

Practical UHPLC:

Selectivity and Rapid Method Development,
Method Translations
& Instrument Transfers

Gemma Lo



Contents

- ◆ How can UHPLC help us?
- ◆ Ultra resolution separations
- ◆ UHPLC practical hints and tips
- ◆ Maximising selectivity for UHPLC method development
- ◆ Translating methods to UHPLC
- ◆ Transferring UHPLC methods between instruments

How Can UHPLC Help Us?

- ◆ UHPLC with small particles (sub-2 micron) provide higher performance than HPLC (**but with higher backpressure**)
- ◆ Column efficiency is inversely proportional to particle size (d_p)

$$N \propto \frac{1}{d_p}$$

- ◆ Therefore, decreasing the particle size by 3x increases efficiency by 3x e.g. by changing from 5 μm to 1.7 μm
- ◆ We can also choose to reduce column length by 1/3 to achieve around the same efficiency ($N \propto L$)...this means run time will decrease by a factor of 3 (at constant mobile phase linear velocity).

150 mm, 5 μm	\approx	50 mm, 1.7 μm
15 mins.	\rightarrow	5 mins

How Can UHPLC Help Us?

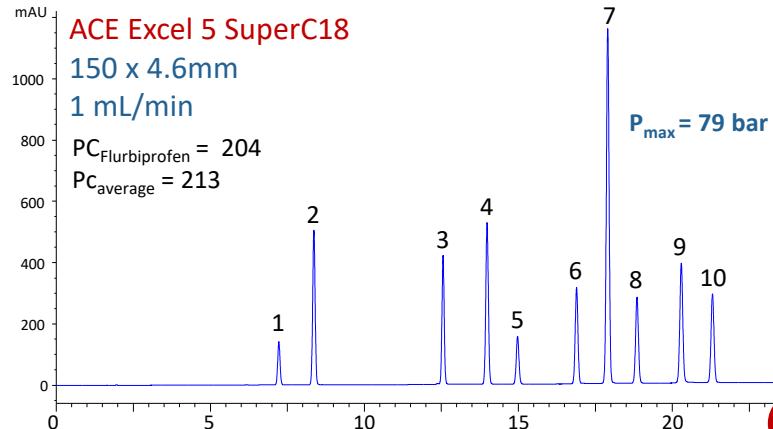
- ◆ UHPLC offers several key potential benefits over HPLC:
 - 1. Speed of analysis
 - 2. Higher resolution
 - 3. ‘Ultra-high’ resolution separations (long columns)
 - 4. Increased Productivity

1 & 2. Speed and Higher resolution

♦ Speed / Resolution

$$P_x = 1 + \frac{t_G}{W_x}$$

$$P_{Average} = 1 + \frac{t_G}{\frac{1}{10} \sum_1^{10} W_p}$$

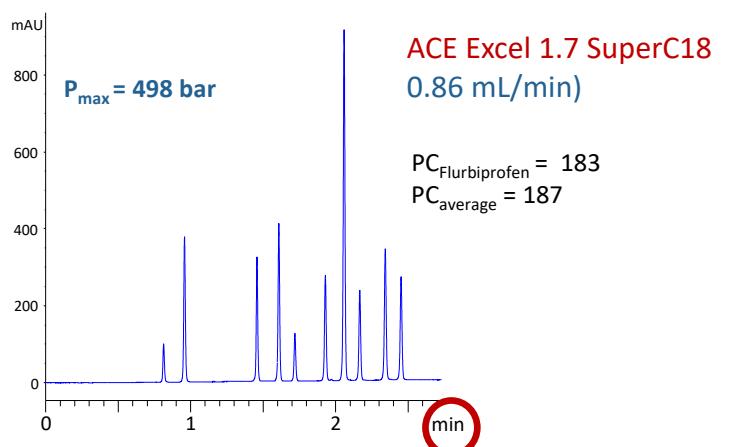


Resolution Values

Peak Pair	150 x 4.6	50 x 3.0	100 x 3.0
1,2	7.8	7.5	11.1
2,3	29.6	26.6	37.4
3,4	9.8	7.9	9.9
4,5	6.2	5.4	6.9
5,6	11.7	10.0	12.7
6,7	6.1	6.0	8.7
7,8	5.6	5.1	6.6
8,9	8.2	8.0	10.3
9,10	5.7	4.9	7.3

Speed

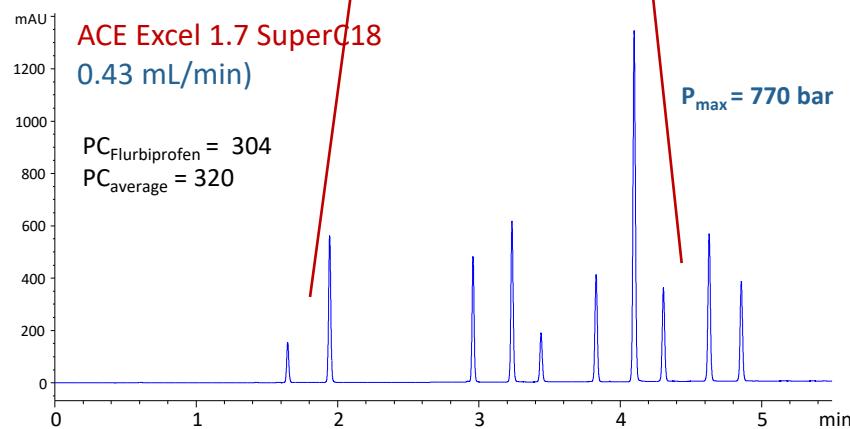
- >> speed
 - ≈ resolution
- 50 x 3.0mm



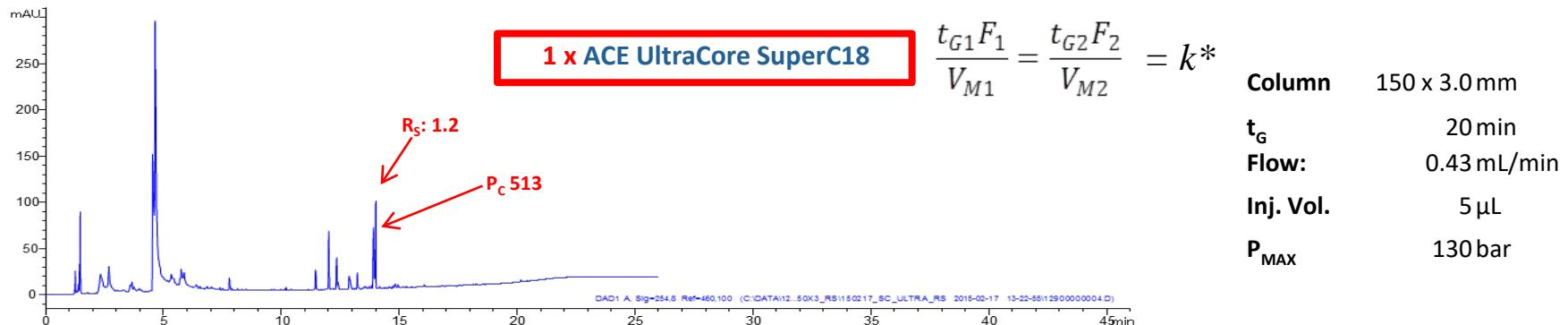
Resolution

- > speed
- >> resolution

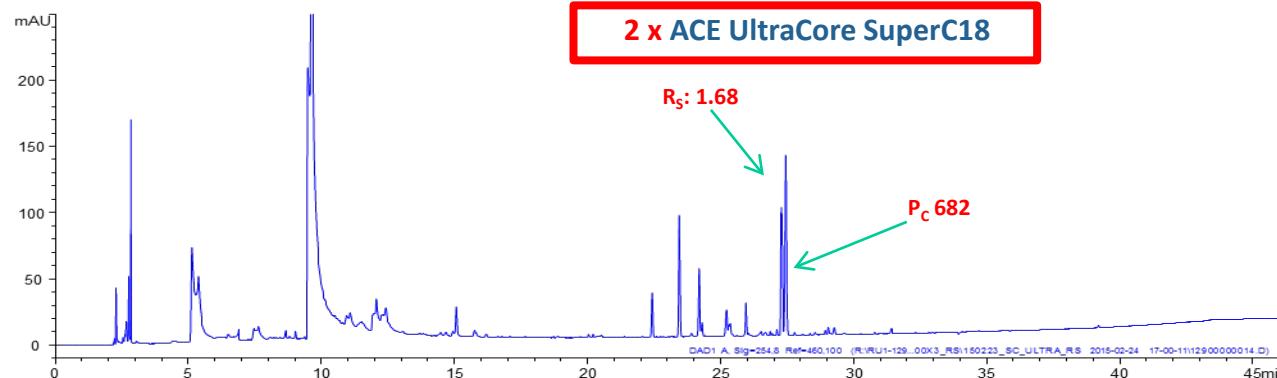
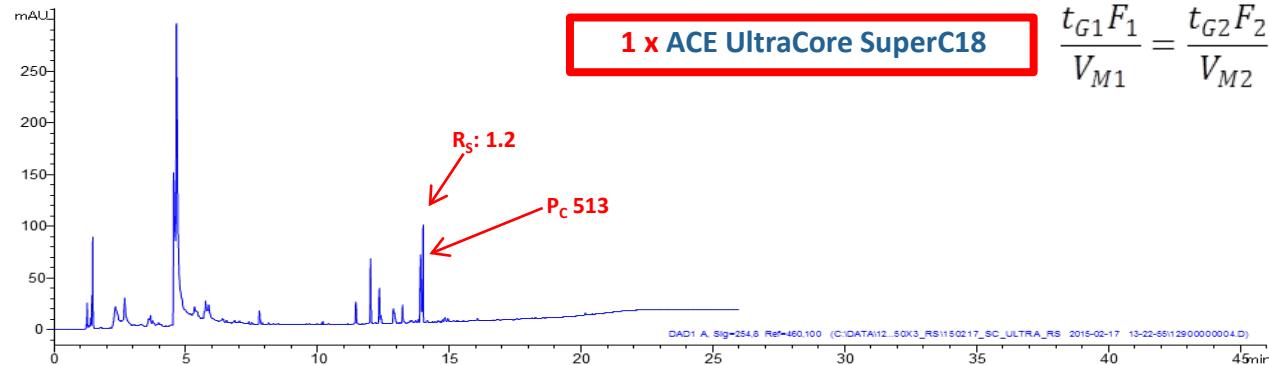
100 x 3.0mm



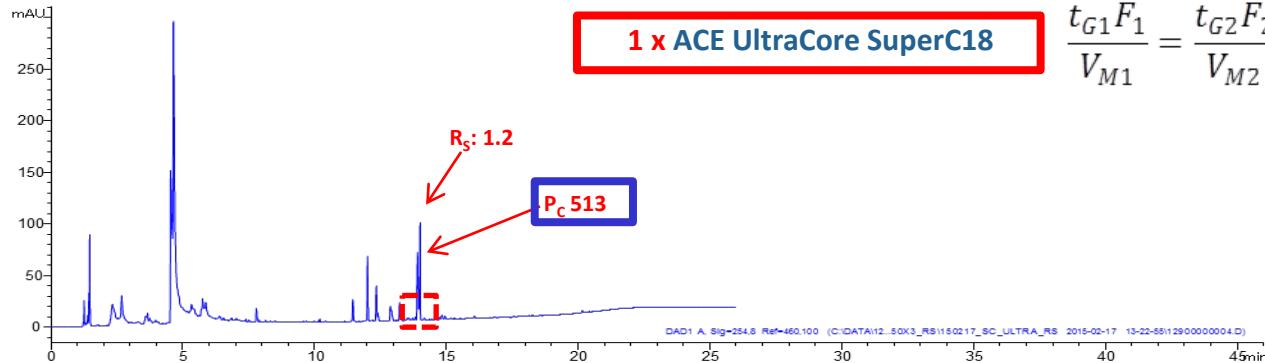
3. Ultra-High Resolution: Echinacea; Longer Column



