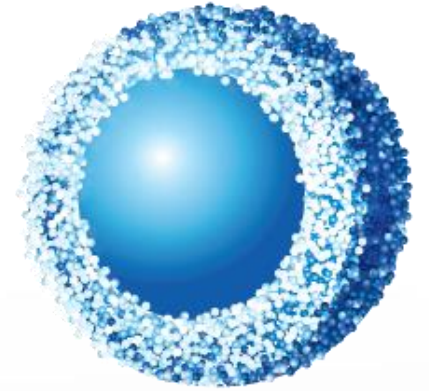


Wide Pore Superficially Porous Particles with Various Bonded Phases for High Resolution Protein Chromatography



William Miles, Barry Boyes, Ben Libert, Stephanie Schuster, Brian Wagner, Conner McHale

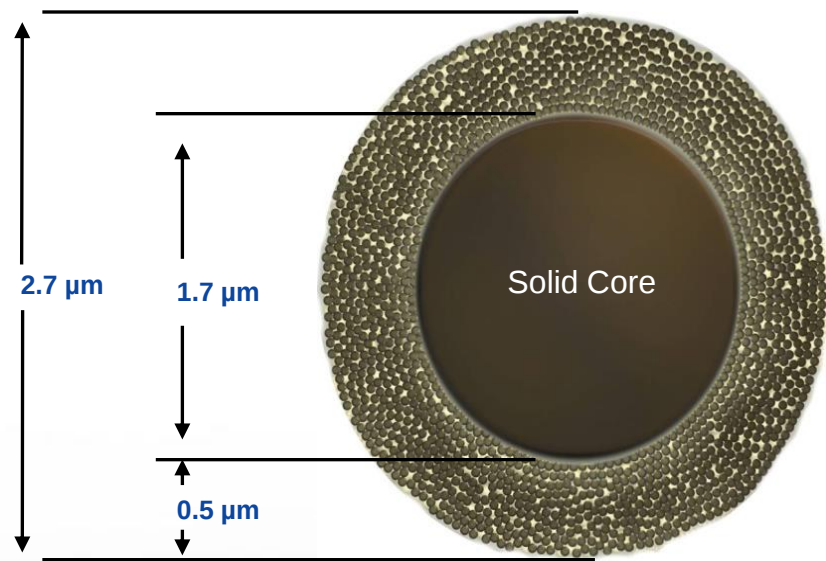
Associate Scientist, R&D

Advanced Materials Technology, Inc.

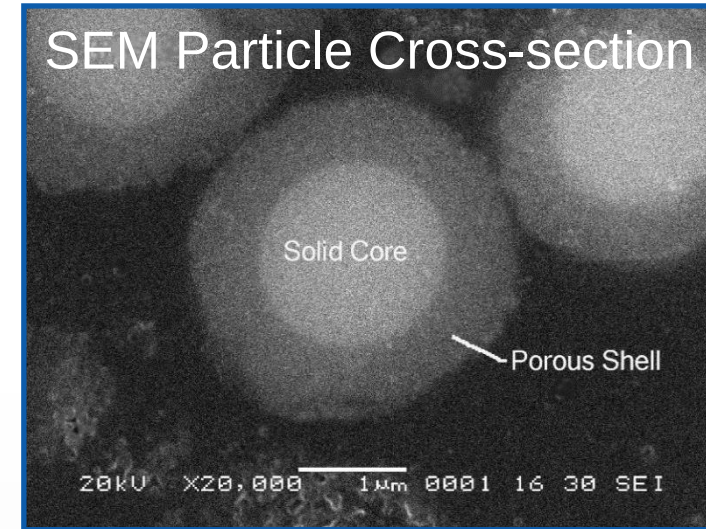
Wilmington, Delaware, USA

wmiles@advanced-materials-tech.com

Superficially Porous Particles (90 Å): 2006



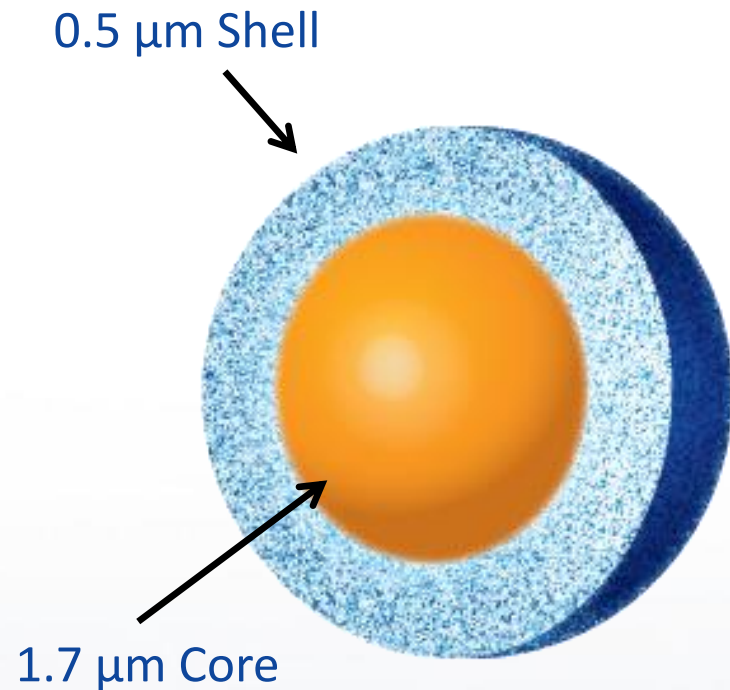
Porous Shell
90 Å



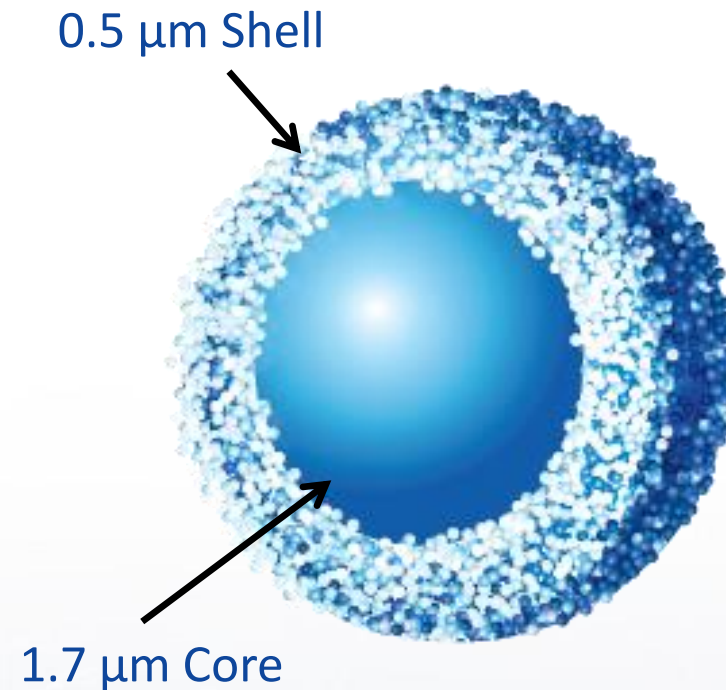
Fully Porous Particles (FPP) vs. Superficially Porous Particles (SPP)

- Lower back pressure
- Higher efficiency / resolution:
 - Narrow particle size distribution
 - Improvements in mass transfer
- Maintains high resolution at high flow rates, flat C-term in van Deemter plots

Developments in HALO[®] Fused-Core[®] Pore Size



90 Å, 2.7 μm
135 m^2/g
2006

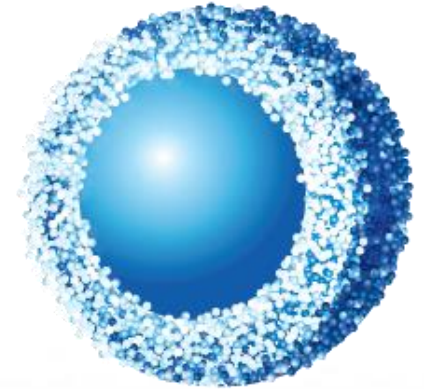


1000 Å, 2.7 μm
22 m^2/g
2017

Wide Pore SPP Benefits Protein Science

What is needed for high performance separations of larger (Bio) molecules?

