

# Optimising Sample Throughput in Bioanalytical Workflows

Matt James<sup>1</sup>, Tony Edge<sup>1</sup>, Geoff Faden<sup>2</sup>

<sup>1</sup> Avantor, Theale, Reading, Berkshire RG7 4PE, UK, <sup>2</sup> MAC-MOD Analytical Inc., 103 Commons Court, PO Box 587, Chadds Ford, PA 19317 USA

## 1. Background

- Sensitive and selective LC-MS analytical protocols benefit from sample preparation.
- For biological samples, different approaches including Protein precipitation (PPP), SLE and SPE can be utilised.
- Differing approaches remove different amounts of matrix components.
- Differing samples will have different matrix components resulting in a variable effect on the detector response.
- Effective removal of matrix components results in robust assays.

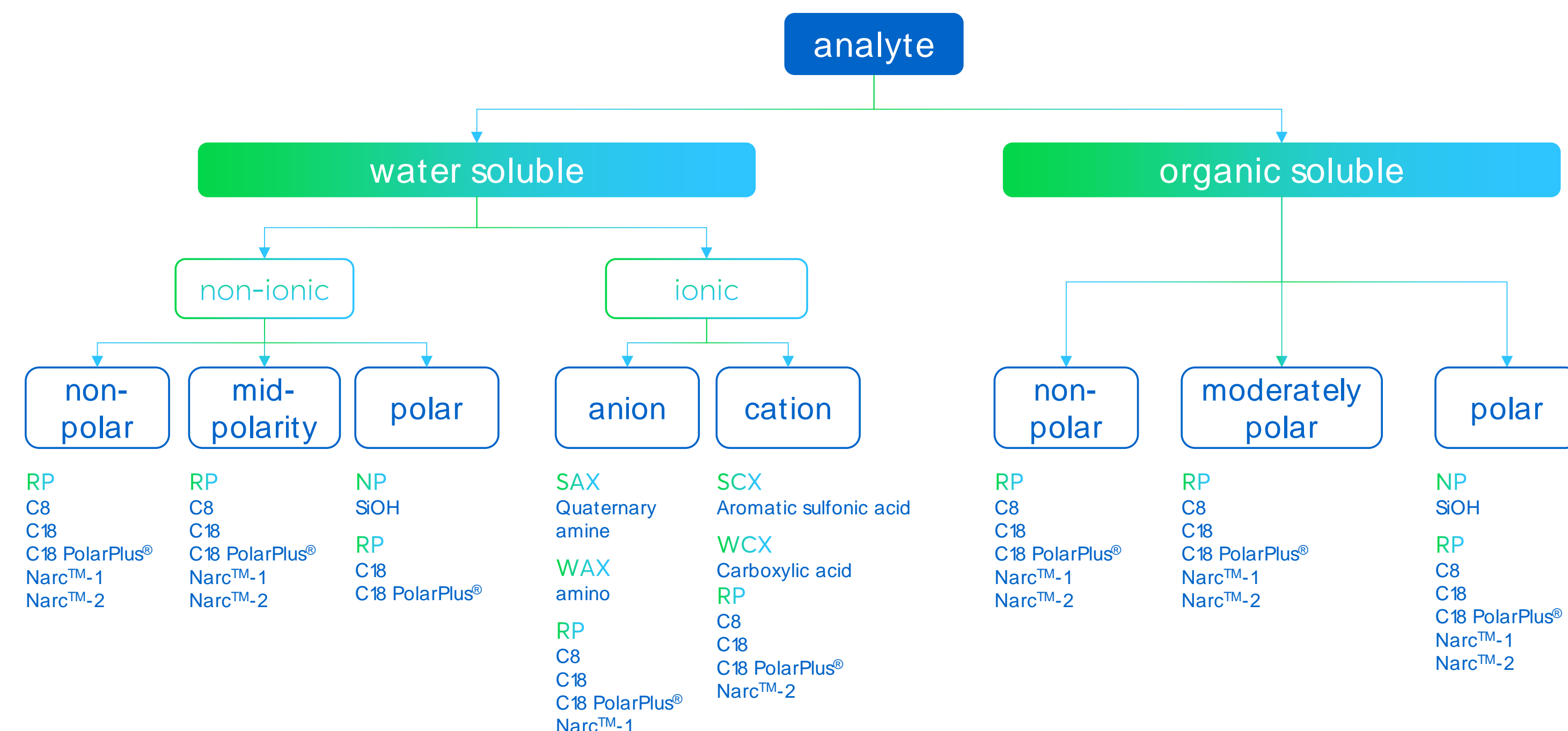
## 2. Which sample prep approach?