3. Ultra-High Resolution: Echinacea; Longer Column

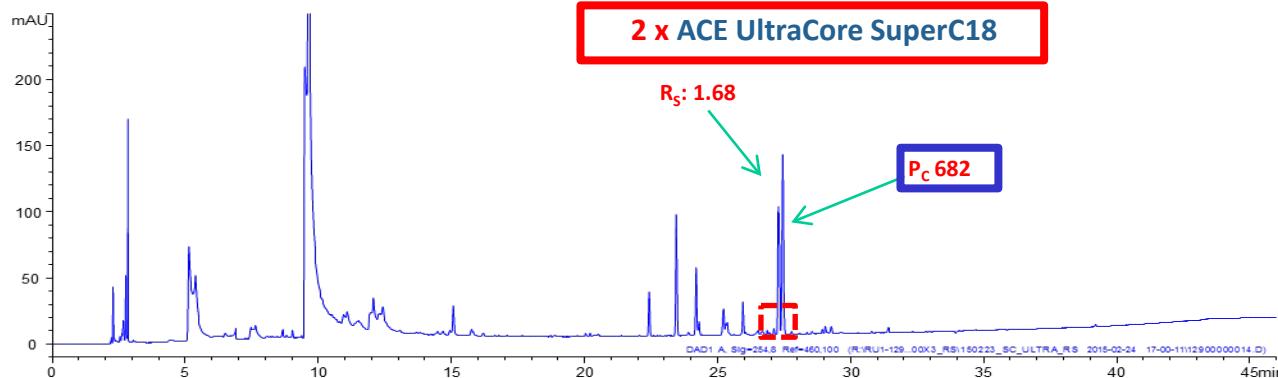


3. Ultra-High Resolution: Echinacea; Longer Column

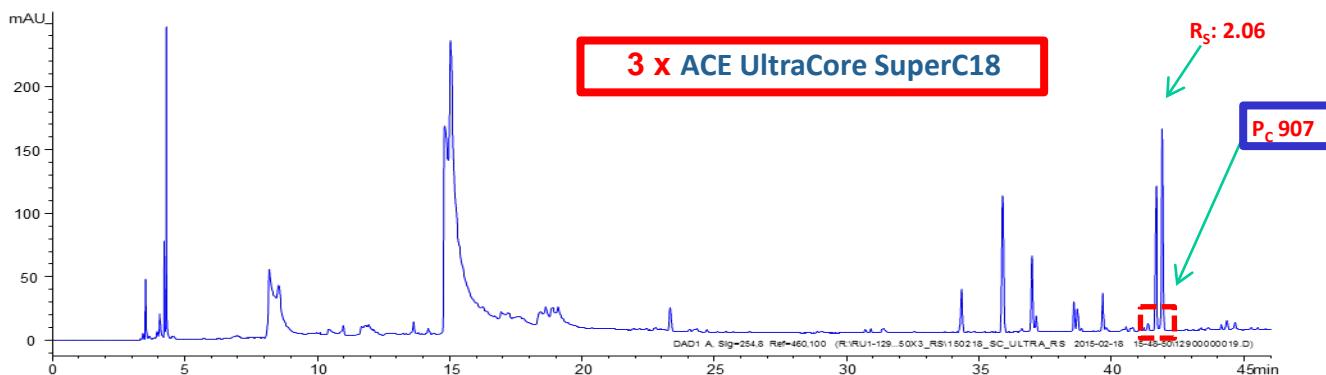


$$\frac{t_{G1}F_1}{V_{M1}} = \frac{t_{G2}F_2}{V_{M2}} = k^*$$

Column	150 x 3.0 mm
t_G	20 min
Flow:	0.43 mL/min
Inj. Vol.	5 µL
PMAX	130 bar

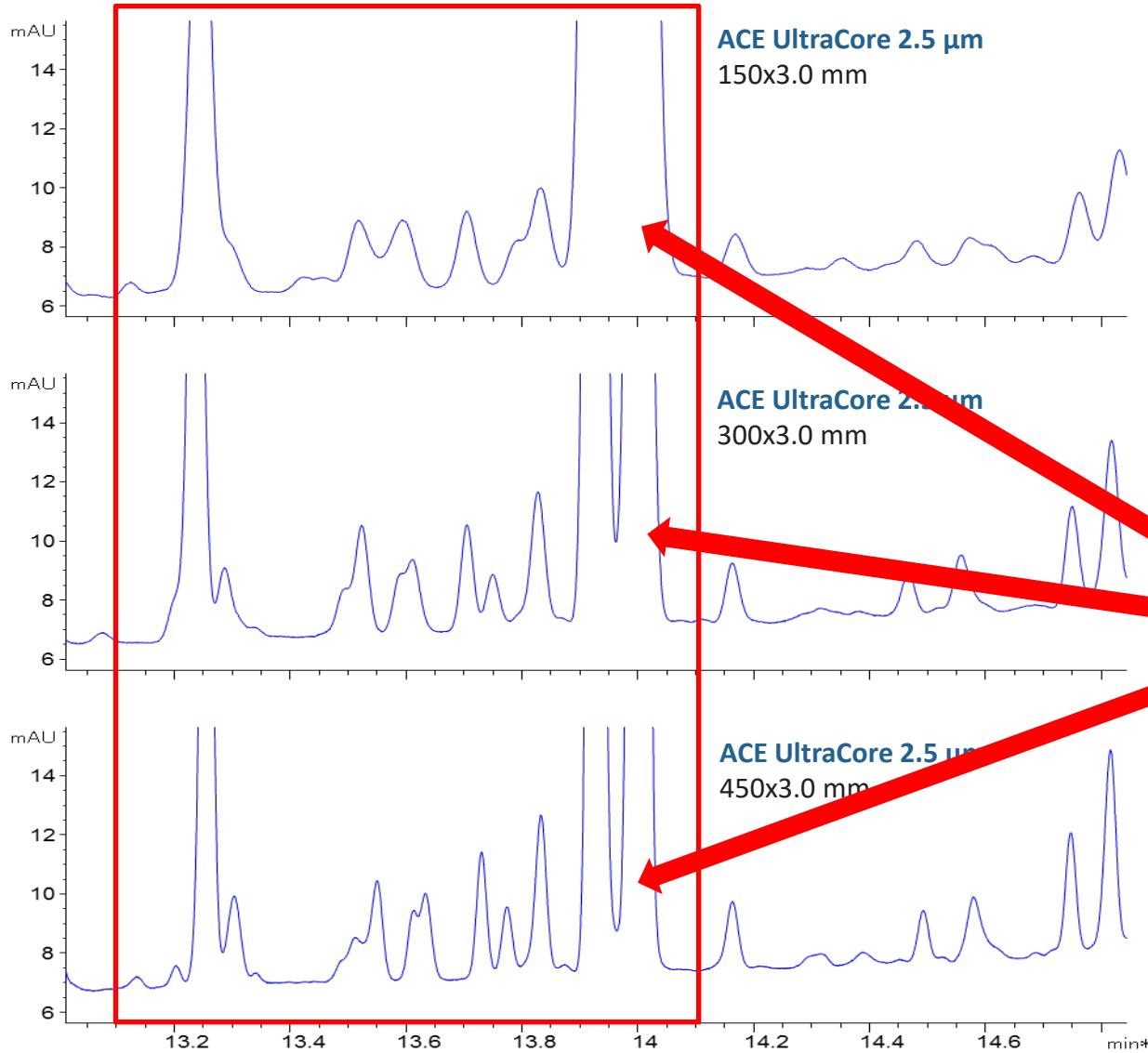


Column	300 x 3.0 mm
t_G	40 min
Flow:	0.43 mL/min
Inj. Vol.	10 µL
PMAX	223 bar



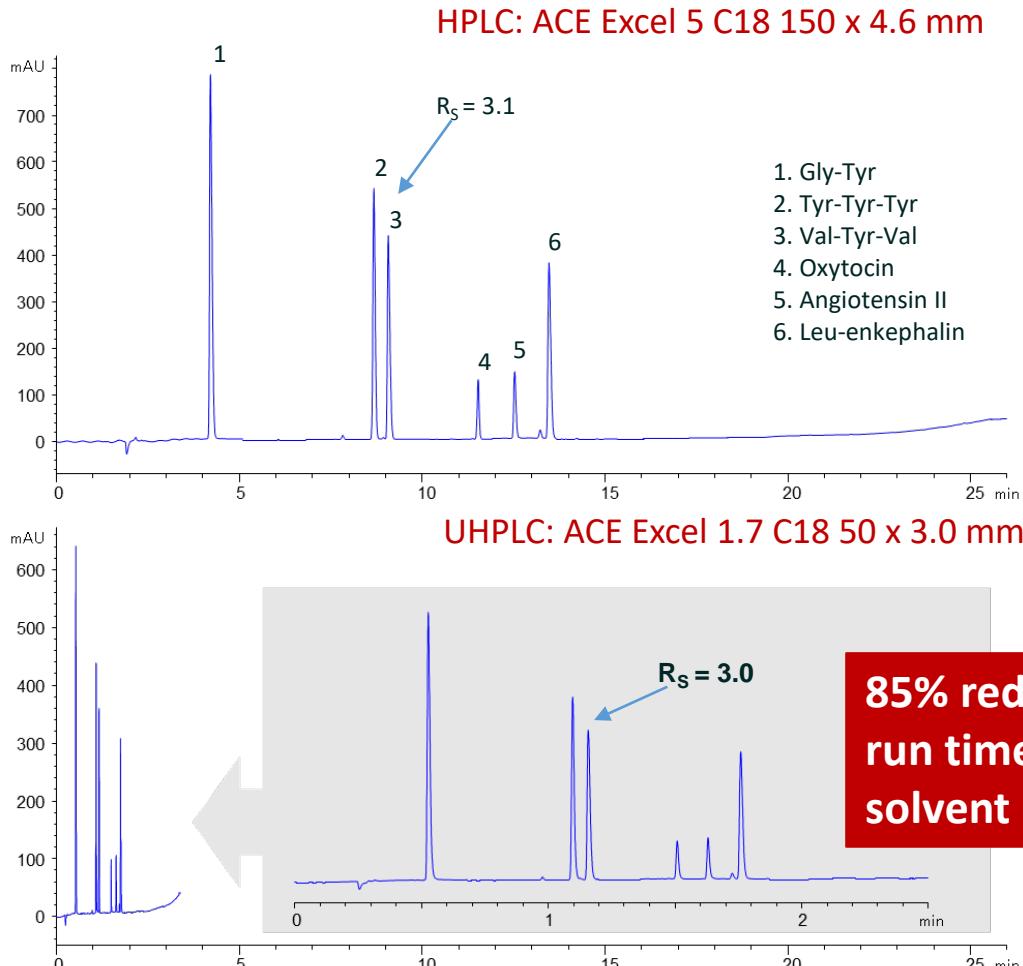
Column	450 x 3.0 mm
t_G	59.96 min
Flow:	0.43 mL/min
Inj. Vol.	15 µL
PMAX	338 bar

3. Ultra-High Resolution: Echinacea; Aligned Data (ZOOM IN)





4. Increased Productivity



- ◆ Improve productivity by translating existing HPLC methods to UHPLC.
- ◆ The ACE LC Translator spreadsheet is available as a free download to help achieve method translation.

Download free at:
MAC-MOD.com

Mobile Phase: A: 0.05% TFA in H₂O, B: 0.05% TFA in MeCN, Temperature: 60 °C, Detection: UV, 220 nm
HPLC conditions: 1.00 mL/min, 4-40 %B in 17 minutes, then 40-90 %B in 6 minutes
UHPLC conditions: 1.00 mL/min, 4-40 %B in 2.41 minutes, then 40-90 %B in 0.85 minutes

UHPLC Practical Hints & Tips

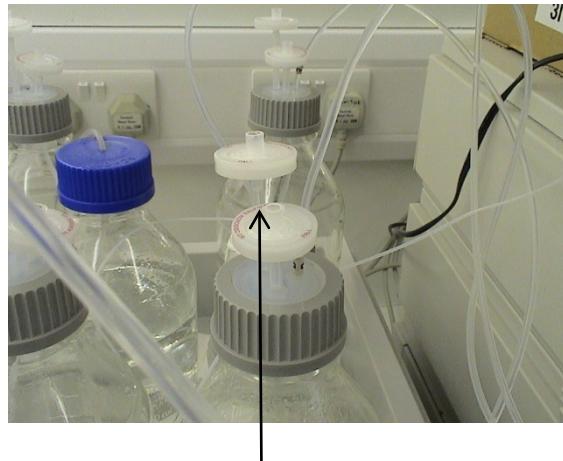
UHPLC Hints & Tips: Filtering Mobile phase



- ◆ General good practice: replace mobile phases regularly to prevent bacterial growth.
- ◆ Filter mobile phases through a 0.2 µm membrane filter prior to use
 - ◆ Removes damaging microparticulates and bacterial contaminants
- ◆ Nylon filters: aqueous solvents
- ◆ PTFE (Teflon): organic solvents
- ◆ Universal filters also available
- ◆ Use high quality solvents, particularly for LC-MS.

Solvents / Buffer Solutions on Instruments

- ◆ Ensure you seal mobile phase reservoirs on the UHPLC instrument
- ◆ This prevents **dirt / dust** and **microbial** contamination



0.45 µm syringe filter fitted to solvent bottle cap



S.C.A.T. SafetyCaps on solvent bottles

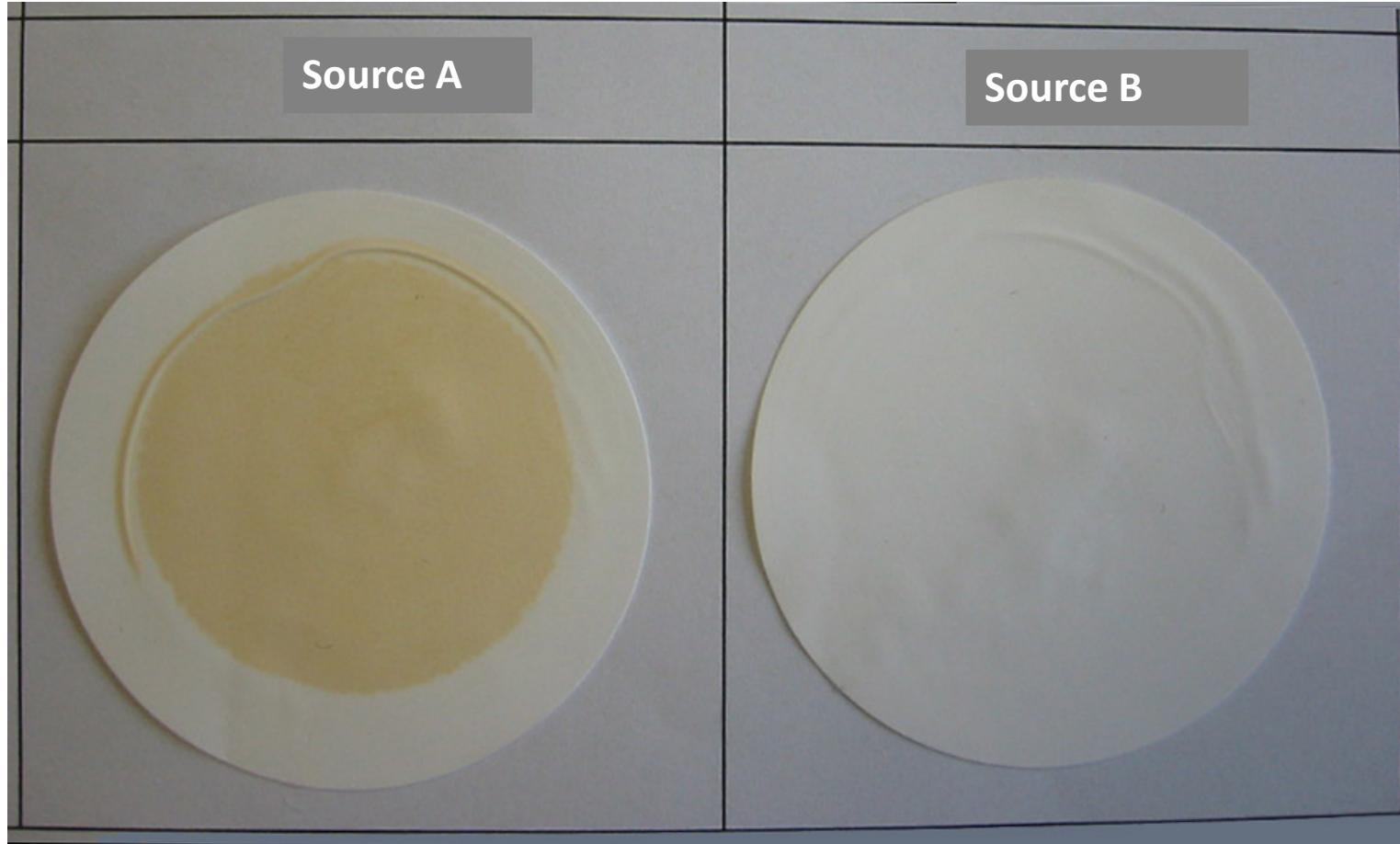


S.C.A.T. SafetyWasteCaps on waste bottles

Particularly if 100% aqueous is used on a solvent line

Not all HPLC water is good!

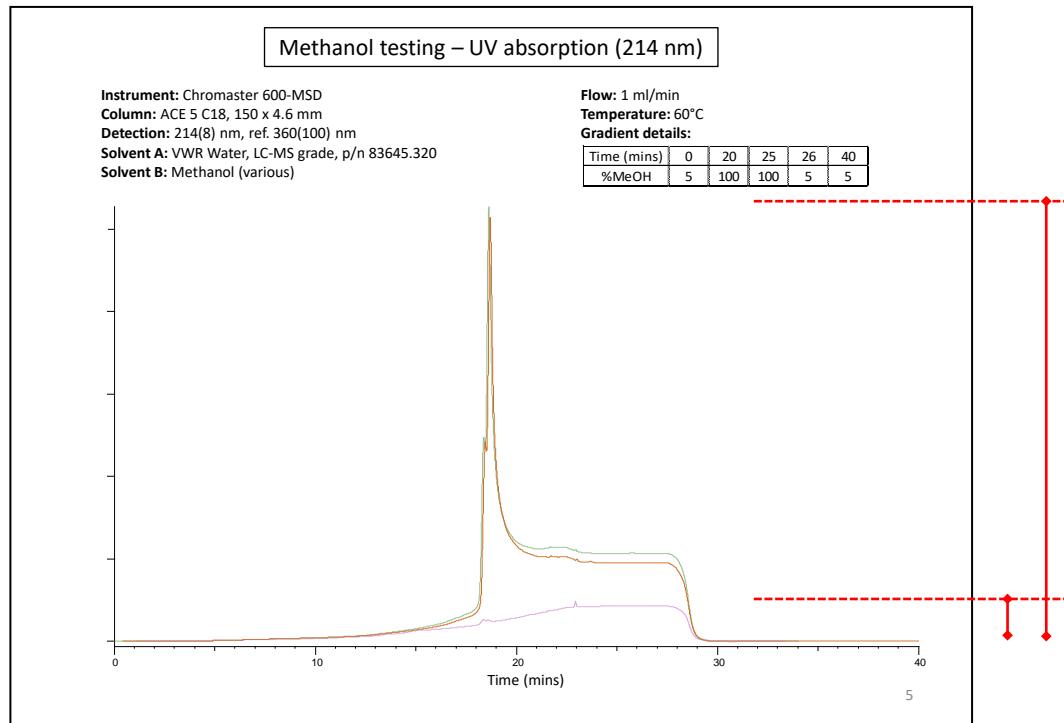
Bottled HPLC grade water!