- **Pore size must “fit” molecule size**
 - Restricted diffusion limits efficiency and load capacity
 - Peak capacity effects by kinetic and retention limitations
- **Particle morphology must optimize surface area/volume**
 - Shell thickness determines diffusion path and surface area
 - Must have “Right” size and desirable particle distribution
- **Surface chemistry appropriate to samples**



HALO
1000 Å, 2.7μm

Very Large Pore SPP

Surface Chemistry Options

Experimental: HPLC Methods

Reversed-Phase Liquid Chromatography (RPLC) separations of proteins and mAbs

Standard Protein RPLC methods

- Gradient elution *30-45% ACN in 15 minutes*
- Strong Solvent *acetonitrile (ACN), n-propanol (nProp)*
- Weak Solvent (H_2O)
- Ion Pair Reagent *trifluoroacetic acid (TFA) or difluoroacetic acid (DFA)*
- Columns *2.1x150mm*
- Flow Rate *0.2-0.6 mL/min with 2.1mm ID*
- High column temperature *60-90°C*
- 280nm UV absorbance

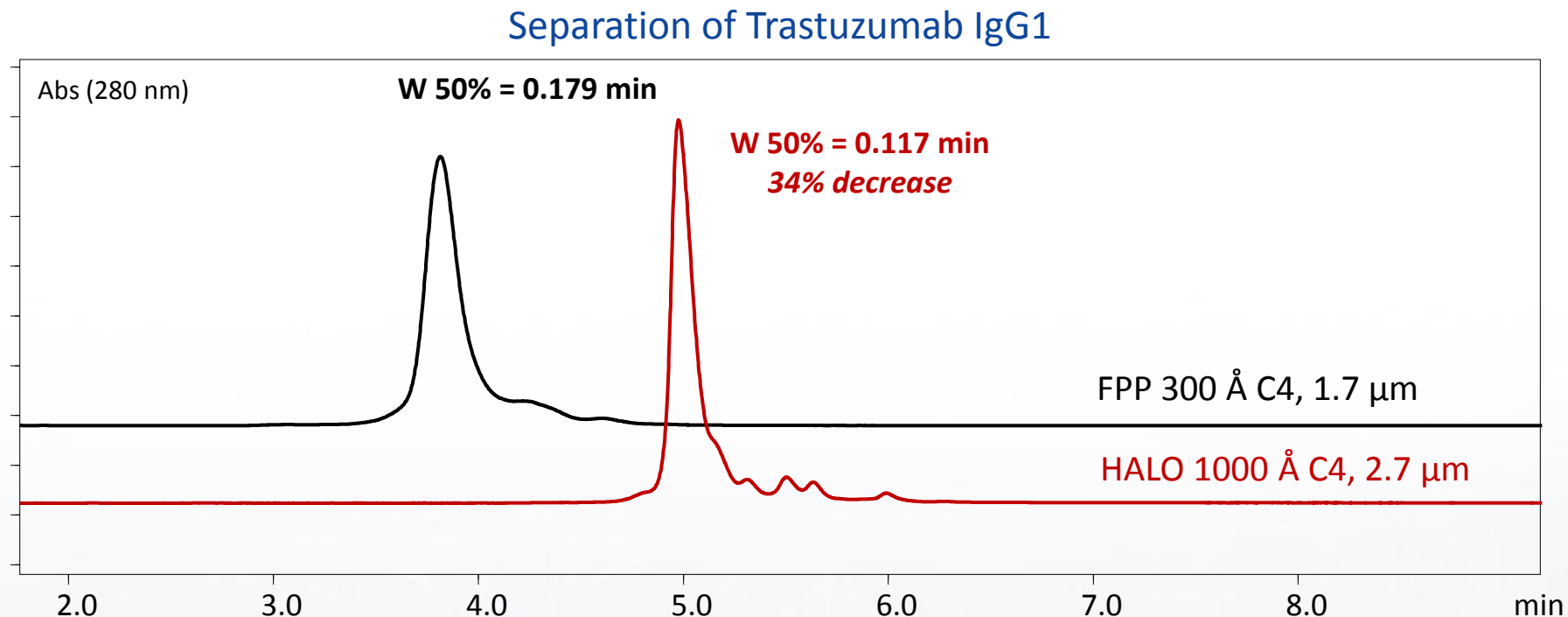
IgG1 Separation on HALO 1000 Å vs FPP 300 Å

Peak Width

- Larger improvement for HALO 1000 Å

Retention

- Increase for HALO 1000 Å



MP A – aq. 0.1% TFA; MP B – ACN + 0.1% TFA; 34-42% B in 16 min
Flow: 0.4 mL/min; Temp: 60°C; Inj.Vol: 2 μL (4 ug)

IgG1 Separation on HALO 1000 Å vs FPP 300 Å

Peak Width

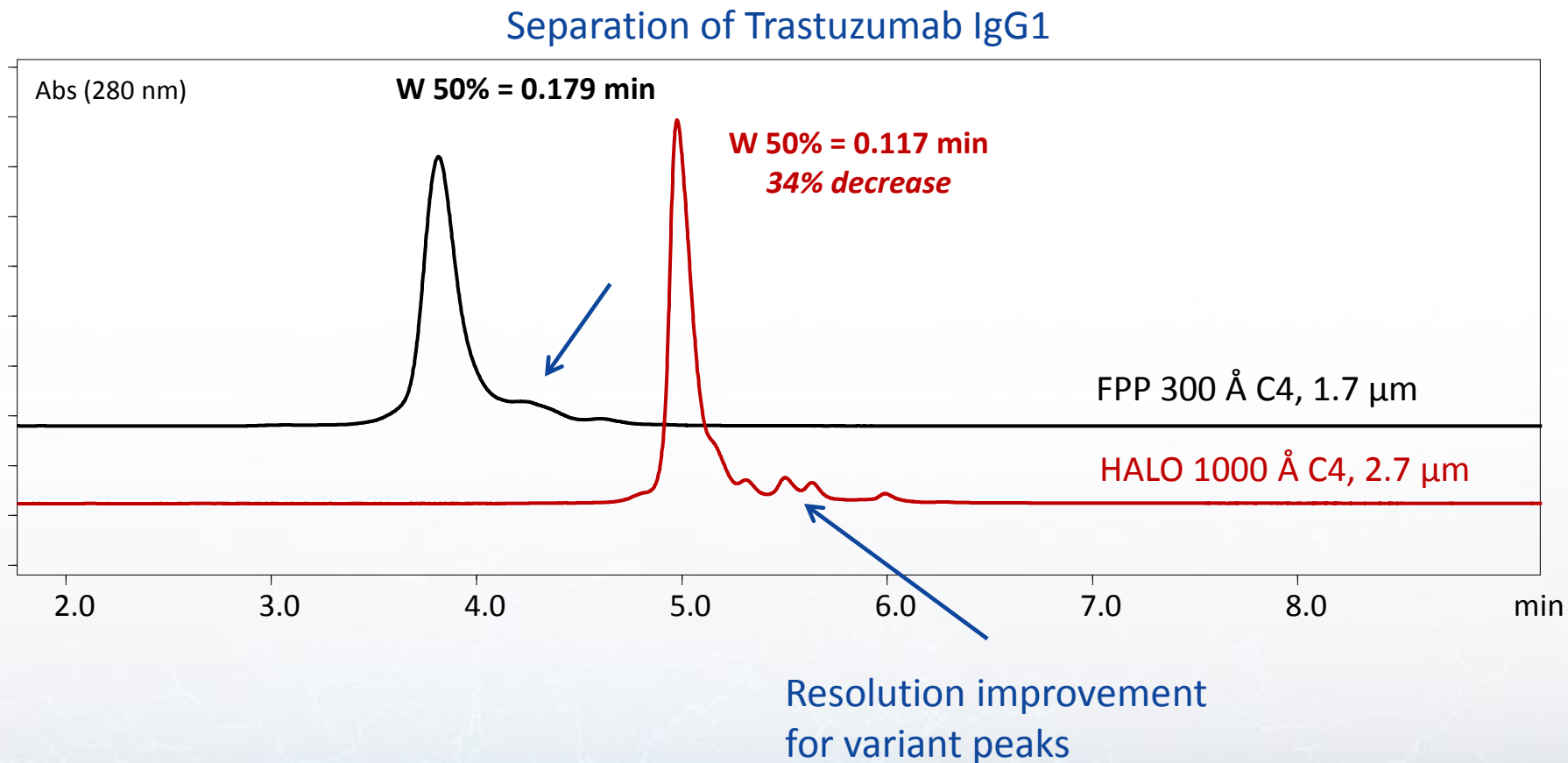
- Larger improvement for HALO 1000 Å

Retention

- Increase for HALO 1000 Å

Resolution

- Increase in R_s for later eluting minor variant peaks on HALO 1000 Å



MP A – aq. 0.1% TFA; MP B – ACN + 0.1% TFA; 34-42% B in 16 min
Flow: 0.4 mL/min; Temp: 60°C; Inj.Vol: 2 μL (4 ug)