- SLE, PPP & SPE all have their advantages:

	SLE	PPP	SPE
Selectivity	😊😊😊😊	😊	😊😊😊😊😊😊
Cost	😊😊	😊😊😊😊😊😊	😊
Time	😊	😊😊😊😊😊😊	😊
Method development time	😊😊	😊😊😊😊😊😊	😊😊
Cleanliness	😊😊😊😊	😊	😊😊😊😊😊😊
Range of applicable analytes	😊😊	😊😊😊😊😊😊	😊😊😊😊😊😊

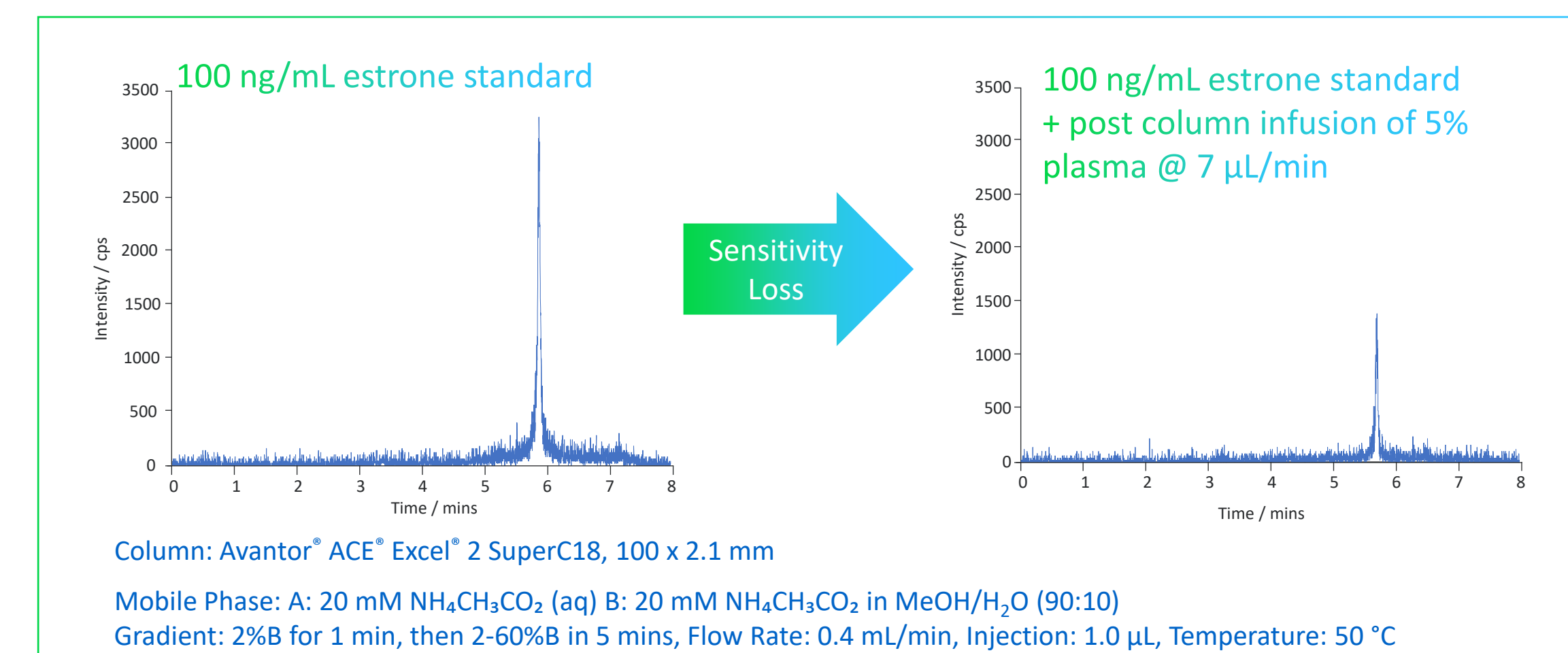


- Selection of an appropriate SPE phase, base on analyte physico-chemical properties:

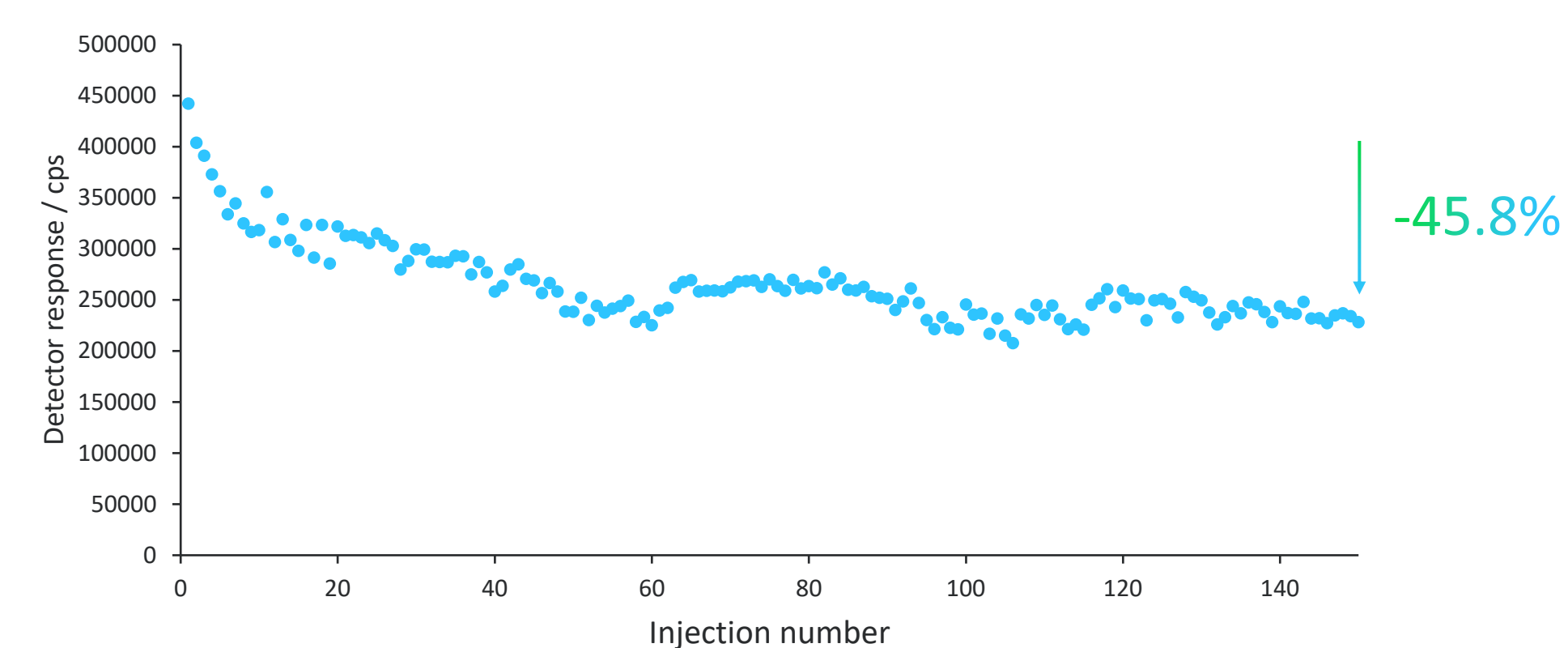


## 3. Investigating sources of signal suppression

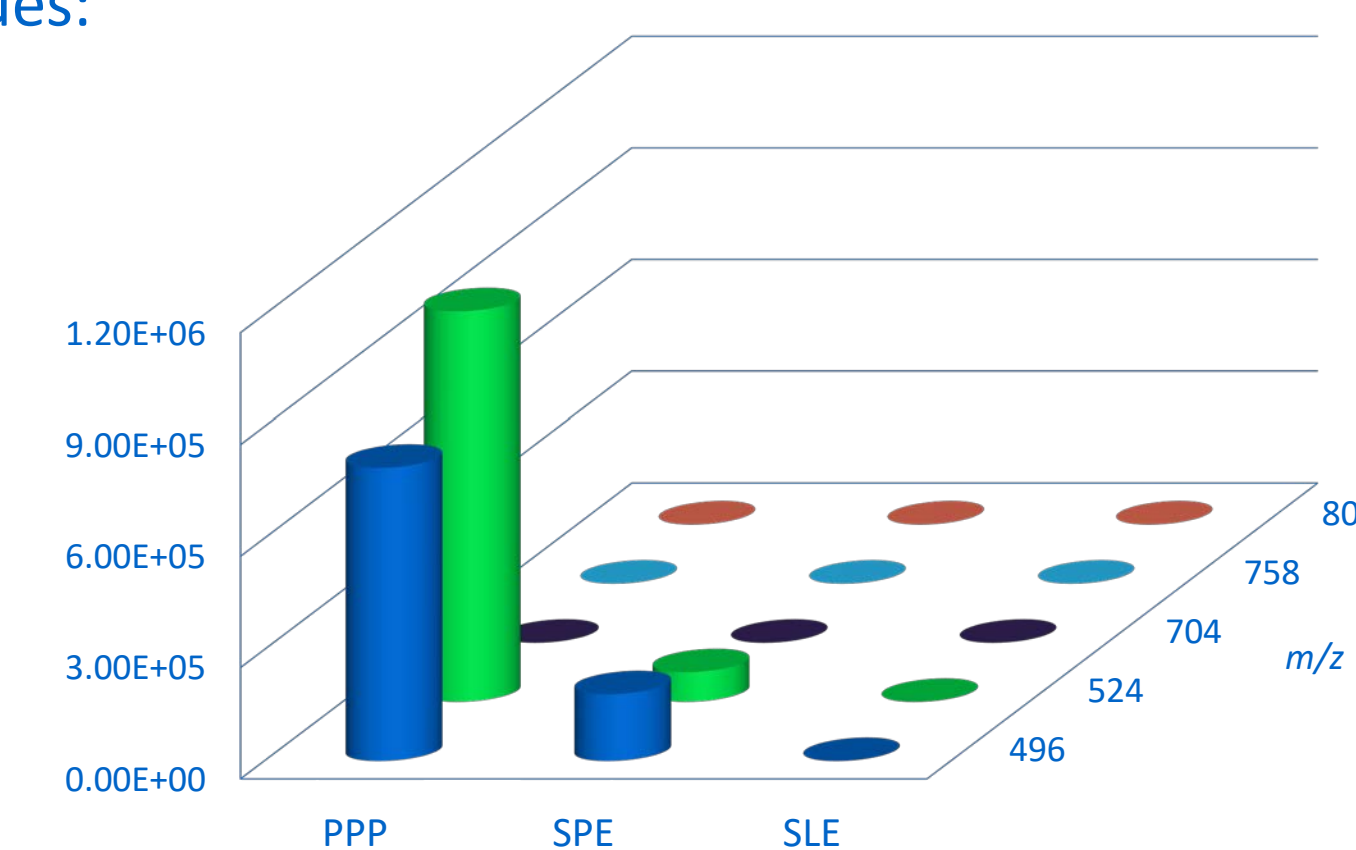
- Matrix components can impact analyte response (matrix effects):
  - Suppression
  - Enhancement
- Demonstration of the influence of matrix effects on the analysis of estrone (MRM 269.1→ 145.0) by LC-MS.
- Not removing the matrix reduces the signal.
- Magnitude of reduction depends on amount of matrix component eluting, which can be highly variable.



- Impact of 150 repeat injections (1 µL) of a 1:20 dilution of protein precipitated dog plasma on MS response:



- Removal of phospholipid components by different sample preparation techniques:



## 4. Chromatographic separation & calibration data

- Optimisation of methodology by careful selection of approach, sorbents, load, wash and elution solvents, pH and chromatographic conditions will produce a method that is validatable in accordance with the regulatory guidelines, in this case for Fluticasone.

### Sample Preparation

2 µL fluticasone-d5 internal standard was added to 398 µL rat plasma and diluted with 4 volumes of water. The following SPE protocol applied.

SPE Plate: J.T.Baker®, BAKERBOND™ spe C18, 60 mg, 96 well plate  
 Conditioning: 400 µL of MeCN.  
 Equilibration: 400 µL of H<sub>2</sub>O.  
 Sample Load: Load 25 µL of pre-treated sample.  
 Wash: Wash with 200 µL water  
 Elution: Elute analytes with 200 µL of DCM.  
 Post Elution: Evaporate to dryness & reconstitute in start mobile phase.  
 Analysis: Eluent analysed by LC-MS/MS