Organic Solvents LC-UV

- Always use the highest quality organic solvents you can afford...

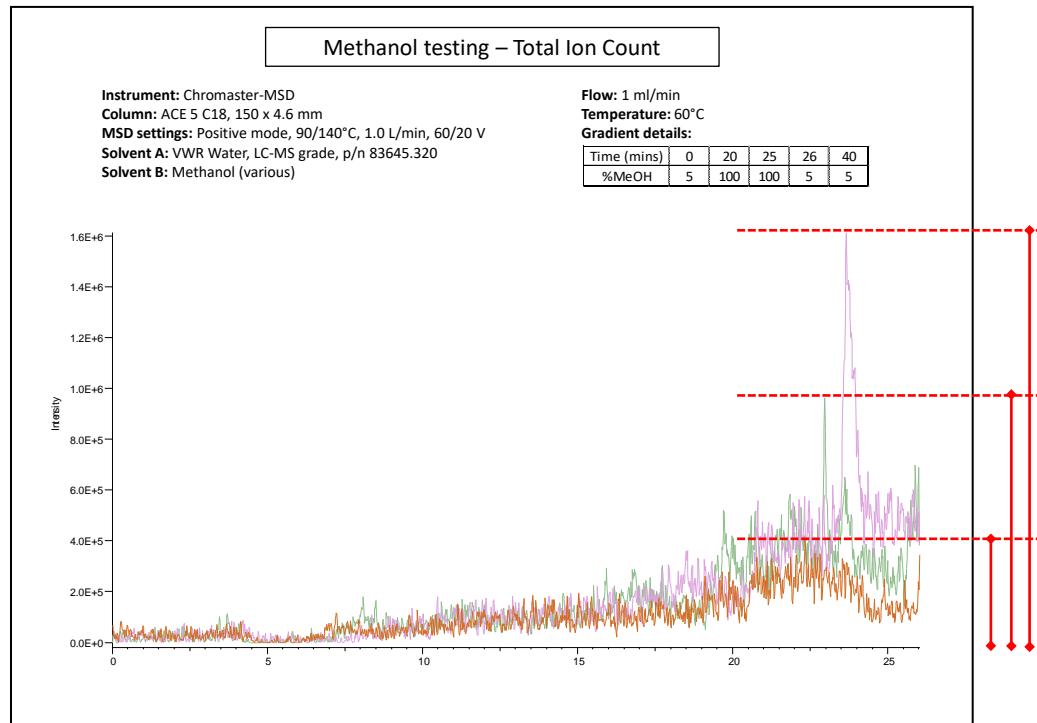


Especially for UHPLC, gradient analyses and LC-MS work



Organic Solvents LC-MS

- Always use the highest quality organic solvents you can afford...



Especially for UHPLC, gradient analyses and LC-MS work

UHPLC Method Development: Using Selectivity



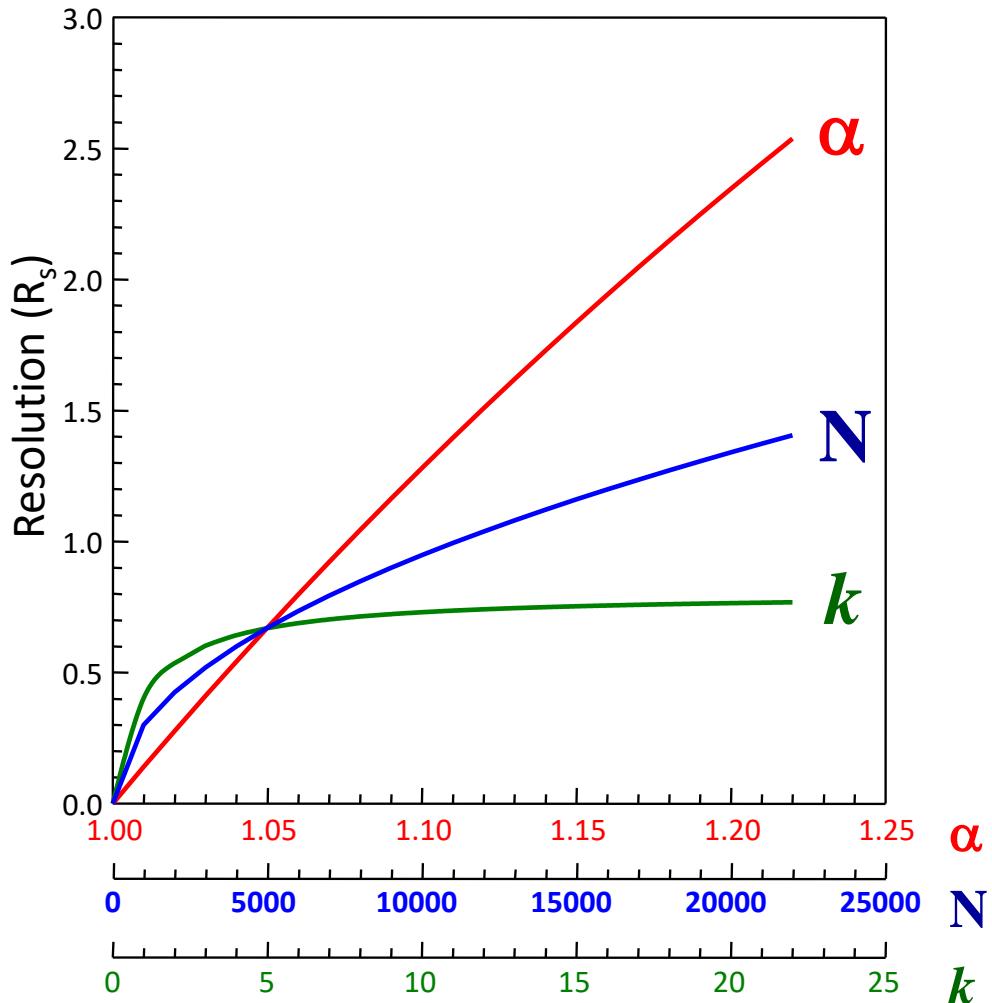
The Power of Selectivity

The Resolution Equation

Efficiency Selectivity Retention

$$R_s = \frac{\sqrt{N}}{4} \cdot \frac{\alpha-1}{\alpha} \cdot \frac{k}{1+k}$$

Selectivity is the key to analyte resolution with efficiency boosting performance.





Which Factors^a Affect Selectivity?

Isocratic Separations

- ◆ Column stationary phase
- ◆ Organic Modifier
- ◆ pH (ionised analytes only)
- ◆ % Organic modifier
- ◆ Buffer selection
- ◆ Column temperature
- ◆ Buffer concentration

MOST
Influence



Gradient Separations

- ◆ All parameters for isocratic
- ◆ Gradient steepness
- ◆ k^* (t_G , F, V_m , $\Delta\Phi$, Mw)

$$k^* = \frac{0.87t_GF}{\Delta\Phi V_m M}$$

- ◆ Dwell volume
- ◆ Column dimensions

LEAST
Influence

- ◆ Stationary phase chemistry: one of the most powerful parameters.
- ◆ ACE novel phases have been specifically designed to offer complementary selectivity – great for method development

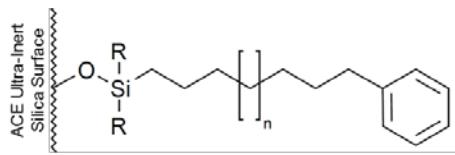


ACE Novel Reversed Phase Columns

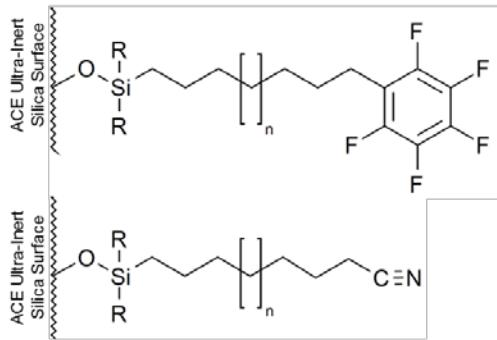
- ◆ The ACE range contains a wide range of stationary phases:
 - Traditional phases (C18, C8, C4, CN, Phenyl)
 - Novel phases (C18-AR, C18-PFP, C18-Amide, CN-ES, SuperC18)
- ◆ The **ACE novel stationary phases** have been specifically designed to provide different retention mechanisms and therefore **different selectivity.**
- ◆ These phases are a powerful method development tool and can provide separations that may not be achievable using a traditional C18.
- ◆ Available in 1.7, 2, 3, 5 and 10 µm HPLC / UHPLC format.

ACE Novel Phases

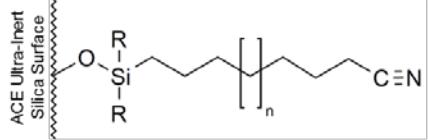
- ◆ ACE novel chemistries: C18 like retention, reproducibility and stability.



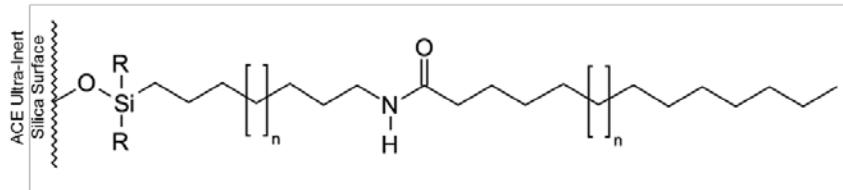
ACE C18-AR (hydrophobic and phenyl interactions)



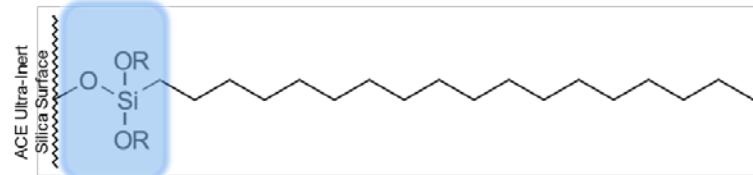
ACE C18-PFP (hydrophobic and PFP interactions)



ACE CN-ES (hydrophobic and polar interactions)



ACE C18-Amide (embedded amide group to increase retention of polar components and alternative selectivity).



ACE SuperC18 (high retentivity with extended pH stability).

ACE® Stationary Phases: Key Mechanisms of Interactions

- ◆ Tanaka¹ characterisation can help understand mechanisms and weightings with different column chemistries

Bonded Phase	Separation Mechanism and Relative Strength ¹				
	Hydrophobic Binding	π-π Interaction	Dipole-Dipole	Hydrogen Bonding	Shape Selectivity
ACE C18	****	-	-	*	**
ACE C18-AR	****	*** (donor)	*	**	***
ACE C18-PFP	****	*** (acceptor)	****	***	****
ACE SuperC18	****	-	-	-	**
ACE C18-Amide	****	-	**	****	**/***
ACE CN-ES	***	*	***	**	*

Approximate value – determined by semi-quantitative mechanism weightings and/or by reference to other ACE phases using >100 characterising analytes.

¹ K. Kimata, K. Iwaguchi, S. Onishi, K. Jinno, R. Eksteen, K. Hosoya, M. Arki, N. Tanaka, *J. Chromatogr. Sci.* 27 (1989) 721.

ACE® Stationary Phases: Key Mechanisms of Interactions

- ◆ Specifically designed phases to maximize selectivity

Bonded Phase	Separation Mechanism and Relative Strength ¹				
	Hydrophobic Binding	π-π Interaction	Dipole-Dipole	Hydrogen Bonding	Shape Selectivity
ACE C18	****	-	-	*	**
ACE C18-AR	****	*** (donor)	*	**	***
ACE C18-PFP	****	*** (acceptor)	****	***	****
ACE SuperC18	****	-	-	-	**
ACE C18-Amide	****	-	**	****	**/***
ACE CN-ES	***	*	***	**	*

Approximate value – determined by semi-quantitative mechanism weightings and/or by reference to other ACE phases using >100 characterising analytes.

- ◆ Multiple mechanisms of interaction: ideal for method development

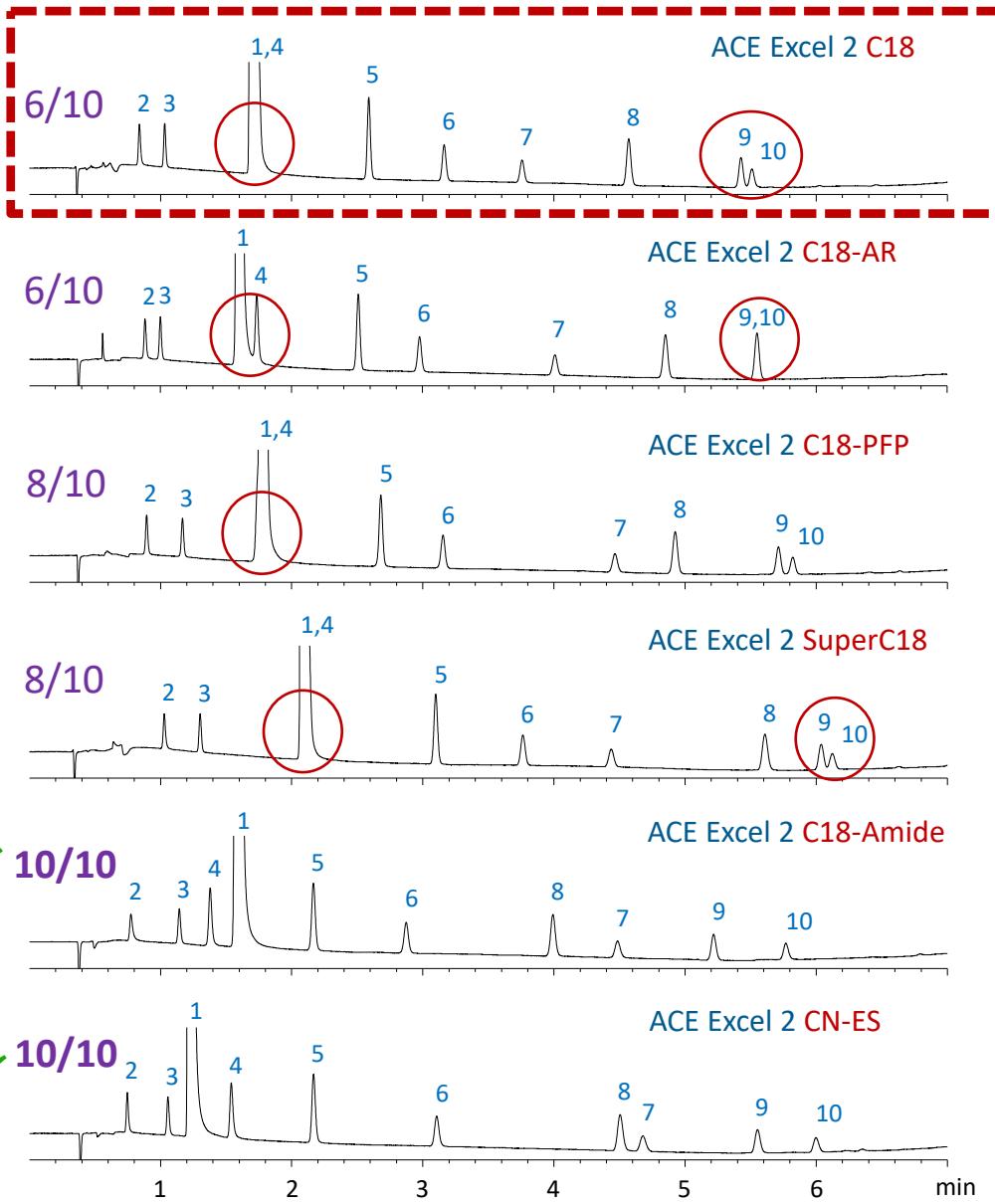
Screening combinations of column chemistries and solvents is systematic and helpful...plus can be semi-automated

The Complete Method Development Solution

MeOH

- ◆ Paracetamol and related compounds (10 analytes).
- ◆ Sample screened on six 100 x 3.0 mm ACE columns
- ◆ Methanol and MeCN used separately as the organic modifiers.