IgG1 Separation on HALO 1000 Å vs FPP 300 Å

Peak Width

- > 30% decrease in width for both 1000 Å phases

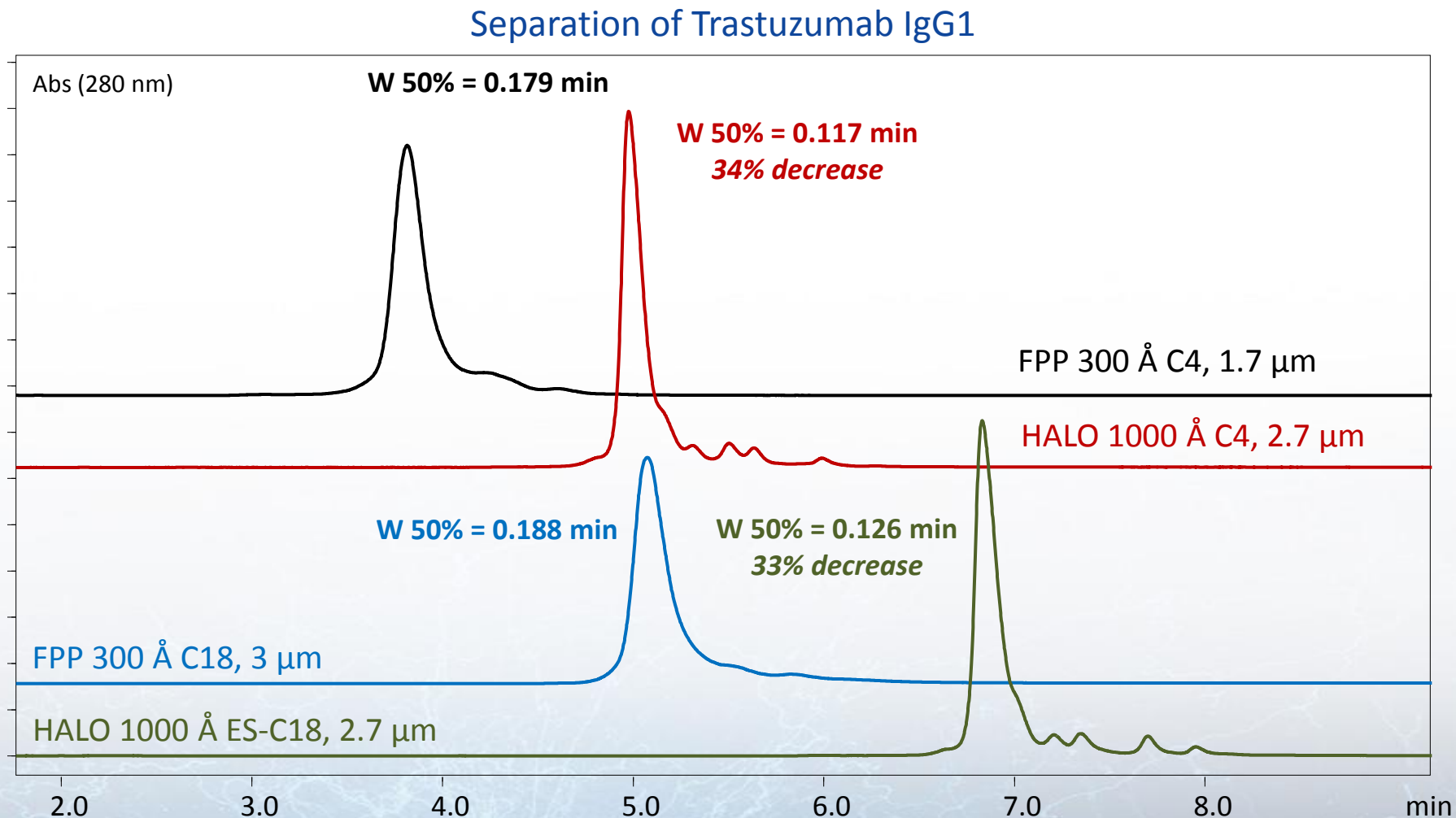
Retention

- Increase for HALO 1000 Å

Resolution

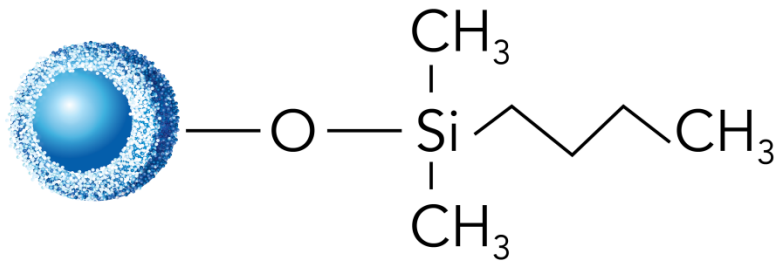
- Increase in R_s for later eluting minor variant peaks on HALO 1000 Å

Improvements seen on both C4 and ES-C18 bonded-phase comparisons



MP A – aq. 0.1% TFA; MP B – ACN + 0.1% TFA; 34-42% B in 16 min
Flow: 0.4 mL/min; Temp: 60°C; Inj.Vol: 2 μL (4 ug)

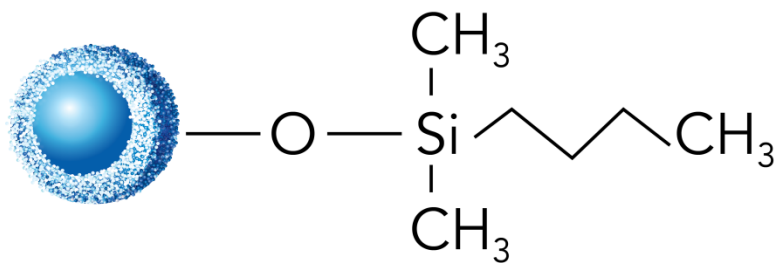
HALO 1000 Å Bonded-Phases



C4

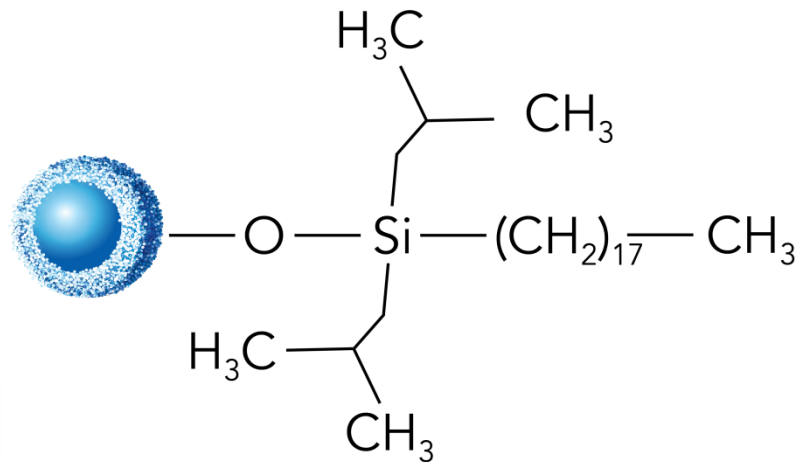
- Traditional alkyl phase for protein separations
- 4.1 $\mu\text{mol}/\text{m}^2$ (0.65% C)

HALO 1000 Å Bonded-Phases



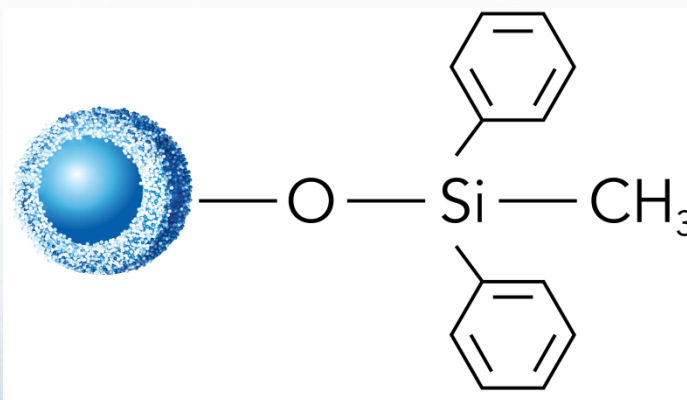
C4

- Traditional alkyl phase for protein separations
- $4.1 \mu\text{mol}/\text{m}^2$ (0.65% C)



ES-C18

- Sterically-protected, long chain alkyl phase
- $1.9 \mu\text{mol}/\text{m}^2$ (1.4% C)



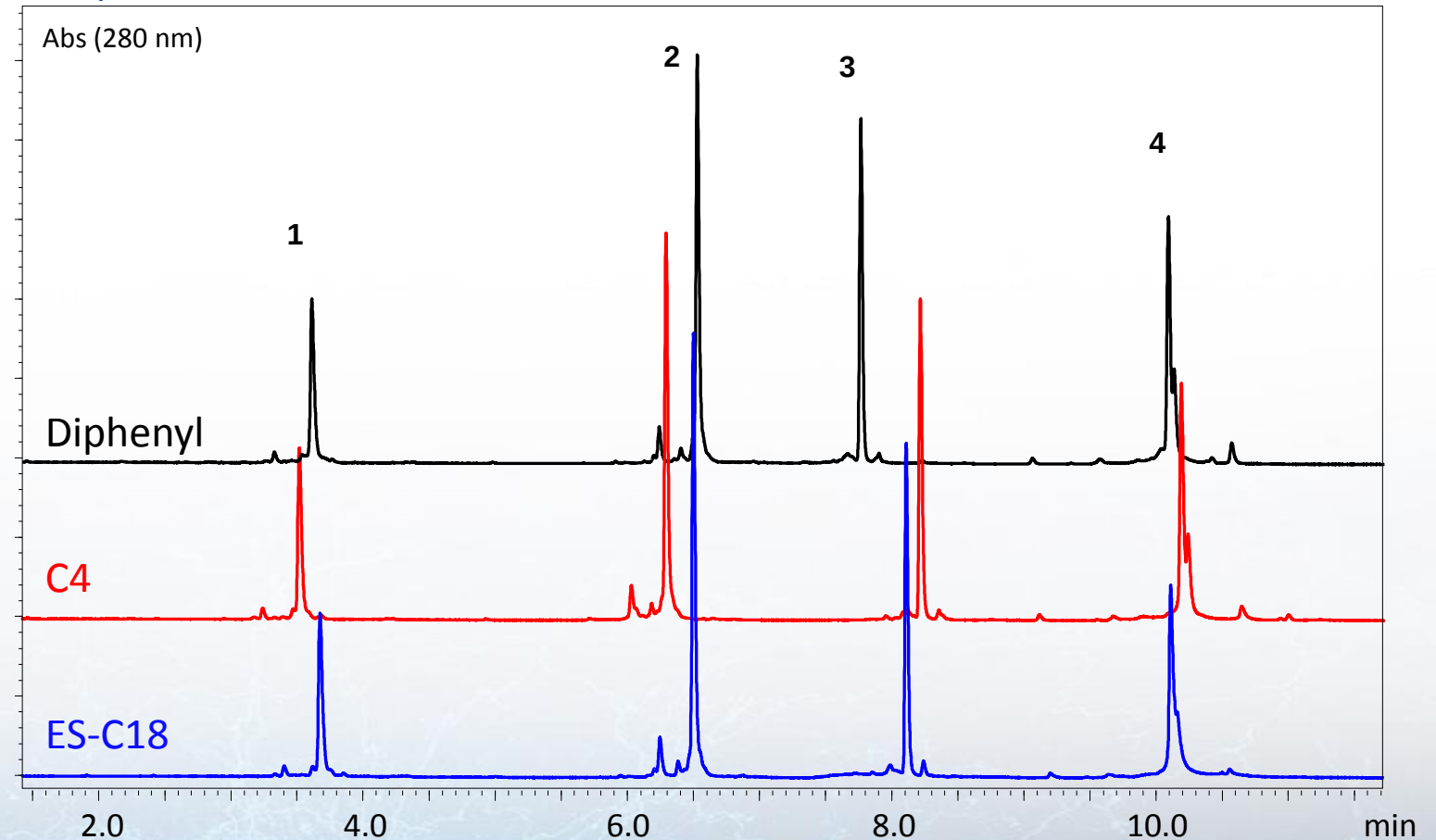
Diphenyl

- Aromatic phenyl phase
- Selectivity differs from alkyl phases
- $2.7 \mu\text{mol}/\text{m}^2$ (1.0% C)