### Chromatography

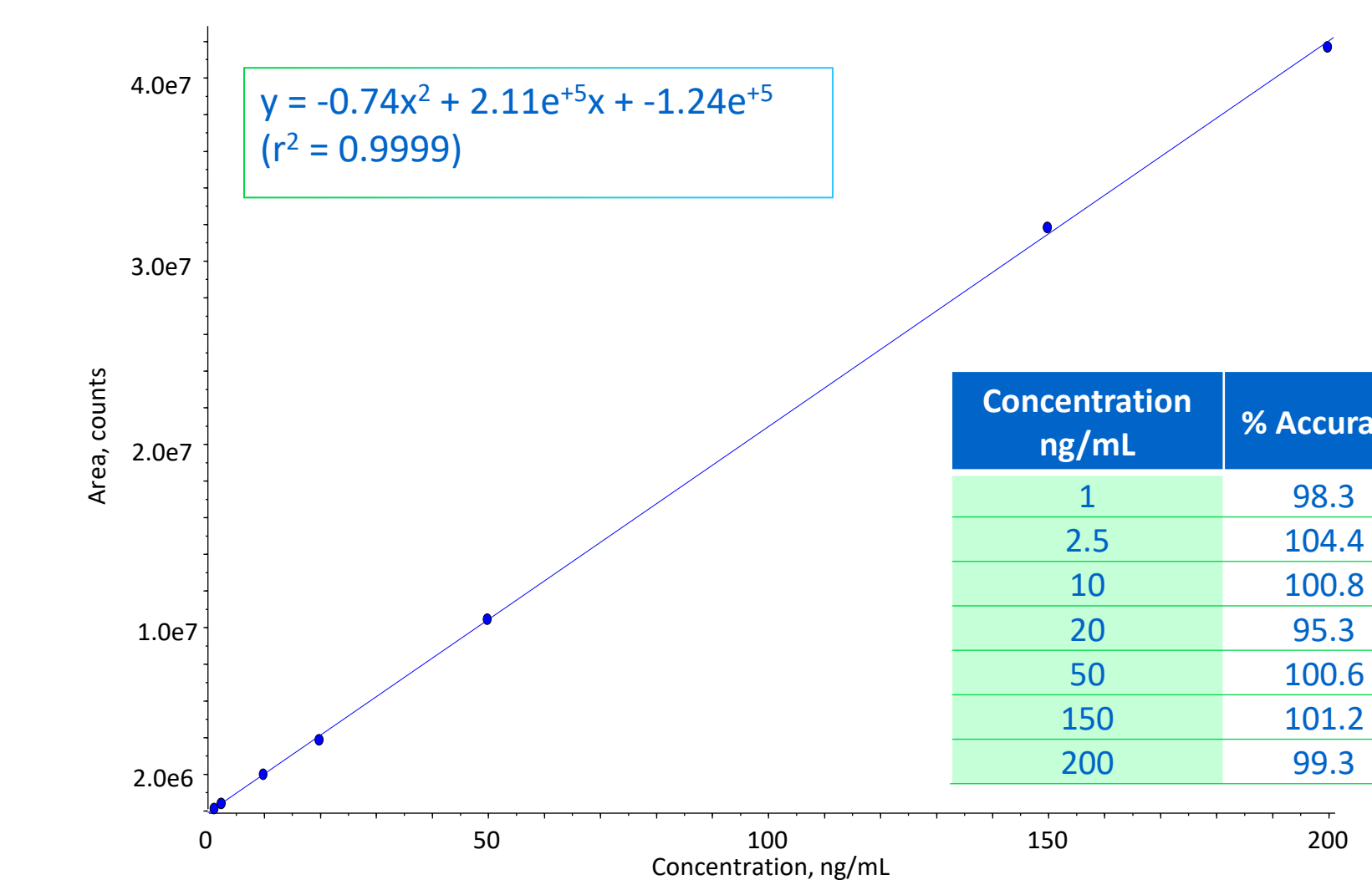
Column: Avantor® ACE® Excel™ 2 SuperC18  
 Particle Size: 2 µm  
 Dimensions: 50 x 2.1 mm  
 Mobile Phase: A: 0.1% formic acid (aq)  
 B: 0.1% formic acid in MeOH

Gradient:

Time (mins)	% B
0	45
0.1	45
2.0	95
5.0	95
5.1	45

Flow Rate: 0.4 mL/min  
 Injection: 5 µL  
 Column temp.: 45 °C  
 Detection: Sciex QTRAP® 6500+ LC-MS/MS system.

### Calibration



- Data obtained from the SPE-LC-MS/MS assay is excellent.
- Demonstrates importance of optimising the sample preparation and chromatographic conditions to remove adverse matrix effects.

## 5. Summary and Conclusions

- Different approaches to sample preparation can be employed, with different advantages and disadvantages to each approach.
- Ion suppression can have dramatic impact on assay stability and also the detection limits.
- Matrix components are removed quantitatively different using different approaches.
- Excellent linearity, accuracy and precision demonstrated for an optimised SPE-LC/MS/MS assay.
- Rapid LC-MS/MS separation developed on an Avantor® ACE® Excel™ 2 SuperC18.