Most common starting point



General scouting conditions:

100 x 3.0 mm columns

A: 20 mM Ammonium acetate pH 6.0

B: 20 mM Ammonium acetate pH 6.0 in MeOH or MeCN:H₂O (9:1 v/v)

Gradient: 5 to 95% B in 10 mins

Temp: 40°C, 2mL injection, Flow rate: 1.2 ml/min

Sample: Acetaminophen with rel subs at 0.5% w/w

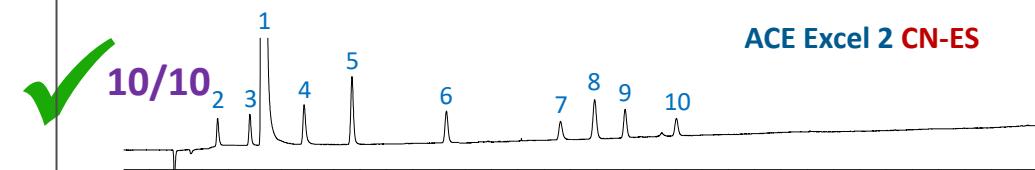
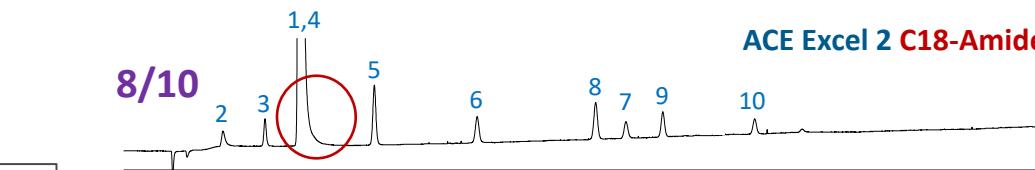
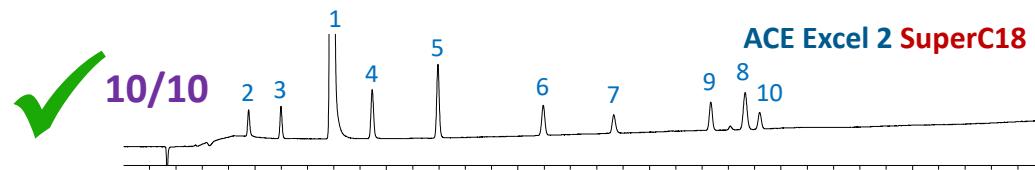
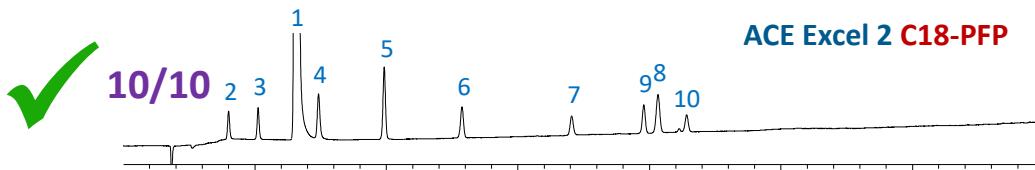
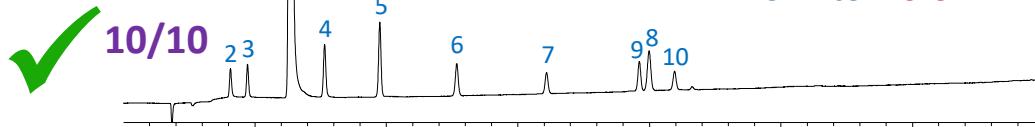
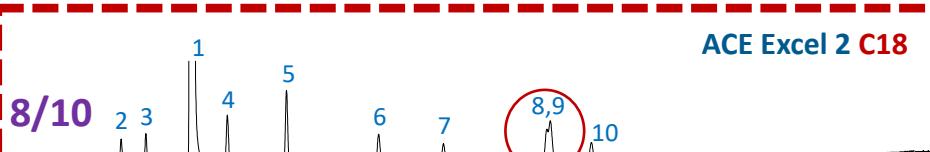
The Complete Method Development Solution

MeCN

- ◆ Paracetamol and related compounds (10 analytes).
- ◆ Sample screened on six 100 x 3.0 mm ACE columns
- ◆ Methanol and MeCN used separately as the organic modifiers.

Total of 6 options identified from the screen where all 10 analytes separated

Most common starting point



General scouting conditions:

100 x 3.0 mm columns

A: 20 mM Ammonium acetate pH 6.0

B: 20 mM Ammonium acetate pH 6.0 in MeOH or MeCN:H₂O (9:1 v/v)

Gradient: 5 to 95% B in 10 mins

Temp: 40°C, 2mL injection, Flow rate: 1.2 ml/min

Sample: Acetaminophen with rel subs at 0.5% w/w

Method Translation

HPLC to UHPLC

Isocratic Translations



Isocratic Method Translations: General Principles

- Maintain a constant length to particle size ratio, L/d_p (for the same phase type and phase vendor)
$$N = \frac{L}{HETP}$$
- Will give ~ similar performance i.e. efficiency (selectivity, resolution)*
- Thus, $300 \times 3.9 \text{ mm}, 10 \mu\text{m} \approx 150 \times 4.6 \text{ mm}, 5 \mu\text{m} = 30,000$

	Column Length (mm)					
	50	75	100	150	250	300
Particle Size (μm)	29,412	44,118	58,824			
1.7	29,412	44,118	58,824			
1.8	27,778	41,667	55,556			
1.9	26,316	39,474	52,632			
2	25,000	37,500	50,000	75,000		
2.5	20,000	30,000	40,000	60,000	100,000	
2.6	19,231	28,846	38,462	57,692	96,154	
2.7	18,519	27,778	37,037	55,556	92,593	
3	16,667	25,000	33,333	50,000	83,333	
5	10,000	15,000	20,000	30,000	50,000	
10	5,000	7,500	10,000	15,000	25,000	30,000

Columns meeting L/d_p of 30,000



Isocratic Method Translations: General Principles

- Once the new column format has been selected, isocratic translations are fairly straightforward.

- Step 1: Scale injection volume (V_i) to new column dead volume (V_M)

- Step 2: Scale flow rate (F)

$$V_{i2} = \frac{V_{i1} \times V_{M2}}{V_{M1}}$$

- If no change in d_p

$$F_2 = \frac{F_1 \times d_{c2}^2}{d_{c1}^2}$$

- If d_p changes can use

$$F_2 = \frac{F_1 \times d_{c2}^2 \times d_{p1}}{d_{c1}^2 \times d_{p2}}$$

- Step 3: Determine new run time

$$t_2 = \frac{t_1 F_1 V_{M2}}{F_2 V_{M1}}$$

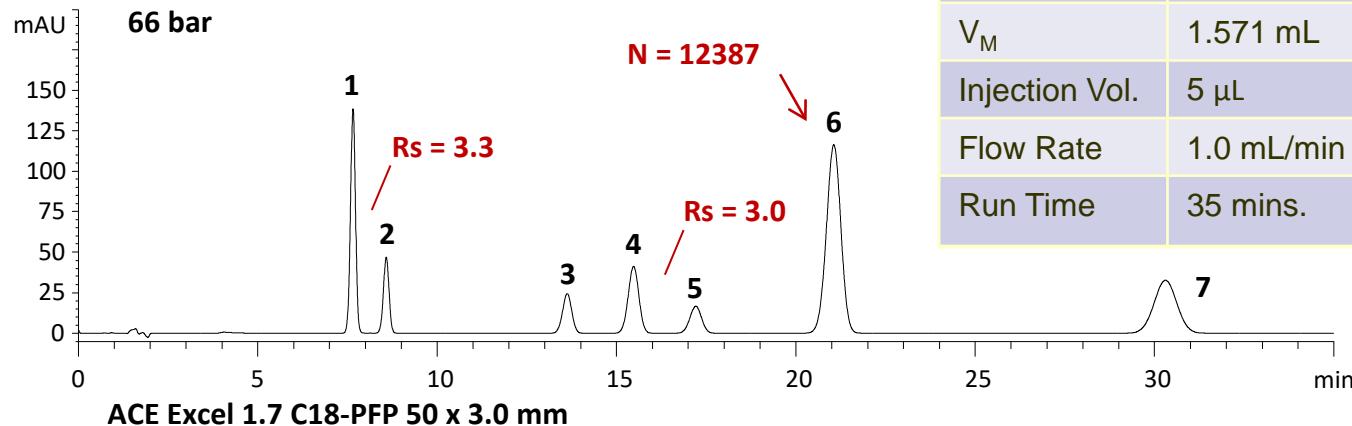
- Backpressure of new method can be estimated

$$P_2 = \frac{P_1 \times F_2 \times L_2 \times d_{c1}^2 \times d_{p1}^2}{F_1 \times L_1 \times d_{c2}^2 \times d_{p2}^2}$$

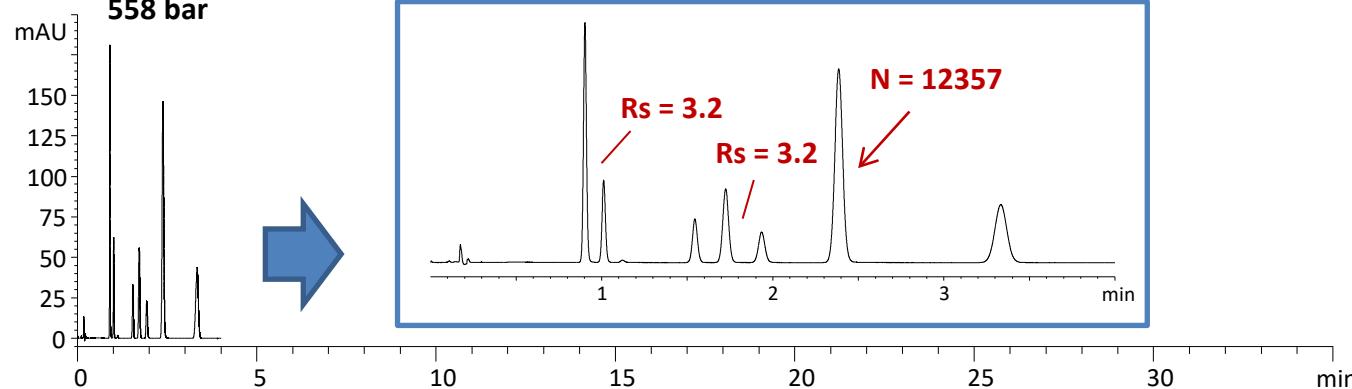


Isocratic Method Translations: HPLC to UHPLC

Methoxybenzene Structural Isomers
ACE 5 C18-PFP 150 x 4.6 mm
66 bar



ACE Excel 1.7 C18-PFP 50 x 3.0 mm



	Original method	Translated method	Difference
Run time	35 minutes	4 minutes	-88.6%
Solvent consumption	35.0 mL	5.0 mL	-85.7%

FREE ACE® Method Translation Tool: Isocratic Example

V1.3

Method Translation

Methods are frequently translated from one column format to another and among different brands and models of instruments (different system volumes, dwell volumes etc). In order to maintain chromatographic and method performance through the translation process, a number of method parameter changes are required, such as flow rate, injection volume and t_g . Separate tools for isocratic and gradient methods are included on this page. For details on how to determine system dwell volumes, please refer to the Dwell Volume tab.

Isocratic

Column Information		Translated	
Current		Translated	
Column Length (L)	150:mm	Column Length (L)	50:mm
Column i.d. (d_c)	4.6:mm	Column i.d. (d_c)	3.0:mm
Particle Diameter (d_p)	5.0: μm	Particle Diameter (d_p)	1.7: μm
L/d_p	30000	L/d_p	29412
Column Porosity	0.63:What's This?	Column Porosity	0.63:What's This?
Column Volume (V_M)	1.570 mL	Column Volume (V_M)	0.223 mL
Method		Translated	
Current		Translated	
Injection Volume	10.0: μL	Injection Volume	1.4 μL
Flow Rate	1.00:mL/min	Flow Rate (scaled to linear velocity)	0.43 mL/min
Run Time	25.0:mins	Flow Rate (scaled to particle size)	1.25 mL/min
Recorded Backpressure	60:bar	Input Flow Rate	1.25 mL/min
Solvent Use	25 mL	New Run Time	2.8 mins
		Estimated Backpressure	508 bar
		Estimated Solvent Use Difference	-86 %

Main Menu

Gradient

Fill in the grey input boxes to translate the method

Displays new run time and solvent saving

Optional input for flow rate – great for allowing greater analyst choice

****Please complete all input fields****

Download free at
MAC-MOD.com

Gradient Translations



Translations of Gradient Methods – The Basics

1. Calculate column volumes

- Better to experimentally determine porosity for accuracy

$$V_M \approx \pi \left(\frac{d}{2}\right)^2 L \varepsilon$$

2. Translate injection volume

- To give similar response

$$Inj_2 = Inj_1 \times \left(\frac{V_{m2}}{V_{m1}} \right)$$

3. Translate flow rate

- Constant linear velocity

$$F_2 = F_1 \times \frac{d_{c_2}^2}{d_{c_1}^2}$$

- Scaled to new d_p

$$F_2 = \frac{F_1 \times d_{c2}^2 \times d_{p1}}{d_{c1}^2 \times d_{p2}}$$

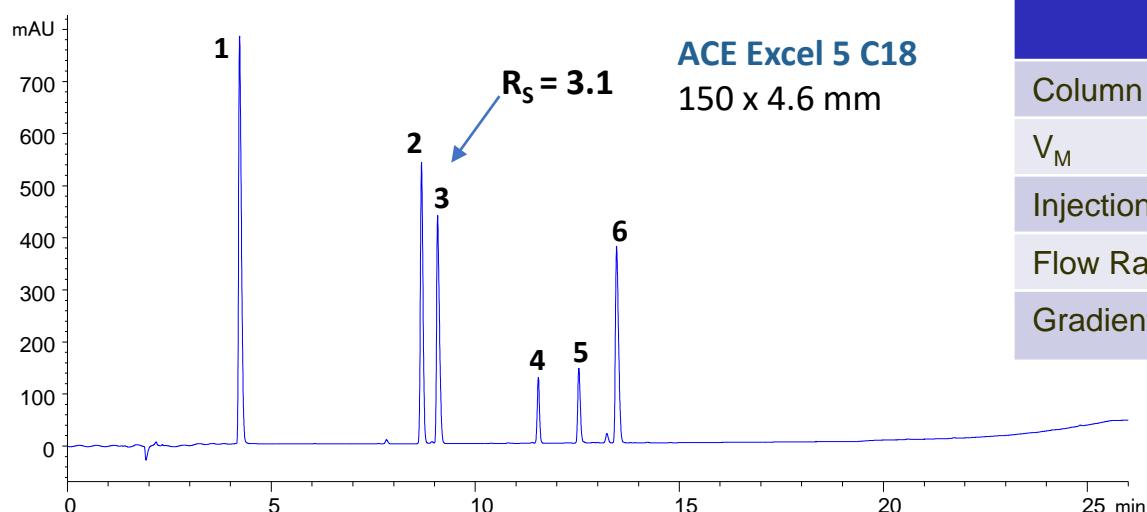
4. Translate gradient time

- To maintain constant k^*

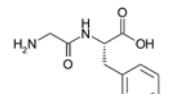
$$t_{G2} = \frac{t_{G1} V_{M2} F_1}{V_{M1} F_2}$$

5. Consider V_D/V_M ratio

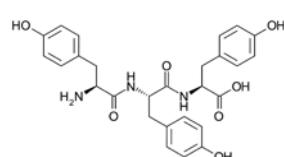
UHPLC Gradient Method Translations: Peptides



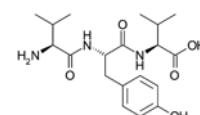
	Original Method
Column	150 x 4.6 mm, 5 μ m
V_M	1.571 mL
Injection Vol.	10 μ L
Flow Rate	1.0 mL/min
Gradient time	17 min.



