal standard directly into the sample prior to processing and thus overcomes systematic errors associated with parallel sample handling. And as demonstrated here, iGlycoMAb can be used to provide both the relative and absolute quantitation of an intact IgG and the Fc glycoforms.

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IgG Expression Levels

HILIC-SRM and ELISA were both utilized to measure the concentration of rh-IgG in the culture broth using two different culture media. These studies show that medium A provides a higher concentration of the rh-IgG and that this is reached by Day 8. However, the cells have a fairly short stationary phase and have entered the decline phase by Day 10. Alternatively, Medium B shows an extended lag phase, which leads to a significantly lower concentration of the desired product. This study also demonstrates the utility of the isotopically labeled standard for the accurate determination of the absolute quantity of intact antibody, and each of the glycoforms.

Characterization of the iGlycoMAb Glycans

The benefit of using an isotopically labeled molecule as an internal standard led us to develop, GlycoMAb, a IgG whose glycans are labeled with >95% 15N, as seen in the mass spectrum below. iGlycoMAb has the typical N-linked IgG glycan, A2F, A3G1F, A2G2F, etc, as shown by the LC-MS analysis of the released glycans, to the left.

Improved Accuracy with iGlycoMAb

A serial dilution of the purified rh-IgG into culture media was performed to evaluate the accuracy of the approach. The result demonstrate a negative deviation from ideality when the concentration of rh-IgG increases, which can potentially be explained by incomplete PNGase F release, overloading of the protein G column, although several other causes are also plausible. The similarity of GlycoMAb to the rh-IgG allows this internal standard to overcome these sources of systematic error and significantly improves the accuracy of the assay.

Improved Precision with iGlycoMAb

The use of GlycoMAb significantly reduces the %CV compared to approaches without internal standards. This can be seen by the decreased error bars obtained when determining the glycan prevalence with the SRM glycopeptide approach. The internal standards also facilitates comparing data between different instruments and across different laboratories.

Conclusions:

A general LC-SRM approach has been created to determine the level of 36 different glycans attached to the Fc region of an IgG. These glycans represent every glycan previously found on a human IgG. The analysis saves time and money because it is performed directly on tryptic digested cell media, without the need for IgG purification or glycan release. The reduction in steps also reduces experimental error. The inclusion of an isotopically labeled IgG (iGlycoMAb) allows results obtained on the glycopeptides to be comparable to that obtained from released glycans. iGlycoMAb facilitates both the absolute and relative quantitation of the intact IgG and each of the glycoforms.

The isotopic standard improves both the accuracy and precision because it overcomes various sources of experimental error. iGlycoMAb can be used to any glycan/glycomics work-up/analysis.