Effect of Bonded-Phase on Retention/Selectivity

- **Hydrophobic, Small Molecule Retention**
 - Diphenyl < C4 << ES-C18
- **Protein Retention**
 - No global retention pattern
 - Differences in individual protein retention time

Separation of 4 Protein Mixture on HALO 1000 Å Bonded-Phases



1. RNase A – 13.7 kDa
2. Lysozyme – 14.3 kDa
3. α -Lactalbumin – 14.2 kDa
4. Enolase – 46.6 kDa

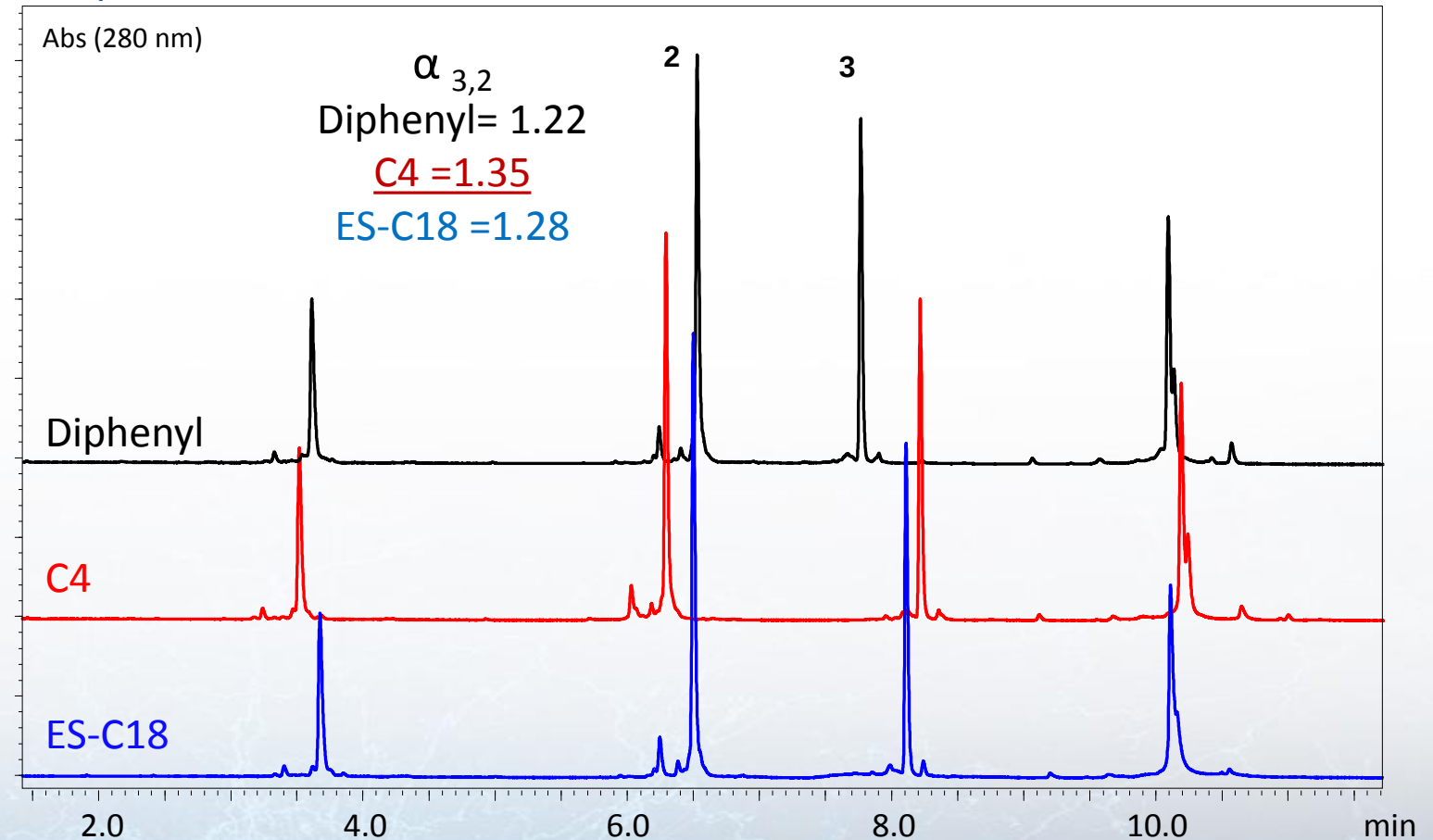
MP A: aq. 0.1% TFA; MP B: ACN + 0.1% TFA; Gradient: 20-60 %B in 15 min;
Injection Volume: 2 μ L; Flow: 0.40 mL/min; Temp: 80 $^{\circ}$ C

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 - Diphenyl < C4 << ES-C18
- **Protein Retention**
 - No global retention pattern
 - Differences in individual protein retention time
- **Protein Selectivity**
 - Subtle but useful changes

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4. Enolase – 46.6 kDa

Separation of 4 Protein Mixture on HALO 1000 Å Bonded-Phases



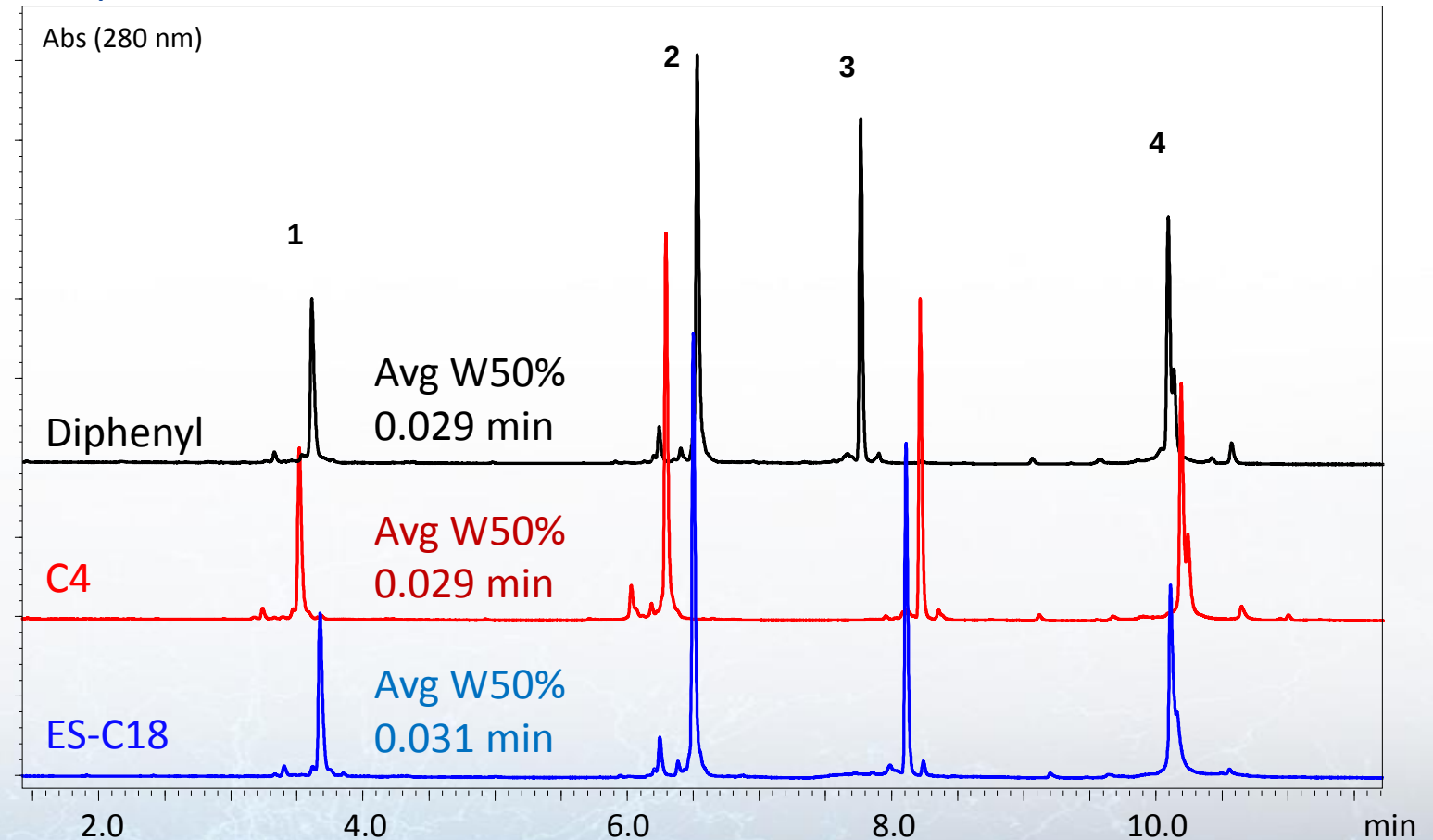
MP A: aq. 0.1% TFA; MP B: ACN + 0.1% TFA; Gradient: 20-60 %B in 15 min;
Injection Volume: 2 μ L; Flow: 0.40 mL/min; Temp: 80 $^{\circ}$ C