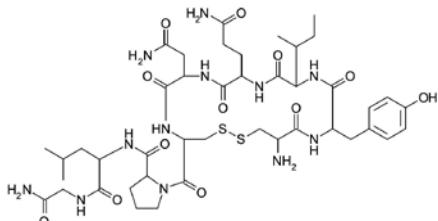
1. Gly-Tyr



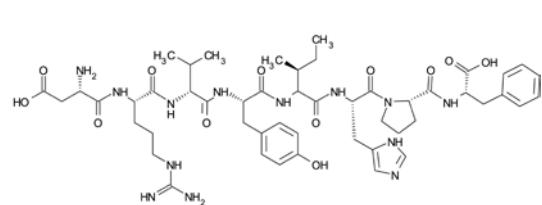
2. Tyr-Tyr-Tyr



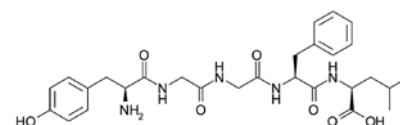
3. Val-Tyr-Val



4. Oxytocin



5. Angiotensin II



6. Leu-enkephalin

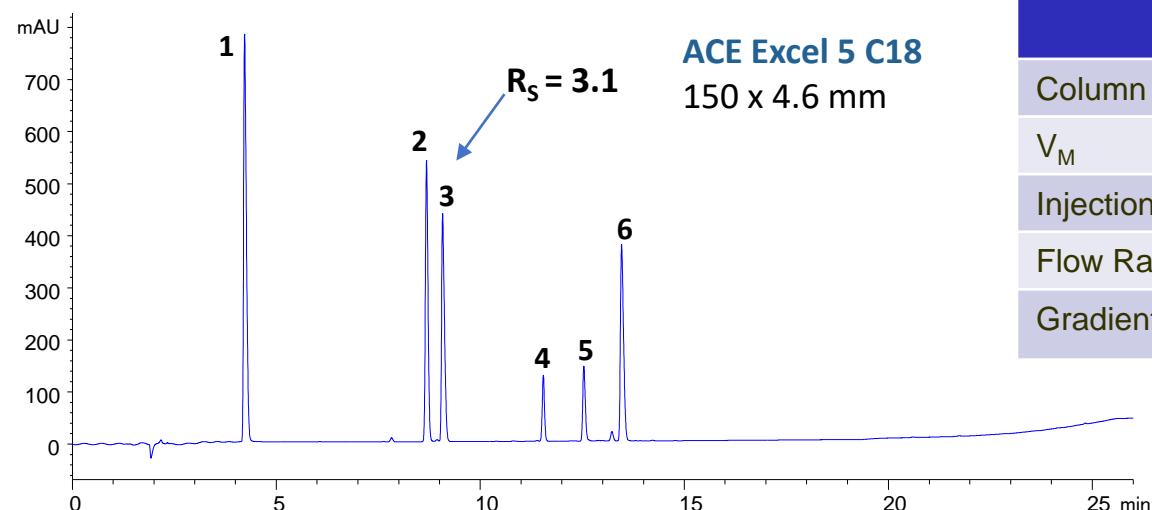
Mobile Phase A: 0.05% TFA (aq), B: 0.05% TFA in MeCN

Gradient: 5 to 40 %B in 17.0 mins, then 40 to 90 %B in 6.0 minutes and hold 90% B for 2.0 mins

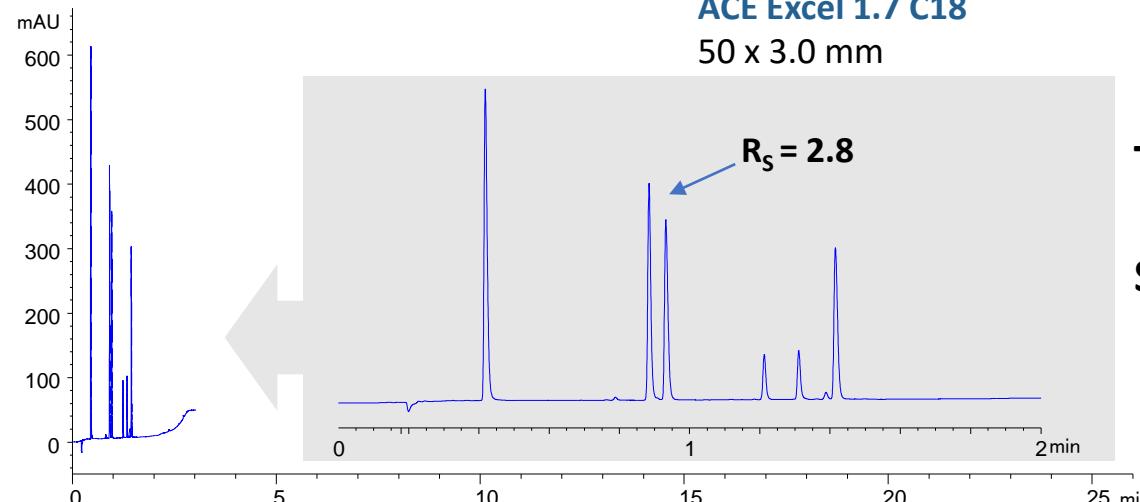
Temp: 60 °C Flow Rate: 1.0 mL/min

Detection: UV 220 nm.

UHPLC Gradient Method Translations: Peptides



	Original Method	Translated Method
Column	150 x 4.6 mm, 5 μ m	50 x 3.0 mm, 1.7 μ m
V_M	1.571 mL	0.223 mL
Injection Vol.	10 μ L	1.4 μ L
Flow Rate	1.0 mL/min	1.25 mL/min
Gradient time	17 min.	1.93 min.



Total run time: -88%

Solvent consumption: -85%



Gradient Method Translations: Correcting for V_D/V_M

- For maximum accuracy of gradient translations, it may be necessary to correct for the change in ratio of V_D/V_M between the original and translated method.

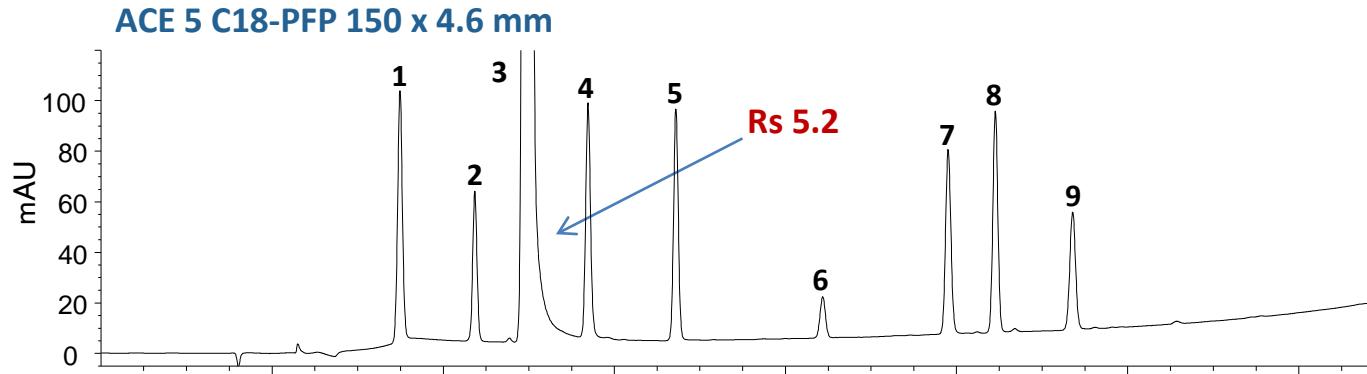
$$\Delta = \left(\frac{V_D}{V_M} \right)_{original} - \left(\frac{V_D}{V_M} \right)_{new} \quad must\ approach\ zero$$

- This is increasingly important when translating to small i.d. (therefore small V_M) UHPLC column formats.
- The correction can be made using

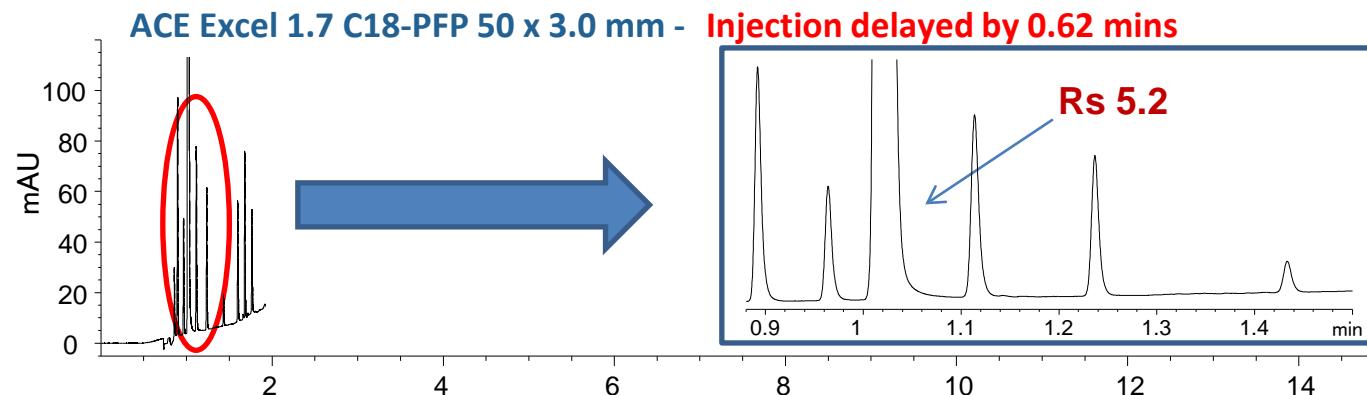
$$x = \left[\left(\frac{V_{D1}}{V_{M1}} \right) - \left(\frac{V_{D2}}{V_{M2}} \right) \right] \times \frac{V_{M2}}{F_2}$$

- Negative value:** injection must be delayed x minutes after gradient starts.
- Positive value:** a pre-gradient isocratic hold of x minutes should be added to gradient program.

Gradient Method Translations: Why correct for V_D/V_M ?



$t_G = 15$ minutes
 $F = 1.0 \text{ mL/min}$



$t_G = 1.70$ minutes
 $F = 1.25 \text{ mL/min}$

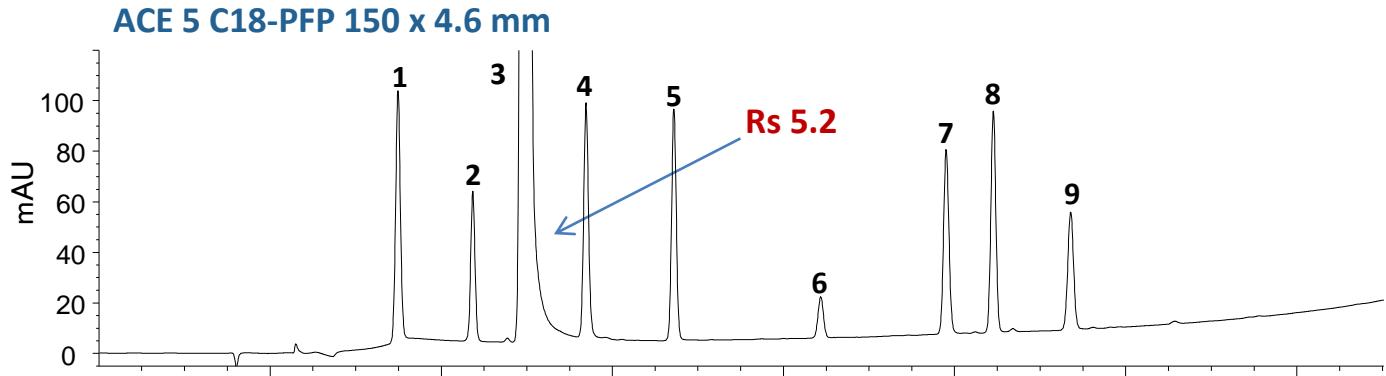
- Calculations tell us to **delay injection** until **0.62 minutes** after the gradient starts to correct for V_D/V_M

$$x = \left[\left(\frac{V_{D1}}{V_{M1}} \right) - \left(\frac{V_{D2}}{V_{M2}} \right) \right] \times \frac{V_{M2}}{F_2} = \left[\left(\frac{1.098}{1.570} \right) - \left(\frac{0.926}{0.223} \right) \right] \times \frac{0.223}{1.25} = \boxed{-0.62 \text{ minutes}}$$

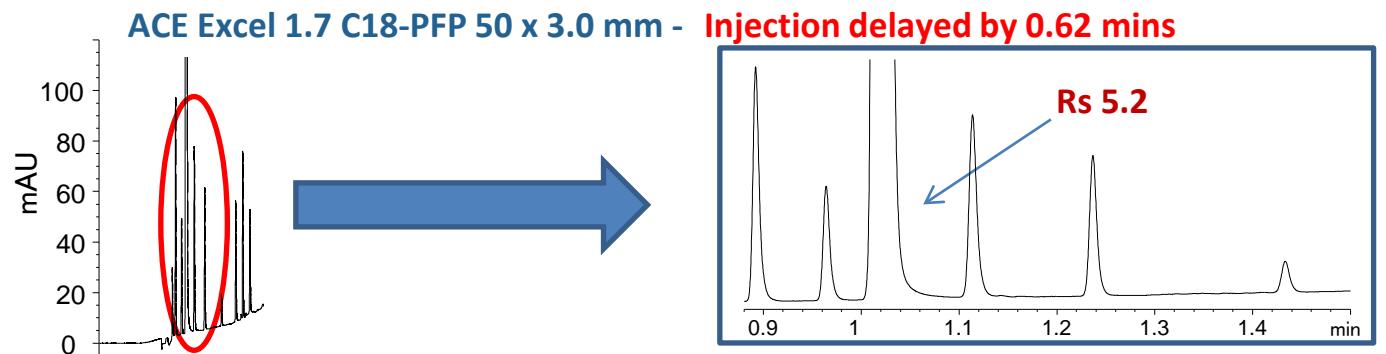
Gradient analysis, A= 20mM ammonium acetate pH 6.0 (aq), B= 20 mM ammonium acetate pH 6.0 in MeCN:water 80:20 v/v, 5-95% B in 15.0 mins, hold 95% B for 2.0 mins, 40°C, 1.0 mL/min, 230 nm.

1. 4-aminophenol, 2. hydroquinone, 3. 4-acetamidophenol (paracetamol), 4. 2-aminophenol, 5. 2-acetamidophenol, 6. phenol, 7. 4-nitrophenol, 8. 4-chloroacetanilide, 9. 2-nitrophenol. Impurities were spiked at 0.5% w/w.

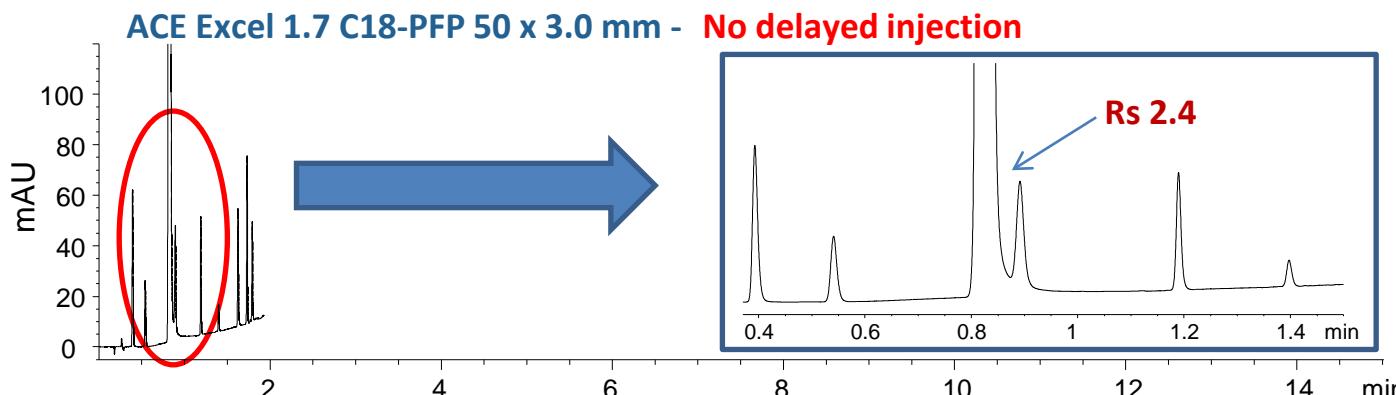
Gradient Method Translations: Why correct for V_D/V_M ?



$t_G = 15$ minutes
 $F = 1.0 \text{ mL/min}$



$t_G = 1.70$ minutes
 $F = 1.25 \text{ mL/min}$



$t_G = 1.70$ minutes
 $F = 1.25 \text{ mL/min}$



FREE ACE® Method Translator Tool: Gradient Example

Input data into
the grey boxes

Input dwell
volume of LC
systems

Gradient

Column Information		Translated													
Current		Translated													
Column Length (L)	150:mm	Column Length (L)	50:mm												
Column i.d. (d_o)	4.6:mm	Column i.d. (d_o)	3.0:mm												
Particle Diameter (d_p)	5.0: μ m	Particle Diameter (d_p)	1.7: μ m												
L/ d_p	30000	L/ d_p	29412												
Column Porosity	0.63:What's This?	Column Porosity	0.63:What's This?												
Column Volume (V_M)	1.570 mL	Column Volume (V_M)	0.223 mL												
Method		Translated													
Current		Translated													
Injection Volume	10.0: μ L	Injection Volume	1.4 μ L												
Flow Rate	1.00:mL/min	Flow Rate (scaled to linear velocity)	0.43 mL/min												
LC Name	HPLC 1	Flow Rate (scaled to particle size)	1.25 mL/min												
Dwell Volume (V_D)	1.098:mL	Input Flow Rate	1.25 mL/min												
Recorded Backpressure	67:bar	LC Name	UHPLC 1												
Gradient	<table border="1"><thead><tr><th>Time</th><th>%B</th></tr></thead><tbody><tr><td>0.00</td><td>5</td></tr><tr><td>15.00</td><td>95</td></tr><tr><td>17.00</td><td>95</td></tr><tr><td>17.50</td><td>5</td></tr><tr><td></td><td></td></tr></tbody></table>	Time	%B	0.00	5	15.00	95	17.00	95	17.50	5			Estimated Backpressure	568 bar
Time	%B														
0.00	5														
15.00	95														
17.00	95														
17.50	5														
Suggested Re-equilibration Time	16.8 mins	Estimated Run Time Difference	-87 %												
		Estimated Solvent Use Difference	-83 %												
Gradient	<table border="1"><thead><tr><th>Time</th><th>%B</th></tr></thead><tbody><tr><td>0.00</td><td>5</td></tr><tr><td>1.70</td><td>95</td></tr><tr><td>1.93</td><td>95</td></tr><tr><td>1.98</td><td>5</td></tr><tr><td></td><td></td></tr></tbody></table>	Time	%B	0.00	5	1.70	95	1.93	95	1.98	5			Time to delay injection after the gradient begins	0.62 mins (770 μ L)
Time	%B														
0.00	5														
1.70	95														
1.93	95														
1.98	5														
		Correct translation of this method requires that the injection is delayed until after the gradient begins by this time. A delayed injection can be added to a method in many LC instrument software packages.													

New Gradient

This section tells you if a
delayed injection of X mins is
required

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MAC-MOD.com

Method Transfer Between Different instruments



Instrument to Instrument Same Method Transfers

◆ Isocratic

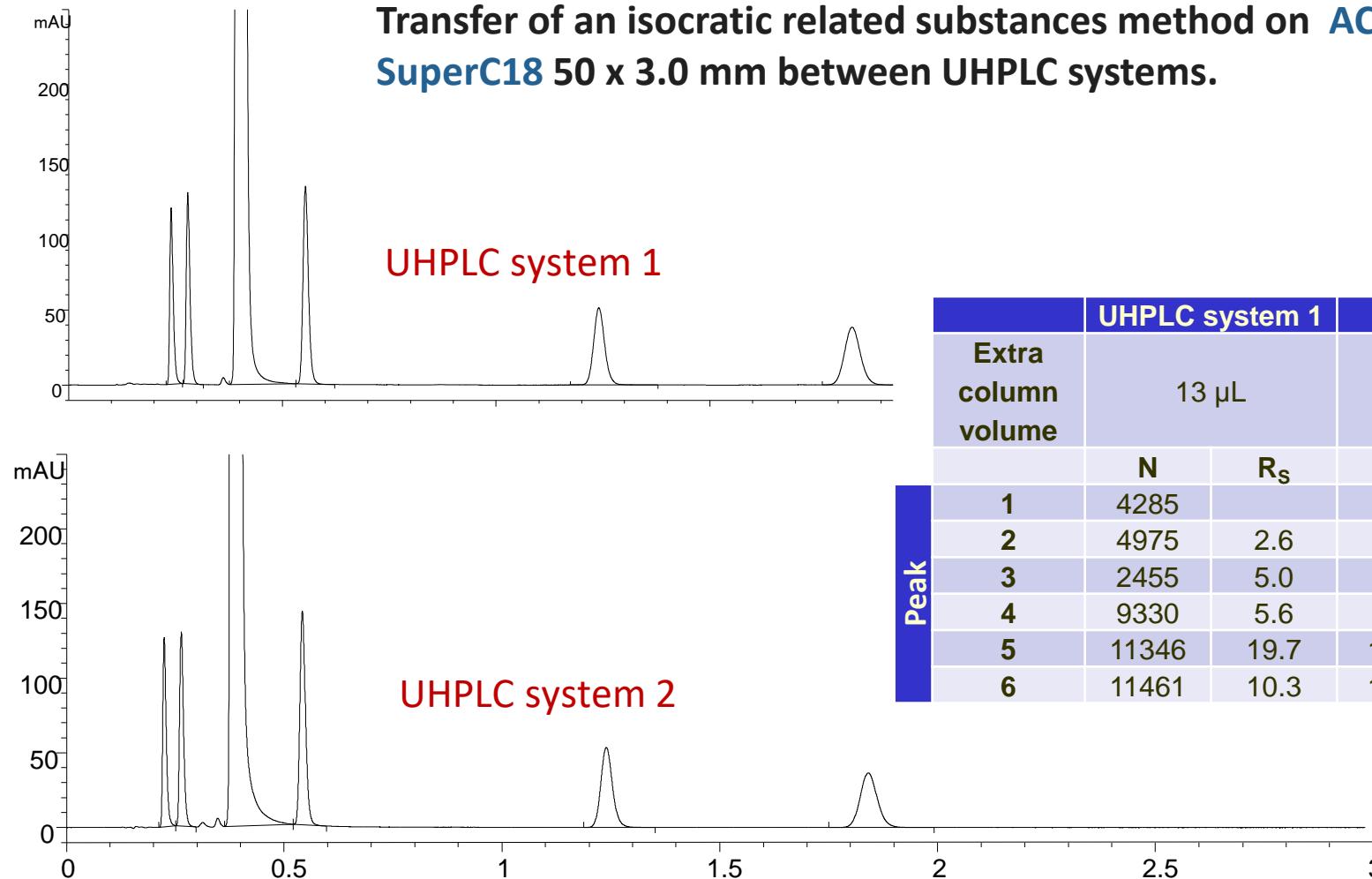
- If column format **remains identical**, **no changes** necessary.
- (If column format changes, translation required.)

◆ Gradient

- If column format **remains identical**, need to **correct for influence of differing system dwell volumes** only. Flow rate, injection volume and gradient times remain **unchanged**.
- (If column format changes, translation plus differing system dwell effects need to be calculated.)



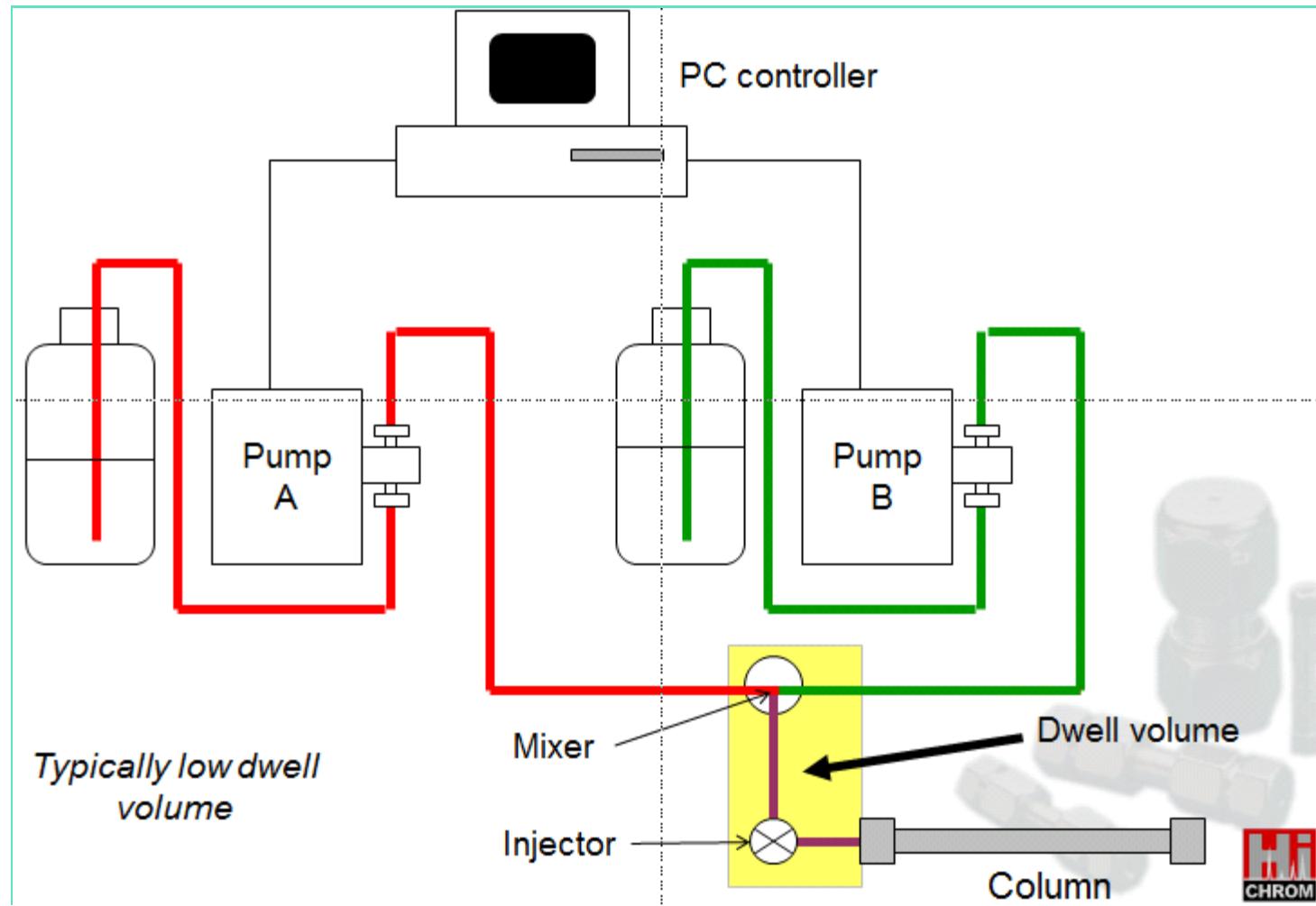
Instrument to instrument method transfer: Isocratic



Isocratic analysis 0.2% H₃PO₄ in MeCN:MeOH:water 35:5:60 v/v/v, 40°C, 1.25 mL/min, 0.7 µL , Injection vol., 254 nm.
1. 4-hydroxybenzoic acid, 2. 4-hydroxyisophthalic acid, 3. acetylsalicylic acid (aspirin), 4. salicylic acid, 5. acetylsalicylsalicylic acid, 6. salsalate. Impurities were spiked at 0.5% w/w.