Effect of Bonded-Phase on Retention/Selectivity

- **Hydrophobic, Small Molecule Retention**
 - Diphenyl < C4 << ES-C18
- **Protein Retention**
 - No global retention pattern
 - Differences in individual protein retention time
- **Protein Selectivity**
 - Subtle but useful changes
- **Protein Peak Width 50%**
 - Average width nearly identical between phases

1. RNase A – 13.7 kDa
2. Lysozyme – 14.3 kDa
3. α -Lactalbumin – 14.2 kDa
4. Enolase – 46.6 kDa

Separation of 4 Protein Mixture on HALO 1000 Å Bonded-Phases



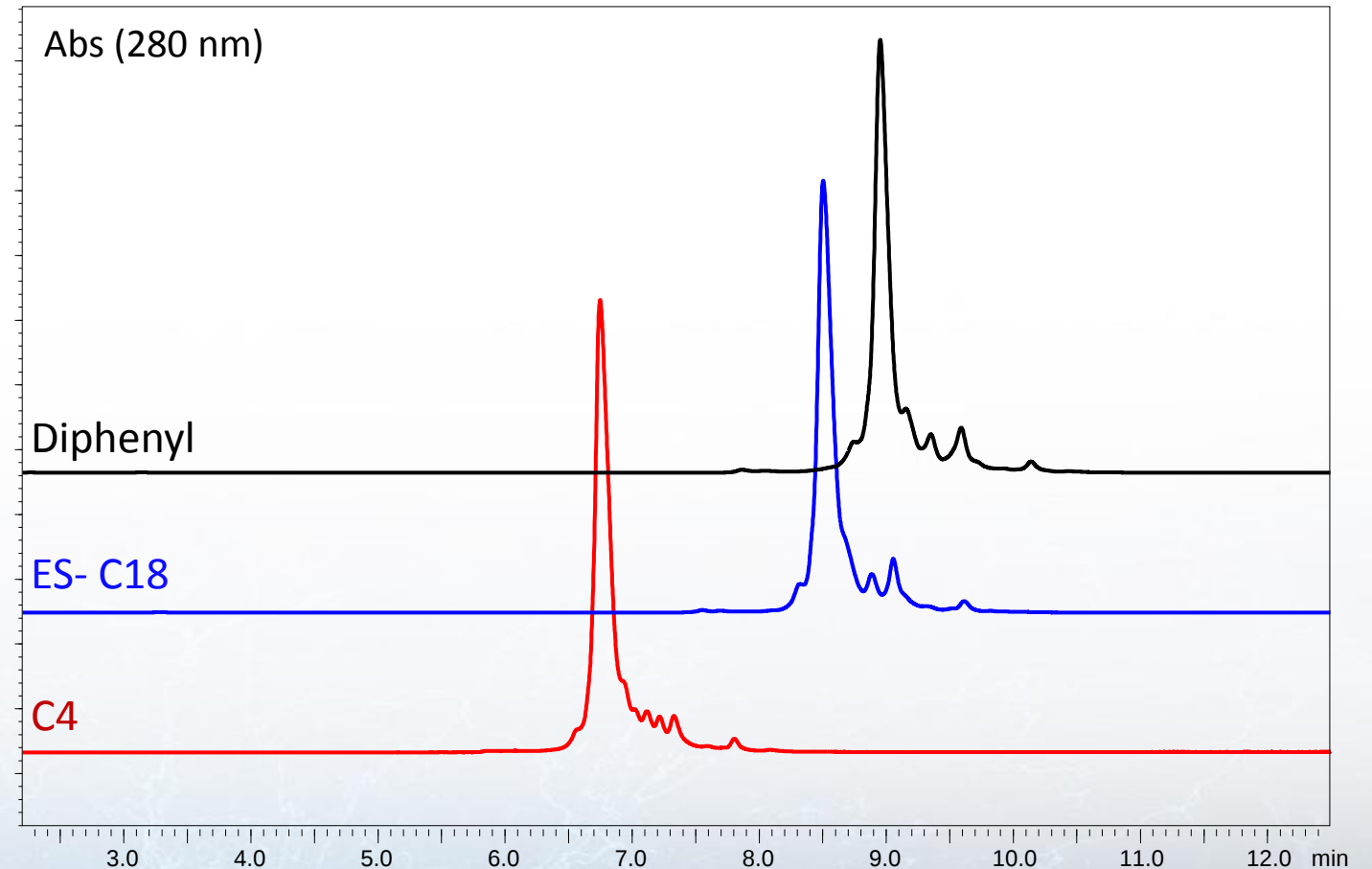
MP A: aq. 0.1% TFA; MP B: ACN + 0.1% TFA; Gradient: 20-60 %B in 15 min;
Injection Volume: 2 μ L; Flow: 0.40 mL/min; Temp: 80 $^{\circ}$ C

Effect of Bonded-Phase on Retention/Selectivity

- mAb Retention

- C4 < ES-C18 < Diphenyl
- Increased retention for ~150 kDa M.W. mAb with Diphenyl bonded-phase

Separation of Trastuzumab IgG1 on HALO 1000 Å Bonded-Phases

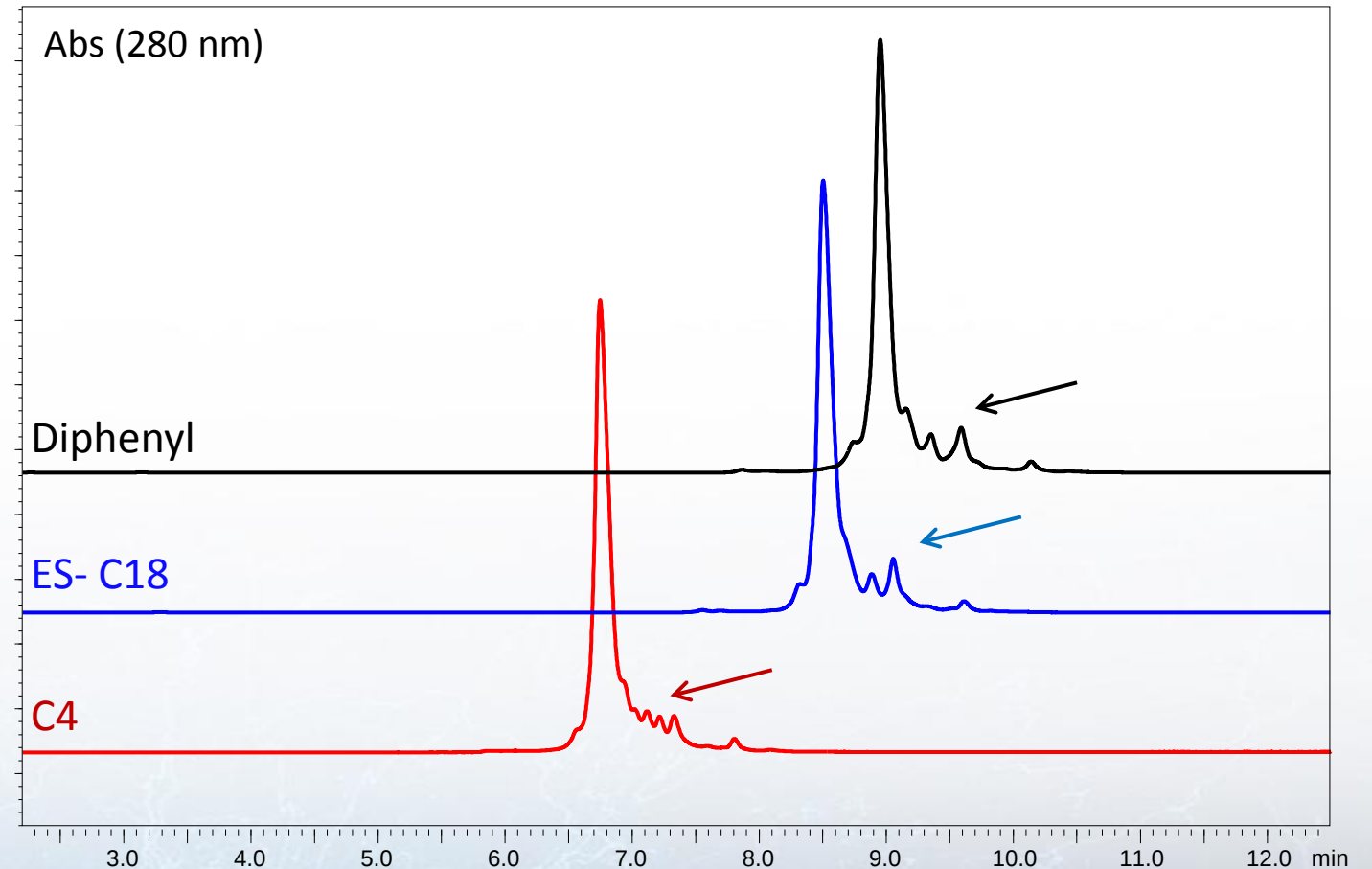


MP A: aq. 0.1% TFA; MP B: ACN + 0.1% TFA; Gradient: 32-40 %B in 16 min;
Flow 0.40mL/min; Inj Vol 2 µL; Temp: 80°C