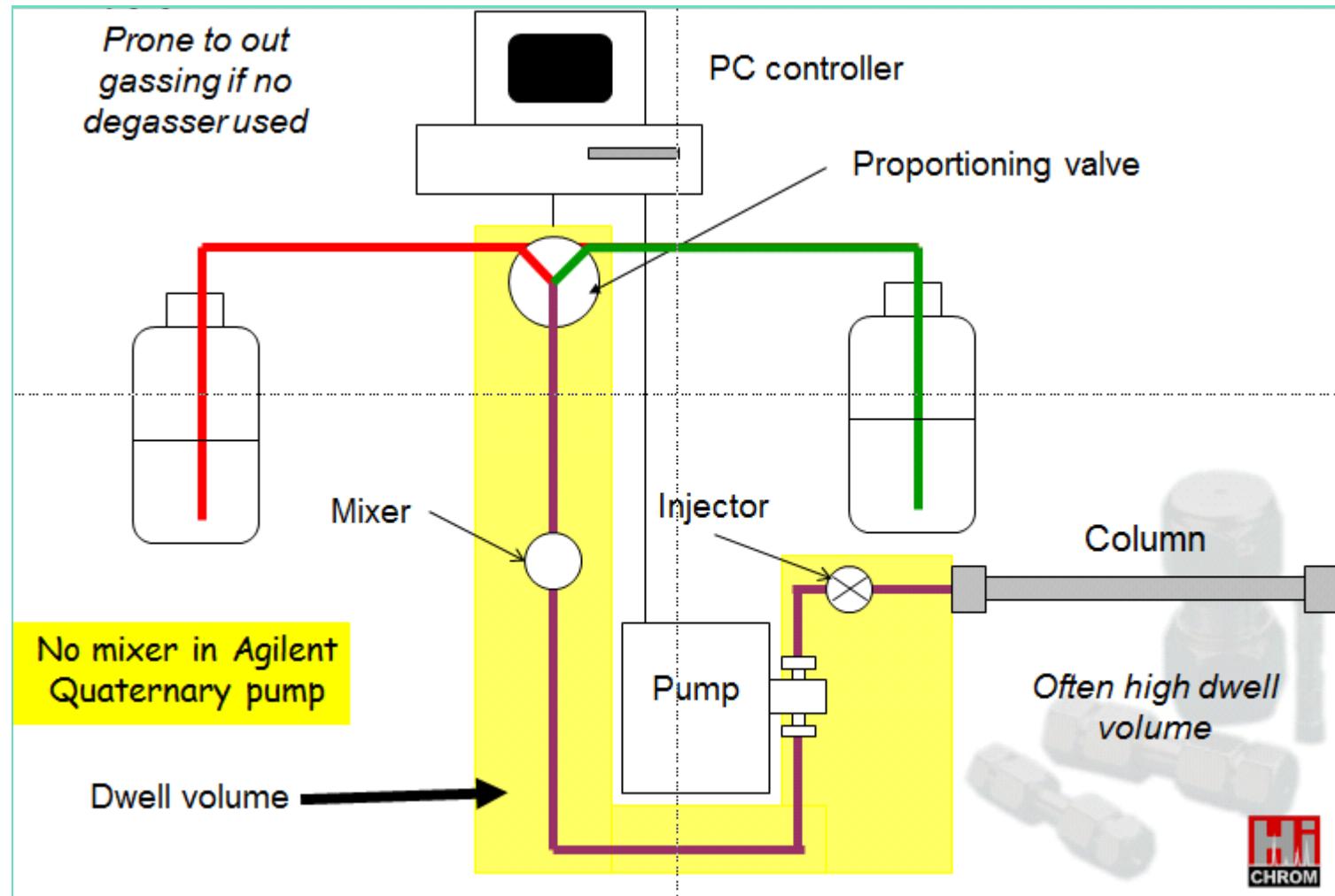


Binary Pump Configuration: High Pressure Mixing





Quaternary Pump Configuration: Low Pressure Mixing





Instrument to Instrument Same Method Transfers

- ◆ For gradient method transfer, t_G , F and V_M all remain identical.
- ◆ Only need to correct for the system dwell volume differences
 - Given that V_M does not change for method transfer, the V_D/V_M equation simplifies to:

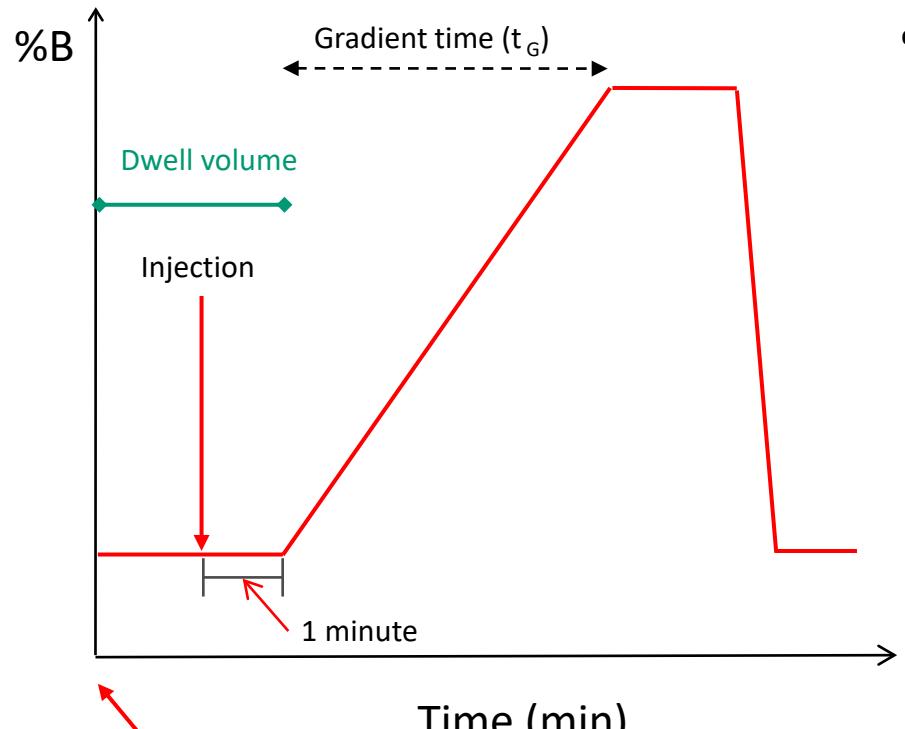
$$x = \left(\frac{V_{D1} - V_{D2}}{F} \right)$$

- Negative value: injection must be delayed x minutes after gradient starts so injection is made onto the gradient.
- Positive value: a pre-gradient isocratic hold of x minutes should be added to gradient program.

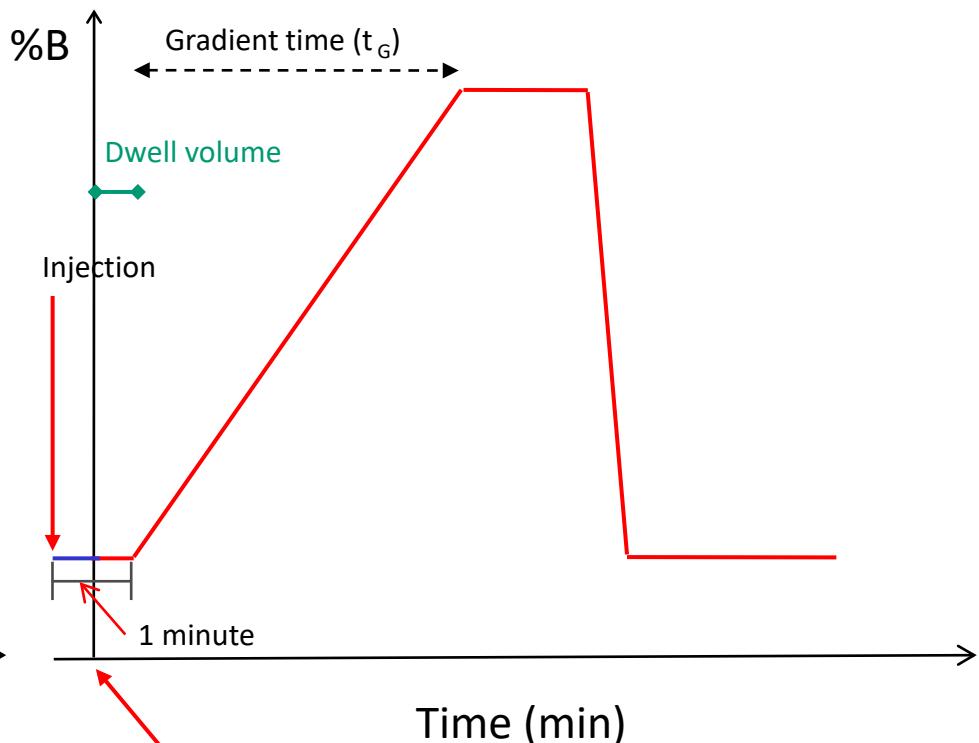


Method Transfers: Dwell Volumes / Injection Times

e.g. small column on a high dwell volume system. The dwell time is artificially long.



e.g. large column on a low dwell volume system. The dwell time is artificially short.



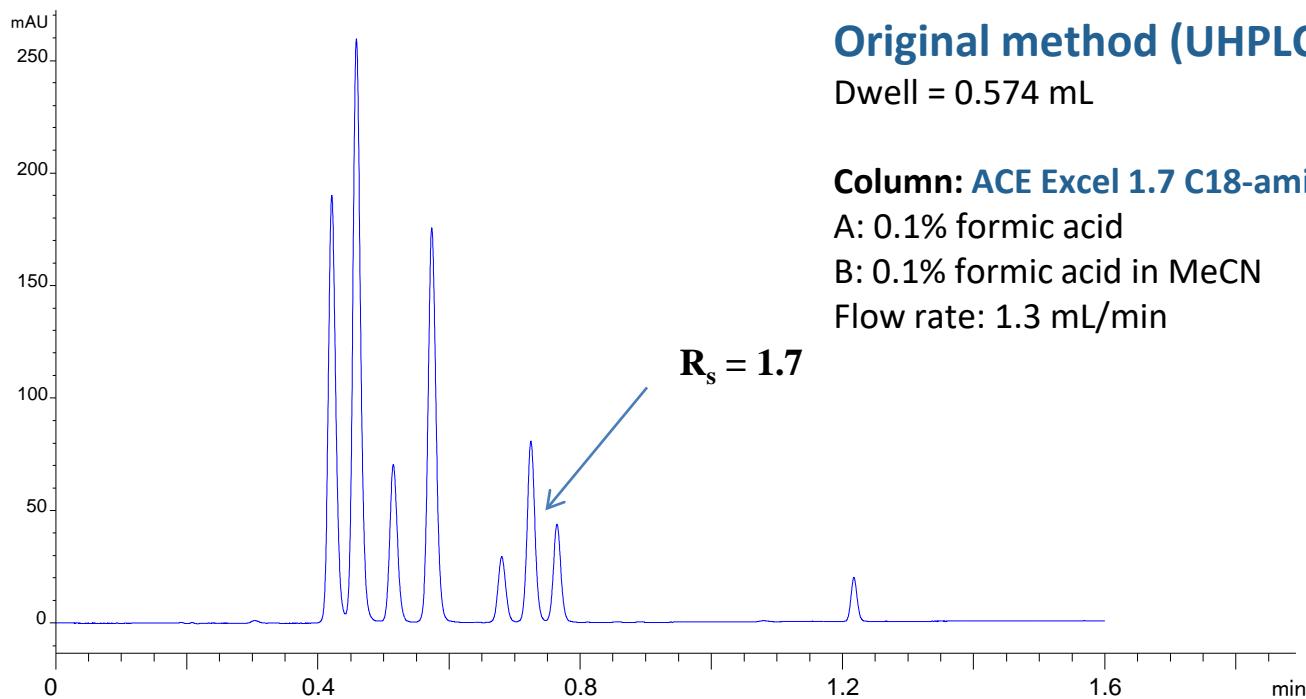
$$\Delta (V_D/V_M) = \text{negative}$$

Injection is delayed until after the start of gradient

$$\Delta (V_D/V_M) = \text{positive}$$

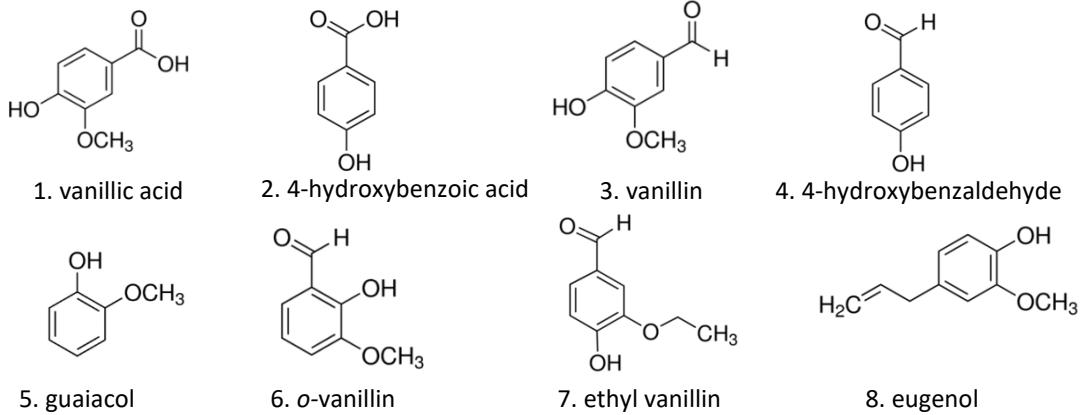
Pre-gradient hold is added. Effectively extends dwell time

UHPLC method transfer - Vanillins



	t	%B
	0	25
	1.32	75
	1.49	75
	1.6	25

t_G 1.32 min
Post time: 2 min
Inj. Vol. 1 μL
P_{MAX} 502 bar





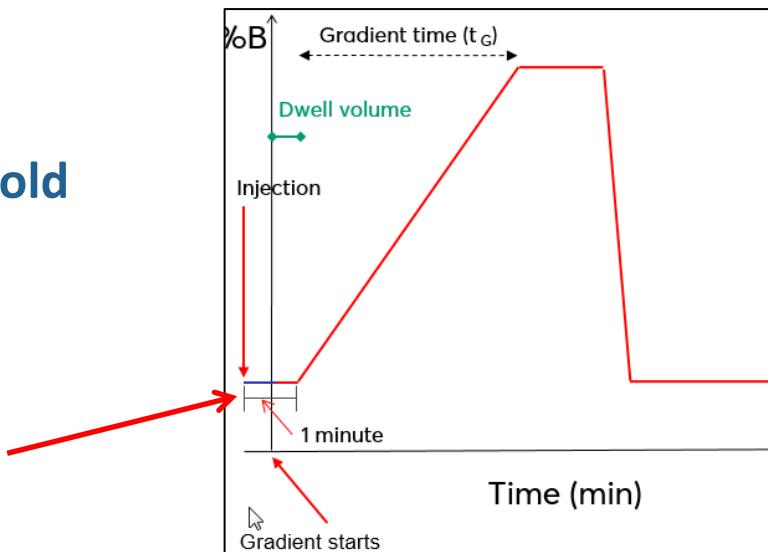
UHPLC method transfer - Vanillins

◆ Aim

- To translate the method from UHPLC System 1 (Quaternary) to UHPLC System 2 (Binary)
- Correct for system dwell volume only
- $V_{D1} - V_{D2} = \Delta = 0.574 - 0.202 = 0.372 \text{ mL}$
- Positive value, therefore a pre-gradient hold (x) is required

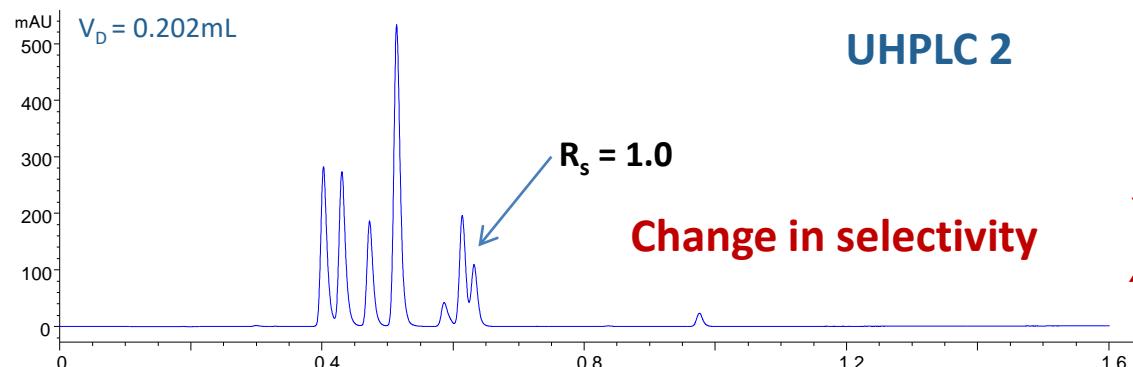
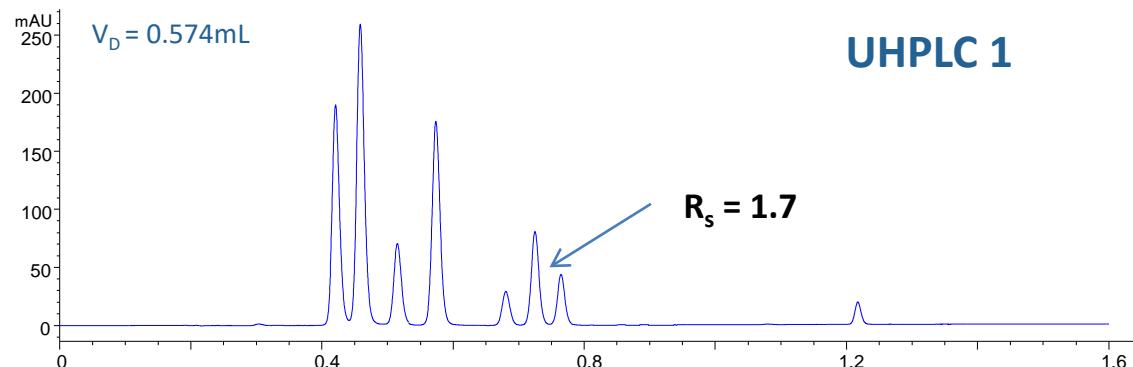
$$x = \frac{|\Delta|}{F_2} = 0.29 \text{ min pre-gradient hold}$$

Essentially introducing
an isocratic hold prior
to gradient start





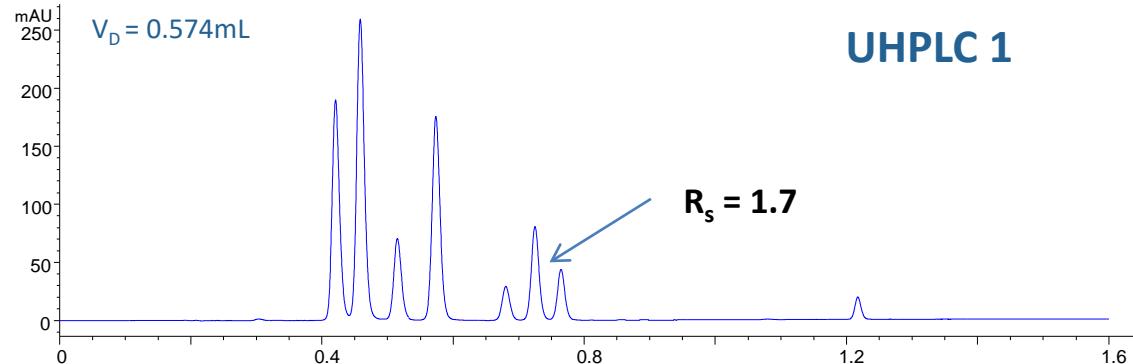
Instrument to instrument method transfer



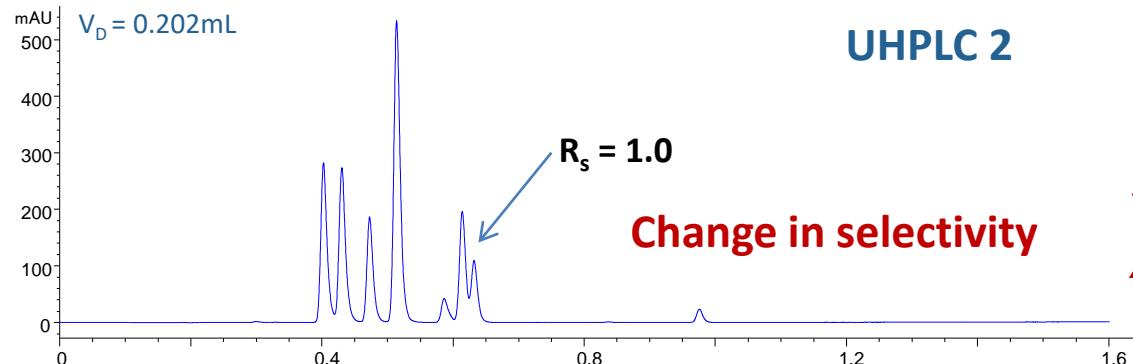
Without isocratic hold



Instrument to instrument method transfer

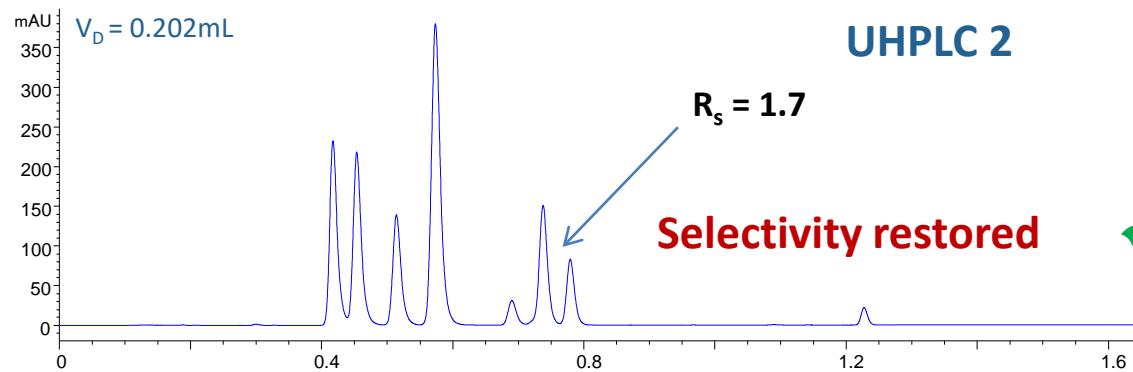


t	%B
0	25
1.32	75
1.49	75
1.6	25



t	%B
0	25
1.32	75
1.49	75
1.6	25

Without isocratic hold



t	%B
0	25
0.29	25
1.61	75
1.78	75
1.89	25

**With correction
for ΔV_D**





Method Transfer Tool Example - Gradient

V1.4

Method Transfer

Method transfer involves moving a method from one LC instrument to another, whilst keeping the column format constant. Isocratic analysis is simple with no method changes required. Gradient methods however, should be adjusted to account for any change in system dwell volume in order to ensure accurate method transfer. This tool automatically determines any correction required to ease method transfer. See the Dwell Volume tab for details of how to determine system dwell volume.

Column Information

Column Length (L)	50 mm
Column i.d. (d _c)	3.0 mm
Particle Diameter (d _p)	1.7 µm
L/d _p	29412
Column Porosity	0.63 <small>(What's This?)</small>
Column Volume (V _M)	0.223 mL

Dwell volumes of both systems entered

Method

Current		Transferred													
Flow Rate	1.300 mL/min	LC Name	Ultra RS												
LC Name	1290Q	Dwell Volume (V _D)	0.284 mL												
Dwell Volume (V _D)	0.574 mL	Gradient	<table border="1"><thead><tr><th>Time</th><th>%B</th></tr></thead><tbody><tr><td>0.00</td><td>25.0</td></tr><tr><td>1.32</td><td>75.0</td></tr><tr><td>1.49</td><td>75.0</td></tr><tr><td>1.60</td><td>25.0</td></tr><tr><td>1.82</td><td>25.0</td></tr></tbody></table>	Time	%B	0.00	25.0	1.32	75.0	1.49	75.0	1.60	25.0	1.82	25.0
Time	%B														
0.00	25.0														
1.32	75.0														
1.49	75.0														
1.60	25.0														
1.82	25.0														

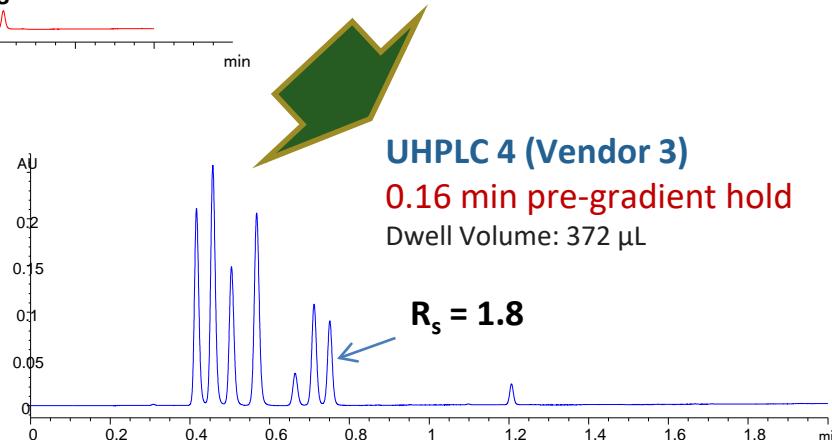
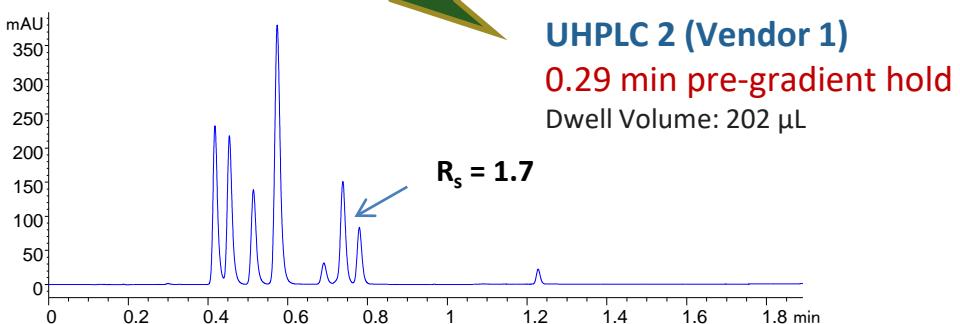
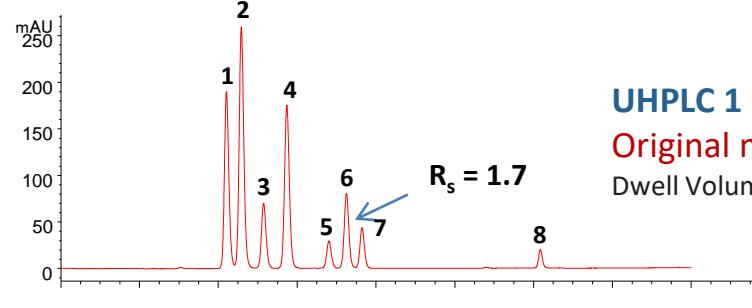
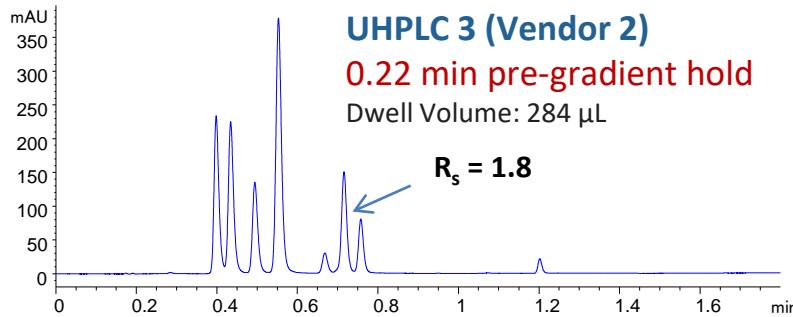
New Gradient with isocratic hold

**Download free at
MAC-MOD.com**



Instrument to instrument method transfer

- Using the ACE Method Transfer Tool, gradient method transfer between different vendor LC systems is easily achieved.



Summary #1

- ◆ UHPLC can provide many benefits:
 - Increased separation speed
 - Increased resolution
 - ‘Ultra-resolution’ with column coupling for complex samples
- ◆ Stationary phase selectivity is a powerful tool for UHPLC (and HPLC!) method development
 - Different stationary phases provide different mechanisms of analyte interaction.
 - Five **ACE novel phases** (C18-AR, C18-PFP, C18-Amide, CN-ES and SuperC18) plus ACE C18 provide total solution for column screening.



Summary #2

- ◆ Translation of HPLC methods to UHPLC can be accomplished by using a few key equations:
 - Isocratic: fairly simple
 - Gradient: more complex (translate t_G and correct for dwell volume)
- ◆ Method transfers between instruments also straightforward

The ACE LC Translator simplifies method translation and instrument transfer

Download free at
MAC-MOD.com

Useful Resources

- ◆ ACE Translation Tool:

- ◆ (+help file)
- ◆ (+ AKN#0023)



- ◆ ACE Knowledge Notes (AKNs):

- ◆ [AKN0001 - How to Determine System Dwell Volume](#)
- ◆ [AKN0006 – UHPLC Column Connections](#)
- ◆ [AKN0011 – Practical UHPLC](#)
- ◆ [AKN0012 - Understanding the Relationship between Particle Size, Performance and Pressure](#)
- ◆ [AKN0017 - How to Determine Extra Column Dispersion and Extra Column Volume](#)
- ◆ [AKN0023 - Gradient Method Translation Using the ACE LC Translator](#)

Gradient Method Translation Using the ACE LC Translator

ABSTRACT

Translating gradient LC methods from one column dimension to another can be challenging. For the most common method translation, which involves moving a gradient segment from one column to another, knowledge of fundamental chromatography principles is required. Optimizing key parameters in subsequent calculations is critical to ensure the method remains effective and accurate when translated to a different column dimension. The ACE LC Translator is based upon fundamental principles and includes all key parameters and calculations required for an accurate gradient method translation process.

Introduction

Method translation involves moving an LC method from one column dimension to another. Additionally, the method may be moved to a new LC system at the same time. While the method is being moved, many minor particles, many legacy LC methods are designed for a specific column dimension (e.g. 10 x 25 x 4.6 mm). For example, a legacy method may be developed for a large column (e.g. 10 x 250 x 4.6 mm). Conversely, while new method development settings are being used (e.g. 10 x 100 x 1.7 mm), there may be a need to translate the method to UHPLC for routine use. The principal cost for method translation is to maintain the chromatographic selectivity and performance of the original method with the newly translated method.

Simplified Method Translation

Assuming gradient method translation requires moving a single gradient segment from one column dimension to the new column dimension, it is often helpful to know the effect of the gradient changes to the column volume and system dwell volume. A simplified method has been developed to help achieve successful method translation by automatically applying calculated gradient parameters using the original method, system dwell variables, and target column details as input.

Example: Translating a method from HPLC to UHPLC

Figure 1 shows an HPLC separation of a mixture of alcohols and ester impurities on an ACE Excel 5 C18-RP, 150 x 4.6 mm column. The separation conditions are 5 uL injection volume, 5 uL/min flow rate, and a gradient from 0 to 50 x 1.7 mm UHPLC column.

Figure 1: Separation of alcohols and ester impurities on an ACE Excel 5 C18-RP, 150 x 4.6 mm (Flow rate: 1 mL/min; injection volume: 5 μ L) See ACE Application Note AN0150 for full method and sample details.

For more information contact your local ACE distributor or visit www.ace-lc.com or email: info@ace-lc.com

- ◆ ACE Method Development Kit Brochure and Webinar

- ◆ Download Resources at mac-mod.com

or info@mac-mod.com

ACE Method Development Kits Intelligent Solutions for Method Development

Intelligent Solutions for Method Development

ACE Method Development Support

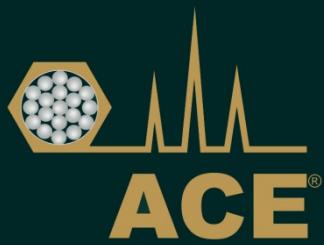
• HPLC and UHPLC method development kits available for the most common column types

• All ACE Method Development Kits include a detailed user manual, access to our extensive knowledge base, and free technical support.

• ACE Method Development Kits are designed to help you quickly and easily develop methods for your applications.

• Learn more about our method development tools at www.ace-lc.com

Learn More: ace-lc.com or info@ace-lc.com



Thank you for your attention

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