Effect of Bonded-Phase on Retention/Selectivity

- **mAb Retention**
 - C4 < ES-C18 < Diphenyl
 - Increased retention for ~150 kDa M.W. mAb with Diphenyl bonded-phase
- **mAb Selectivity**
 - Differences observed with later eluting variant peaks

Separation of Trastuzumab IgG1 on HALO 1000 Å Bonded-Phases



MP A: aq. 0.1% TFA; MP B: ACN + 0.1% TFA; Gradient: 32-40 %B in 16 min;
Flow 0.40mL/min; Inj Vol 2 µL; Temp: 80°C

Effect of Bonded-Phase on Retention/Selectivity

- **mAb Retention**

- C4 < ES-C18 < Diphenyl
- Increased retention for ~150 kDa M.W. mAb with Diphenyl bonded-phase

- **mAb Selectivity**

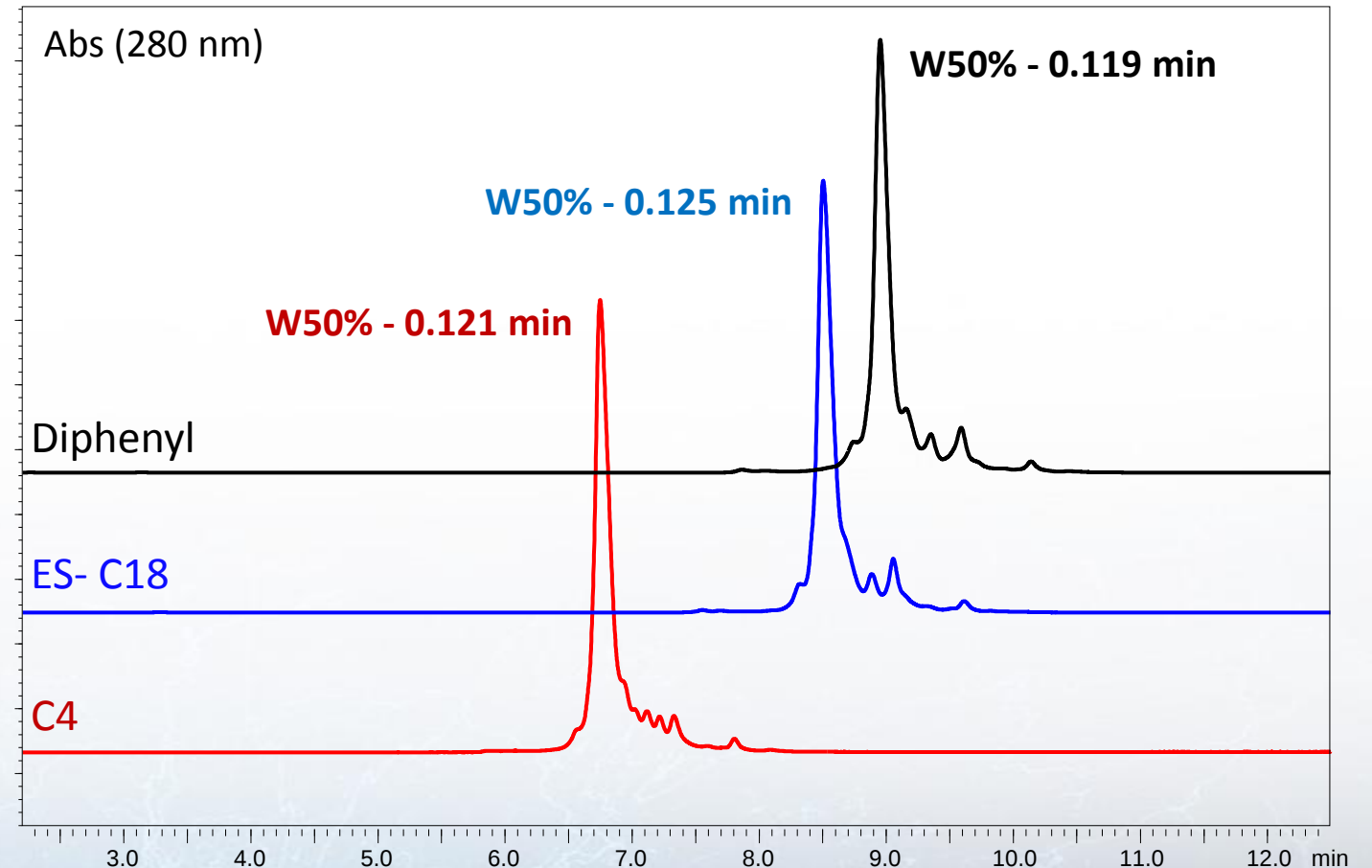
- Differences observed with later eluting variant peaks

- **mAb Peak Width 50%**

- Nearly identical for each phase

Changes in bonded-phase chemistry preserve efficiency of the separation, while providing useful changes in retention and selectivity

Separation of Trastuzumab IgG1 on HALO 1000 Å Bonded-Phases



MP A: aq. 0.1% TFA; MP B: ACN + 0.1% TFA; Gradient: 32-40 %B in 16 min;
Flow 0.40mL/min; Inj Vol 2 µL; Temp: 80°C

Selectivity Manipulation in RPLC of mAbs

1. Bonded-Phase

- C4, ES-C18, and Diphenyl offer changes in retention and selectivity

Easy to change between columns

2. Gradient Elution Variables

- Temperature
- Mobile Phase Strong Solvent
- Ion Pairing Reagent
- Gradient Slope and Time
- Flow Rate

More in-depth method development

Gradient Elution Variables: IgG1 Example

Bonded-Phase

- Diphenyl

Temperature

- Varied

Mobile Phase Strong Solvent

- ACN

Ion Pairing Reagent

- 0.1% TFA

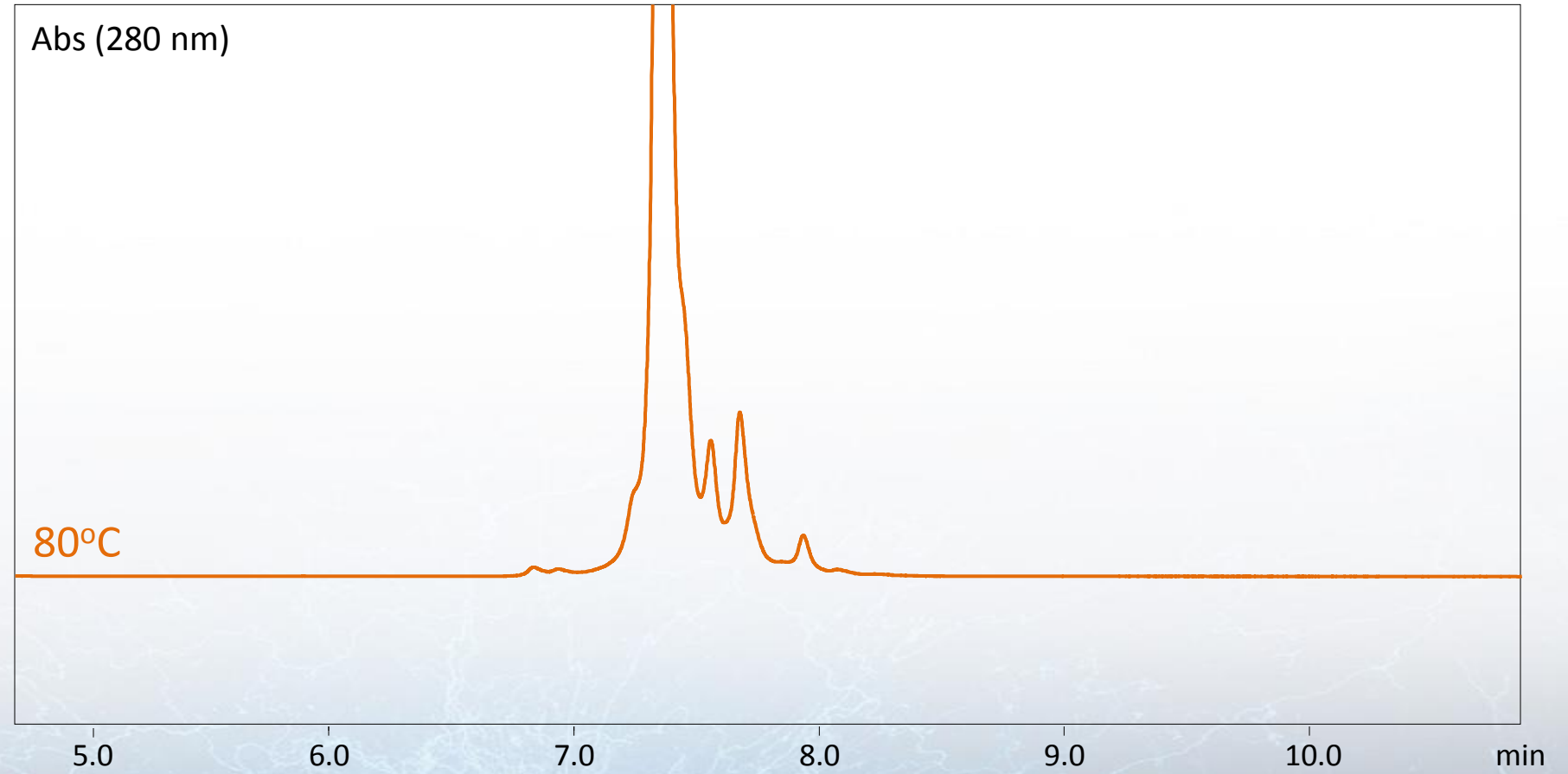
Gradient

- 30-45% B in 15 min

Flow

- 0.40 mL/min

Sample = trastuzumab IgG1



Separation on HALO 1000 Å Diphenyl, 2.7µm, 2.1x150mm
MP A: aq. 0.1% TFA; MP B: ACN + 0.1% TFA; Inj Vol 2 µL (4µg)

Gradient Elution Variables: IgG1 Example

Bonded-Phase

- Diphenyl

Temperature

- Varied

Mobile Phase Strong Solvent

- ACN

Ion Pairing Reagent

- 0.1% TFA

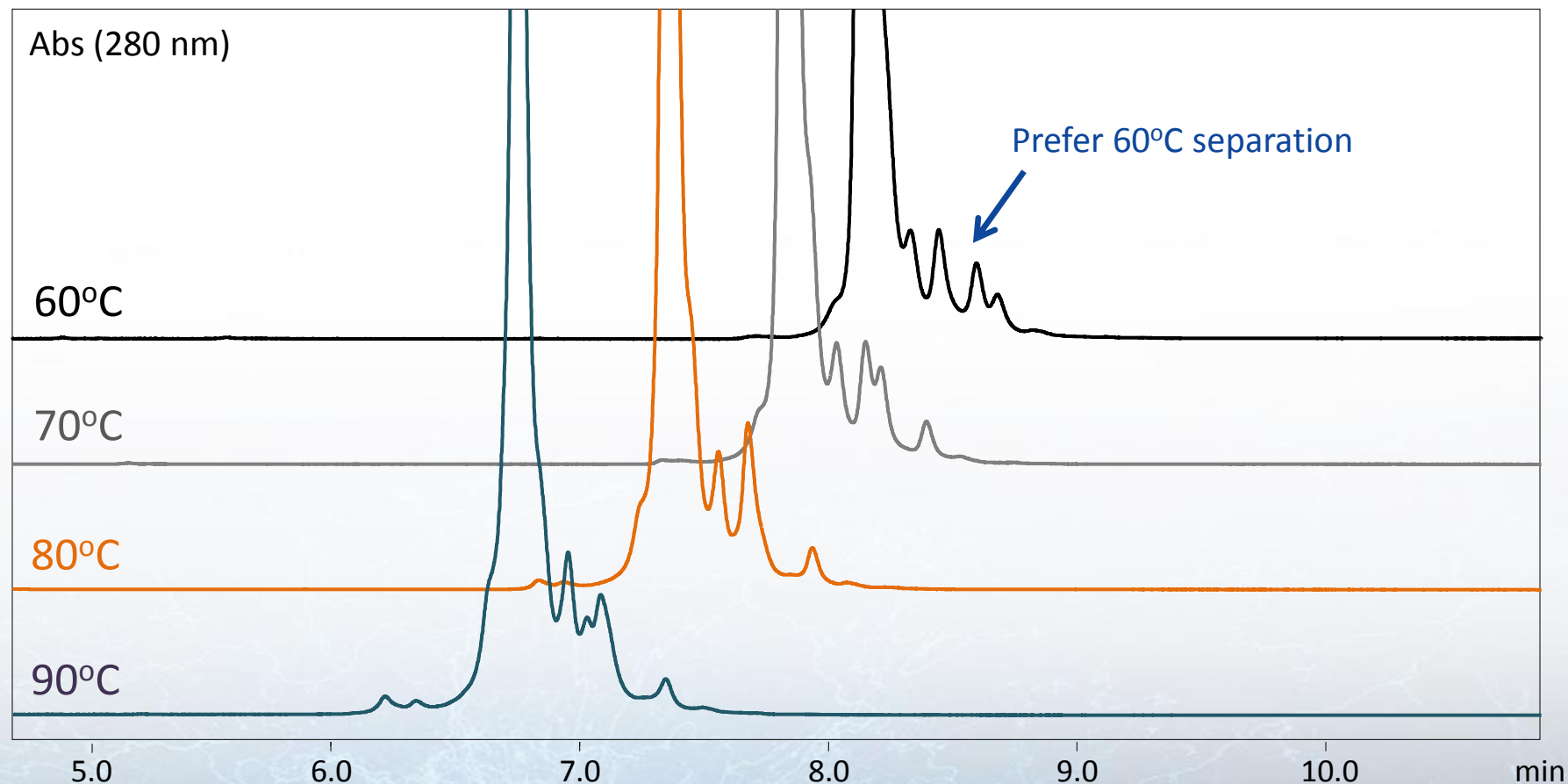
Gradient

- 30-45% B in 15 min

Flow

- 0.40 mL/min

Sample = trastuzumab IgG1



Separation on HALO 1000 Å Diphenyl, 2.7µm, 2.1x150mm
MP A: aq. 0.1% TFA; MP B: ACN + 0.1% TFA; Inj Vol 2 µL (4µg)

Gradient Elution Variables: IgG2 Example

Bonded-Phase

- C4

Temperature

- 80°C

Mobile Phase Strong Solvent

- ACN

Ion Pairing Reagent

- 0.1% TFA

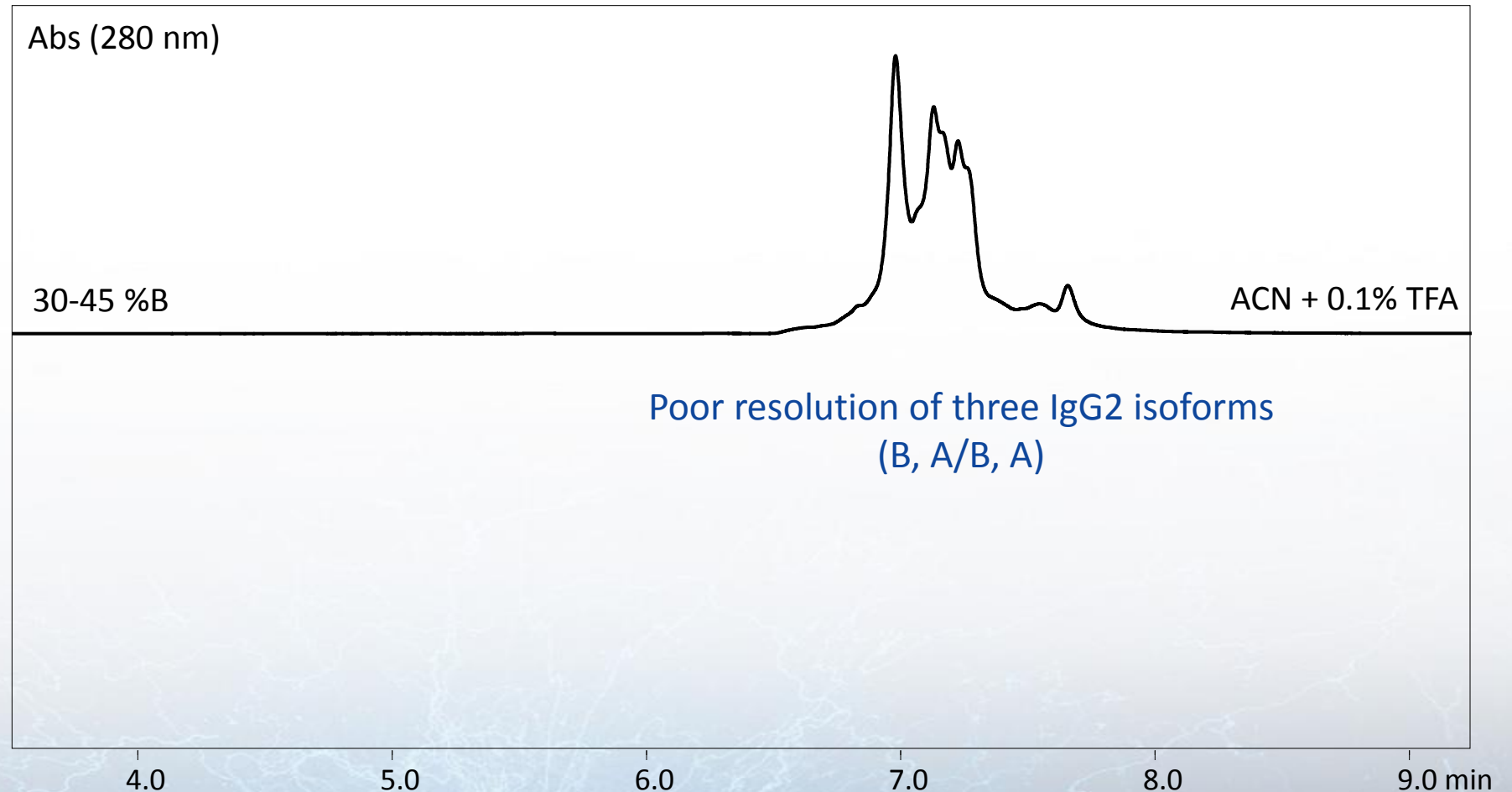
Gradient

- 15% B in 15 min

Flow

- 0.40 mL/min

Sample = denosumab IgG2



Separation on HALO 1000 Å C4, 2.7µm, 2.1x150mm

MP A: aq. 0.1% *specified acid*; MP B: *specified* + 0.1% acid; Gradient: 30-45%B for ACN/TFA,

Temp: 80°C; Inj Vol 2 µL (4µg)

Gradient Elution Variables: IgG2 Example

Bonded-Phase

- C4

Temperature

- 80°C

Mobile Phase Strong Solvent

- ACN
- 50/50 ACN/nProp

Ion Pairing Reagent

- 0.1% TFA

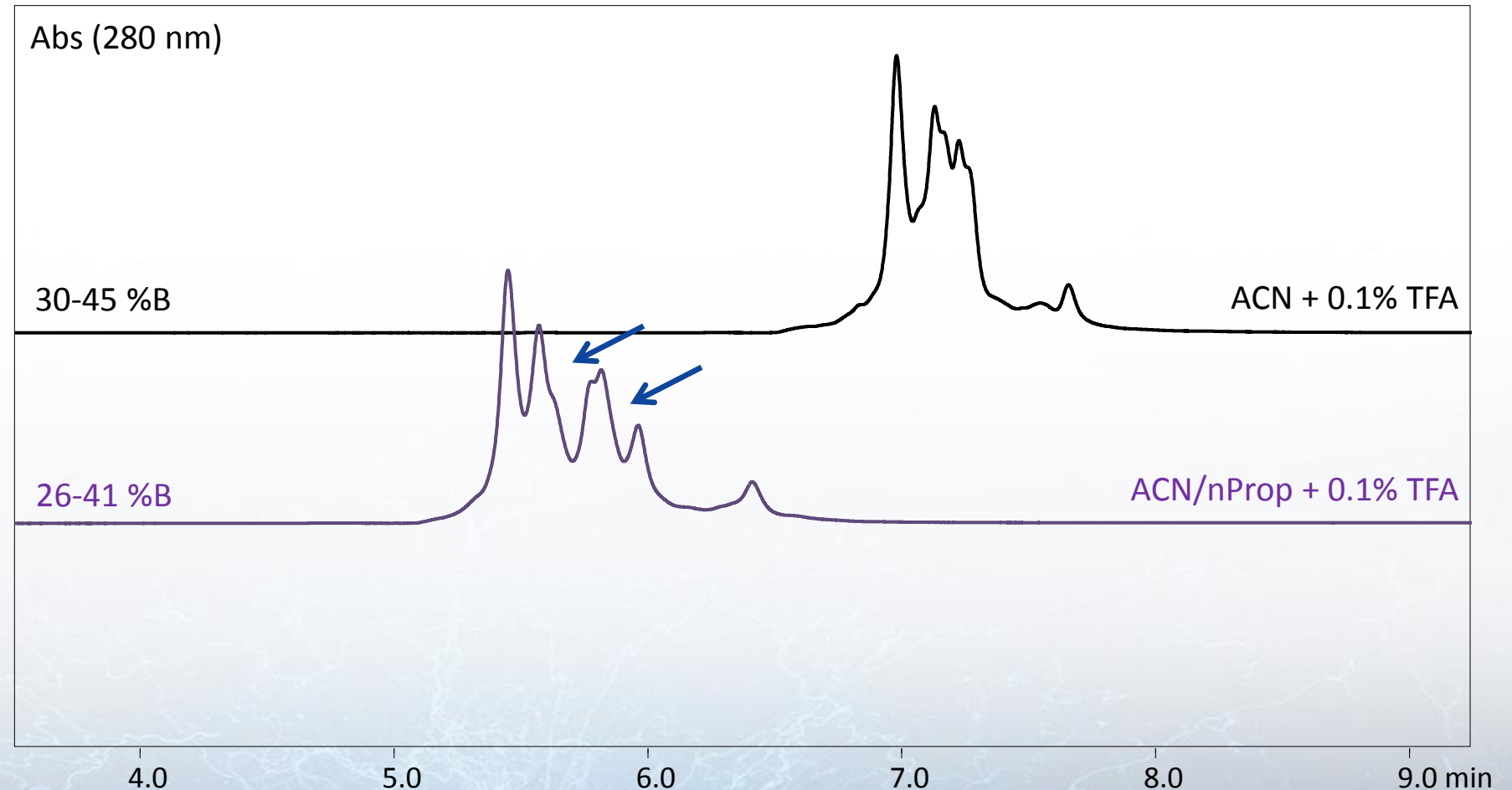
Gradient

- 15% B in 15 min

Flow

- 0.40 mL/min

Sample = denosumab IgG2



Separation on HALO 1000 Å C4, 2.7µm, 2.1x150mm

MP A: aq. 0.1% *specified acid*; MP B: *specified* + 0.1% *acid*; Gradient: 30-45%B for ACN/TFA, 26-41%B for (ACN/nProp/TFA); Temp: 80°C; Inj Vol 2 µL (4µg)

Gradient Elution Variables: IgG2 Example

Bonded-Phase

- C4

Temperature

- 80°C

Mobile Phase Strong Solvent

- ACN
- 50/50 ACN/nProp

Ion Pairing Reagent

- 0.1% TFA
- 0.1% DFA

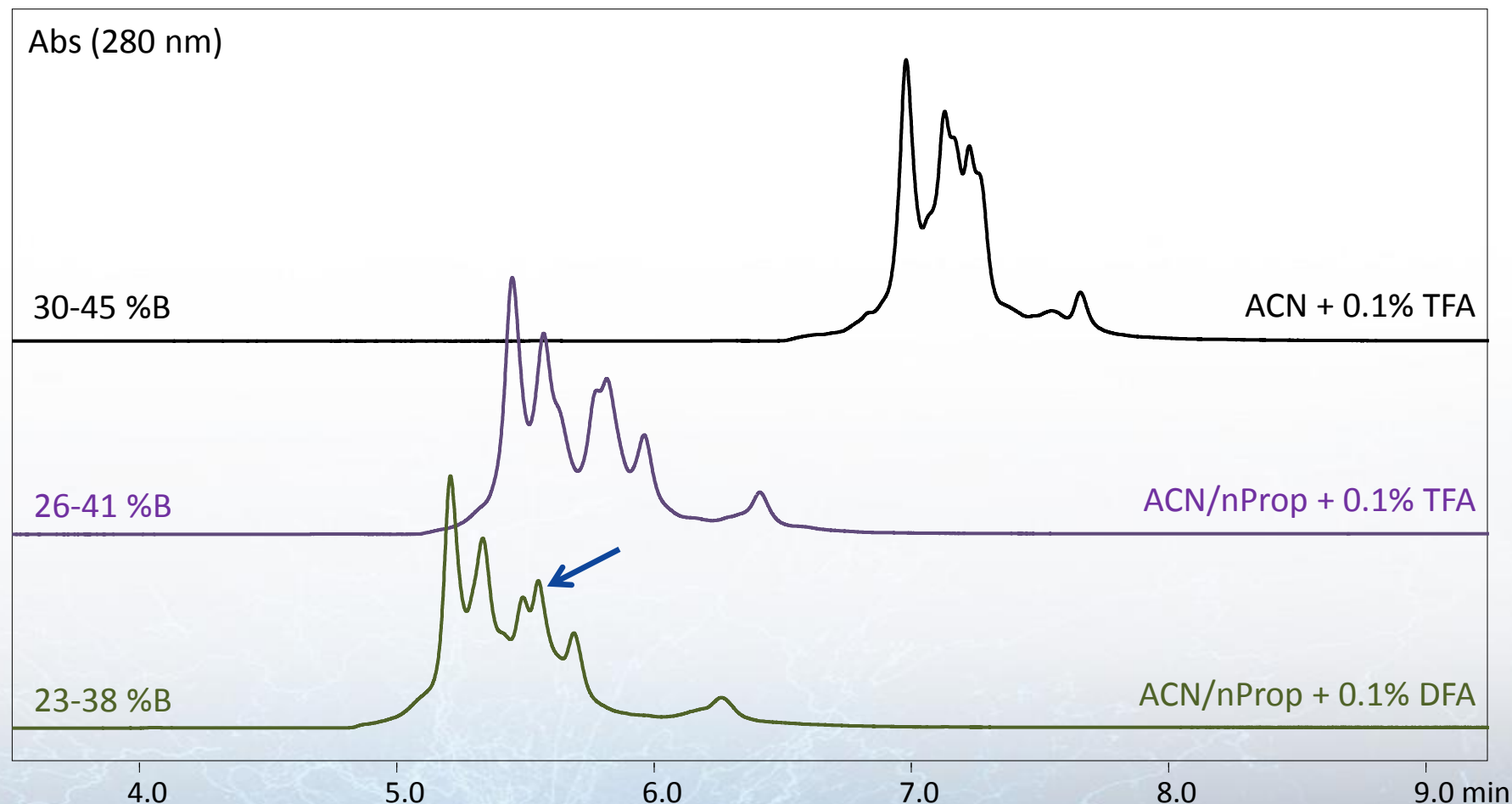
Gradient

- 15% B in 15 min

Flow

- 0.40 mL/min

Sample = denosumab IgG2



Separation on HALO 1000 Å C4, 2.7µm, 2.1x150mm

MP A: aq. 0.1% *specified acid*; MP B: *specified* + 0.1% *acid*; Gradient: 30-45%B for ACN/TFA, 26-41%B for (ACN/nProp/TFA); 23-38%B for (ACN/nProp/DFA); Temp: 80°C; Inj Vol 2 µL (4µg)

Summary and Future Work

- Improving protein separations is combination of both particle morphology and bonded-phase chemistry
- Wide pore HALO 1000Å columns demonstrate superior biomolecule separations
- Subtle, but useful, differences in selectivity are available through changes in bonded-phase
- Gradient elution variables can alter selectivity and resolution of complex mAbs
- Work continues on optimizing pore size and geometry for silica SPP
- The resolution gained by these new technologies provides a greater level of detail in the structural differences of well characterized biotechnology products

Acknowledgements

- The Research and Development team at Advanced Materials Technology including, Justin Godinho and Bob Moran (AMT) are thanked for advice, technical assistance, and samples.
- AMT management including Joe DeStefano and Tim Langlois for ongoing support on protein analytical science.
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Thank you for your Attention